

COMPARATIVE ANALYSIS OF D1 PROTEIN SEQUENCES IN CYANOBACTERIA

CIPRIAN CHIȘ^{1,2}, IULIANA CHIȘ^{1,2}, OANA SICORA¹,
DALTON CARMEL^{1,3}, AURICA GREC³, AUREL ARDELEAN³,
NICOLAE DRAGOȘ² and COSMIN SICORA^{1,✉}

SUMMARY. The D1 protein of Photosystem II (PSII), encoded by the *psbA* genes, is an indispensable component of oxygenic photosynthesis. Due to strongly oxidative chemistry of PSII water splitting, the D1 protein is prone to constant damage and requires its replacement by a new copy every 5 h under low light conditions and every 20 minutes under intense illumination, whereas most of the other PSII subunits remain ordinarily undamaged. In cyanobacteria the D1 protein is encoded by a *psbA* gene family, ranging from 1 to 6 members. The presence of multiple *psbA* genes encoding different D1 isoforms is an indication of their importance in regulatory mechanisms responsible for maintaining a functional PSII upon changing environmental conditions in natural habitats of cyanobacteria.

Herewith, we present a comparative analysis of the protein sequences encoded by *psbA* gene family in model cyanobacteria strains with sequenced genomes, highlighting their characteristic features that give indication of their putative function.

Keywords: cyanobacteria, D1 protein, Photosystem II, photosynthesis;

Introduction

Photosynthesis is one of the most important processes responsible for the maintenance of the oxygen level in the atmosphere and the reduction of CO₂ to carbohydrates (Loll et. al., 2005). It creates organic matter out of inorganic compounds. In plants, as well as in algae and cyanobacteria, photosynthesis is initiated in Photosystem II (PSII) using light energy to drive two distinct chemical reactions - the photo-oxidation of water and the reduction of plastoquinone (Singhal et. al., 1999).

¹ Biological Research Center, Jibou, Romania

² Babeș-Bolyai University, Cluj-Napoca, Romania

³ Western University "Vasile Goldis, Arad, Romania

✉ **Corresponding author: Cosmin Sicora**, Biological Research Center Jibou, Romania.

E-mail: cosmin.sicora@gmail.com

PSII is a large homo-dimeric protein-cofactor complex consisting of 20 protein subunits and more than 77 co-factors. Two proteins form the reaction center core of PSII: the D1 and D2 proteins. While higher plants have one *psbA* gene encoding the D1 protein, cyanobacteria generally display a small *psbA* gene family, consisting of 1 to 6 members, differentially expressed, according to environmental conditions (Mulo *et al.*, 2009). The D1 protein of PSII is an indispensable component of oxygenic photosynthesis. The aminoacid structure of the *PsbA* peptide gives clues about the hydrophobic nature for D1 protein with five membrane spanning helices (Dwivedi and Bhardwaj, 1995). The D1 protein is photodamaged because of the strongly oxidative chemistry of the Photosystem II, requiring its replacement, whereas most of the other subunits of PSII remain undamaged. The D1 protein is the primary component damaged and replaced in all organisms that perform oxygenic photosynthesis. In normal growth conditions a D1 protein is replaced in the PSII every 5 h, and under different stress conditions the replacement occurs every 20 minutes (Mulo *et al.*, 2009). When the D1 protein is damaged, the Photosystem II is inactivated; the dimers separate, followed by the partial disassembly of the Photosystem II. The damaged D1 protein is removed by an FtsH type protease and degraded. Subsequently a new copy of D1 protein is translated, and the new D1 protein is co-translational inserted into the membrane. The C-terminus of the D1 protein is post-translational processed. The monomers are assembled, then the functional dimers are formed and the PSII is active again (Mulo *et al.*, 2009).

Due to the advance of sequencing techniques, new genomes of cyanobacteria are being sequenced and annotated all the time (<http://genome.microbedb.jp/cyanobase/>), so periodically new *psbA* gene sequences and of the proteins they encode become available. These sequences allow for putative functional identification of the D1 isoforms and offer clues about their role. The studies of gene expression showed that under stress conditions, cyanobacterial cells adopt different strategies to cope with the changes. One strategy involves the production of larger amount of the same D1 protein, compensating for the higher rate of damage under stress, behavior best characterized in *Synechocystis* sp. PCC 6803, where one type of D1 protein is encoded by two *psbA* genes (Mate *et al.*, 1998). Another strategy involves the replacement of a D1 protein isoform present in normal growth conditions with another one under stress conditions, this strategy was first described at *Synechococcus* sp. PCC 7942 (Sane *et al.*, 2002). The D1 protein isoforms exchange occurs helping the cell to resist better in stress conditions and increases the adaptability to different environmental conditions (Clarke *et al.*, 1993). A parallel strategy involves the induction of another *psbA* gene, considered silent for a long time (Sicora *et al.*, 2009, Summerfield *et al.*, 2008). It was shown that during the microaerobic conditions, a distinct D1 protein isoform is induced, the D1' isoform, hence its postulated role in adaptation of the cells to low

oxygen conditions (Sicora *et al.*, 2009; Summerfield *et al.*, 2008). Recently, a new class of D1 protein termed 'D1 rogue' (D1r), present in diazotrophic cyanobacteria, was postulated by Murray (2012). Based on its structural characteristics, the protein D1 rogue impairs the PSII function during night allowing for nitrogen fixation.

In conclusion, five, functionally different, D1 protein isoforms are known to date (Mulo *et al.*, 2009):

D1m or D1major is an isoform present in PSII in normal growth conditions, and induced in most stress conditions.

D1:1 is expressed in PSII under normal growth conditions, repressed under stress conditions.

D1:2 are repressed in PSII under normal growth conditions, induced under stress conditions. The D1:1 and D1:2 proteins differ at 25 of 360 amino-acid positions, with a conserved change of Gln with Glu at position 130 (Campbell *et al.*, 1998).

D1' protein is induced in microaerobic conditions. The sequence of amino-acid shows three conserved mutations at positions 80, 158 and 286; these changes are characteristic to the D1' isoform: position 80 is a Gly to Ala mutation, at position 158 is a Phe to Leu mutation and at position 286 is a Thr to Ala mutation (Sicora *et al.*, 2009).

D1r protein is a hypothetical protein, which blocks the Photosystem II during night and has 8 characteristic amino-acids differences; the most relevant from them are at positions: 61 Asp to Glu, 170 Asp to Glu/Ser, 189 Glu to Asp/Ala/Arg, 333 Glu to Ala/Ser, 342 Asp to Thr/Leu/Val, 344 Ala to Ala/Ser (Murray, 2012).

While functional characterization is needed for proper D1 isoform identification, in this article we identify possible D1 protein isoforms inferred from the existing sequencing data available fitting them in the proposed functional classes. We also propose models for the gene family development in the studied cyanobacteria strains.

Materials and methods

All the cyanobacterial *psbA* gene sequences were obtained from CyanoBase (<http://genome.microbedb.jp/cyanobase/>) and imported in the CLC Sequence Viewer 6.9.1 software (freeware <http://www.clcbio.com>). Using the aminoacid sequences, we made an accurate multiple alignment of all 91 sequences, based on progressive alignment algorithm. The alignment was manual edited, the non-informative regions were deleted resulting a number of approximative 360 positions. This made possible the comparison of the aminoacids conserved at all D1 protein isoforms. Based on

the alignment, we summarized the amino-acid changes in tables 2, 3 and 4. The next step was the construction of phylogenetic trees by Neighbour Joining method, 1000 replicates, made with CLC Sequence Viewer 6.9.1 program. A Venn diagram, showing the grouping and distribution of the studied D1 protein isoforms was constructed.

Results and discussion

In this study we analyzed a number of 30 cyanobacterial strains with sequenced genomes, from Cyanobase and identified a total of 91 *psbA* genes which encode different isoforms of the D1 protein (Table 1). There are cases when several genes encode one D1 protein isoform and other cases when each gene encodes a different D1 protein.

As the functional data accumulated it had become evident that all D1 protein isoforms have specific changes in the peptide structure. Based on this experimental observations it has been concluded that the presence at position in the position 130 of a glutamic acid (Glu), instead of glutamine (Gln) suggests the presence of the D1:2 isoform of the D1 protein strongly induced under stress conditions (Table 2).

Usually, one cyanobacterial strain possessing the D1:2 isoform also has the D1:1 isoform of the D1 protein expressed under optimal growth conditions, so we can see that from the total of 30 cyanobacterial strains studied, 17 strains have 45 *psbA* genes which encode the D1:2 isoform of the D1 protein.

Another specific isoform of the D1 protein (D1') is induced in microaerobic conditions (Sicora *et al.*, 2009). Based on the alignment, we searched the sequenced with the three changes in the aminoacid structure characteristic cu D1' isoform (Table 3). This table, shows that out of thirty cyanobacterial strains studied, ten have the D1' isoform of the D1 protein, the isoform that is induced in microaerobic conditions and probably helps the cells adapt to low oxygen conditions.

By bioinformatic analysis of D1 aminoacid composition Murray (2012) concluded that a possible new isoform of the D1 protein that putatively blocks the Photosystem II during night and allows for nitrogen fixation. This isoform of the D1 protein ('D1 rogue') has 6 major changes in the aminoacids structure. The presence at the position 61 of glutamic acid (Glu) instead of aspartic acid (Asp), at position 130 of a glutamic acid (Glu) or serine (Ser) instead of an aspartic acid (Asp), at position 189 of an aspartic acid (Asp) or alanine (Ala) or arginine (Arg) instead of a glutamic acid aminoacid, are only 3 changes that suggest the presence of the D1 rogue isoform. From what we can see below (Table 4), five out of thirty cyanobacterial strains analyzed have three, four or five changes in the peptide structure characteristic to the D1 rogue protein.

Table 1.Cyanobacterial strains analyzed in this study and the proposed number of *psbA* genes within.

No.	Cyanobacterial strain	Number of <i>psbA</i> genes
1	<i>Synechocystis</i> sp. PCC 6803	3
2	<i>Anabaena</i> sp. PCC 7120	5
3	<i>Thermosynechococcus elongatus</i> BP-1	3
4	<i>Gloeobacter violaceus</i> PCC 7421	5
5	<i>Mycrocystis aeruginosa</i> NIES-843	5
6	<i>Prochlorococcus marinus</i> SS120	1
7	<i>Prochlorococcus marinus</i> MED4	1
8	<i>Prochlorococcus marinus</i> MIT9313	2
9	<i>Synechococcus</i> sp. WH8102	4
10	<i>Synechococcus elongatus</i> PCC 6301	3
11	<i>Synechococcus</i> sp. CC9311	2
12	<i>Synechococcus</i> sp. PCC 7002	3
13	<i>Acaryochloris marina</i> MBIC11017	3
14	<i>Prochlorococcus marinus</i> str. NATL2A	3
15	<i>Anabaena variabilis</i> ATCC 29413	6
16	<i>Synechococcus</i> sp. CC 9902	4
17	<i>Synechococcus elongatus</i> PCC 7942	3
18	<i>Synechococcus</i> sp. JA-2-3B ^a (2-13)	3
19	<i>Synechococcus</i> sp. JA-3-3Ab	4
20	<i>Prochlorococcus marinus</i> str. AS9601	1
21	<i>Prochlorococcus marinus</i> str. MIT9515	1
22	<i>Prochlorococcus marinus</i> str. MIT9303	1
23	<i>Prochlorococcus marinus</i> str. NATL21A	2
24	<i>Prochlorococcus marinus</i> str. MIT 9301	1
25	<i>Synechococcus</i> sp. RCC307	4
26	<i>Synechococcus</i> sp. WH7803	4
27	<i>Prochlorococcus marinus</i> str. MIT9215	1
28	<i>Cyanothece</i> sp. ATCC 51142	5
29	<i>Nostoc punctiforme</i> ATCC 29133	4
30	<i>Arthrospira platensis</i> NIES-39	4
TOTAL	30	91

Table 2.

List of proteins (emphasized by bold characters below strain's name) from studied cyanobacterial strains exhibiting aminoacid changes characteristic to D1:1 and D1:2 isoforms.

D1:2		
130 Gln to Glu		
<i>Synechococcus</i> sp. PCC 7002 SynPCC7002_A1418	<i>Synechococcus</i> sp. PCC 7002 SynPCC7002_A0157	<i>Anabaena</i> sp. PCC 7120 Alr_4592
<i>Synechococcus elongatus</i> PCC 7942 Synpcc7942_1389	<i>Synechococcus elongatus</i> PCC 7942 Synpcc7942_0893	<i>Anabaena</i> sp. PCC 7120 Alr_3727
<i>Synechococcus elongatus</i> PCC 6301 Syc0166_d	<i>Synechococcus elongatus</i> PCC 6301 Syc0647_d	<i>Anabaena</i> sp. PCC 7120 All_3572
<i>Anabaena variabilis</i> ATCC 29413 Ava_1597	<i>Anabaena variabilis</i> ATCC 29413 Ava_2460	<i>Anabaena variabilis</i> ATCC 29413 Ava_3553
<i>Nostoc punctiforme</i> ATCC 29133 Npun_F3544	<i>Nostoc punctiforme</i> ATCC 29133 Npun_R5188	<i>Nostoc punctiforme</i> ATCC 29133 Npun_R2273
<i>Synechococcus</i> sp. WH8102 SYNW0983	<i>Synechococcus</i> sp. WH8102 SYNW1919	<i>Synechococcus</i> sp. WH8102 SYNW2151
<i>Synechococcus</i> sp. CC9902 Syncc9902_1814	<i>Synechococcus</i> sp. CC9902 Syncc9902_2036	<i>Synechococcus</i> sp. CC9902 Syncc9902_1817
<i>Synechococcus</i> sp. CC9311 Sync_2384	<i>Synechococcus</i> sp. WH 7803 SynWH7803_0366	<i>Synechococcus</i> sp. WH 7803 SynWH7803_0790
<i>Synechococcus</i> sp. WH 7803 SynWH7803_2084	<i>Synechococcus</i> sp. RCC307 SynRCC307_1440	<i>Synechococcus</i> sp. RCC307 SynRCC307_2009
<i>Synechococcus</i> sp. RCC307 SynRCC307_2183	<i>Arthrospira platensis</i> NIES-39 NIES39_K03030	<i>Arthrospira platensis</i> NIES-39 NIES39_03080
<i>Arthrospira platensis</i> NIES-39 NIES39_R00140	<i>Thermosynechococcus elongatus</i> BP-1 Tlr1477	<i>Thermosynechococcus elongatus</i> BP-1 Tlr1844
<i>Synechococcus</i> sp. JA-2-3B'a(2-13) CYB_0371	<i>Synechococcus</i> sp. JA-2-3B'a(2-13) CYB_0433	<i>Synechococcus</i> sp. JA-3-3Ab CYA_1811
<i>Synechococcus</i> sp. JA-3-3Ab CYA_1849	<i>Synechococcus</i> sp. PCC 7002 SynPCC7002_A2164	<i>Arthrospira platensis</i> NIES-39 NIES39_O04720

COMPARATIVE ANALYSIS OF D1 PROTEIN SEQUENCES IN CYANOBACTERIA

Table 2. continued

<i>Gloeobacter violaceus</i> PCC 7421 Glr0779	<i>Gloeobacter violaceus</i> PCC 7421 Glr2322	<i>Gloeobacter violaceus</i> PCC 7421 Glr2656
<i>Gloeobacter violaceus</i> PCC 7421 Glr1706	<i>Gloeobacter violaceus</i> PCC 7421 Glr3144	<i>Cyanothece</i> sp. ATCC 51142 Cce_3411

Table 3.

List of proteins from studied cyanobacterial strains that exhibit aminoacid changes characteristic for the D1' isoform.

D1'		
80 Gly to Ala	158 Phe to Leu	286 Thr to Ala
<i>Thermosynechococcus elongatus</i> BP-1 Tlr1844	<i>Thermosynechococcus elongatus</i> BP-1 Tlr1844	<i>Thermosynechococcus elongatus</i> BP-1 Tlr1844
<i>Synechococcus</i> sp. PCC 7002 SynPCC7002_A2164	<i>Synechococcus</i> sp. PCC 7002 SynPCC7002_A2164	<i>Synechococcus</i> sp. PCC 7002 SynPCC7002_A2164
<i>Arthrospira platensis</i> NIES-39 NIES39_O04720	<i>Arthrospira platensis</i> NIES-39 NIES39_O04720	<i>Arthrospira platensis</i> NIES-39 NIES39_O04720
<i>Synechocystis</i> sp. PCC 6803 Slr1181	<i>Anabaena</i> sp. PCC 7120 Alr3742	<i>Synechocystis</i> sp. PCC 6803 Slr1181
<i>Anabaena</i> sp. PCC 7120 Alr3742	<i>Anabaena variabilis</i> ATCC 29413 Ava_1583	<i>Anabaena</i> sp. PCC 7120 Alr3742
<i>Anabaena variabilis</i> ATCC 29413 Ava_1583	<i>Cyanothece</i> sp. ATCC 51142 Cce_3411	<i>Anabaena variabilis</i> ATCC 29413 Ava_1583
<i>Cyanothece</i> sp. ATCC 51142 Cce_3411	<i>Synechococcus</i> sp. JA-3-3Ab CYA_1748	<i>Cyanothece</i> sp. ATCC 51142 Cce_3411
<i>Synechococcus</i> sp. JA-3-3Ab CYA_1748	<i>Synechococcus</i> sp. JA-2-3B'a(2-13) CYB_0216	<i>Synechococcus</i> sp. JA-3-3Ab CYA_1748
<i>Synechococcus</i> sp. JA-2-3B'a(2-13) CYB_0216	<i>Anabaena variabilis</i> ATCC 29413 Ava_4121	<i>Synechococcus</i> sp. JA-2-3B'a(2-13) CYB_0216
<i>Cyanothece</i> sp. ATCC 51142 Cce_3477	<i>Acaryochloris marina</i> MBIC11017 AM1_0448	<i>Cyanothece</i> sp. ATCC 51142 Cce_3477
<i>Anabaena variabilis</i> ATCC 29413 Ava_4121		<i>Anabaena variabilis</i> ATCC 29413 Ava_4121

Table 3. continued

<i>Acaryochloris marina</i> MBIC11017 AM1_0448	<i>Acaryochloris marina</i> MBIC11017 AM1_0448
---	---

Table 4.

List of proteins from studied cyanobacterial strains that exhibit aminoacid changes characteristic to the D1rogue isoform.

D1r		
61 Asp to Glu	170 Asp to Asp/Glu/Ser	189 Glu-Asp/Ala/Arg
<i>Cyanothece</i> sp. ATCC 51142 Cce_3477	<i>Synechococcus</i> sp. JA-3-3Ab CYA_1748	<i>Cyanothece</i> sp. ATCC 51142 Cce_3477
<i>Acaryochloris marina</i> MBIC11017 AM1_0448	<i>Synechococcus</i> sp. JA-2-3B'a(2-13) CYB_0216 <i>Cyanothece</i> sp. ATCC 51142 Cce_3477 <i>Acaryochloris marina</i> MBIC11017 AM1_0448	<i>Anabaena variabilis</i> ATCC 29413 Ava_4121
D1r		
333 Glu to Ala/Ser	342 Asp to Thr/Leu/Val	344 Ala to Ser
<i>Synechococcus</i> sp. JA-3-3Ab CYA_1748	<i>Synechococcus</i> sp. JA-3-3Ab CYA_1748	<i>Anabaena variabilis</i> ATCC 29413 Ava_4121
<i>Synechococcus</i> sp. JA-2-3B'a(2-13) CYB_0216	<i>Synechococcus</i> sp. JA-2-3B'a(2-13) CYB_0216	
<i>Cyanothece</i> sp. ATCC 51142 Cce_3477	<i>Cyanothece</i> sp. ATCC 51142 Cce_3477	
<i>Anabaena variabilis</i> ATCC 29413 Ava_4121	<i>Anabaena variabilis</i> ATCC 29413 Ava_4121	
<i>Acaryochloris marina</i> MBIC11017 AM1_0448	<i>Acaryochloris marina</i> MBIC11017 AM1_0448	

Based on the analysis of the tables 2, 3 and 4, we observed the possible presence of several types of D1 protein at the same cyanobacteria strain, without a very obvious relation between them. To better highlight the number of cyanobacterial strains which contain several different isoforms of the D1 protein, we built a Venn diagram (Fig. 1). We observed that from the total of thirty cyanobacterial strains analyzed, four strains have all the three isoforms of D1 protein (D1:2, D1' and D1r).

Seventeen cyanobacterial strains contain the D1:2 isoform of D1 protein. From these seventeen genomes, nine have just the D1:2 isoform, four have both the D1:2 and D1' isoform and four of them have all the 3 isoforms discussed.

As concerns the D1' isoform, ten cyanobacteria contain this isoform of D1 protein; one strain has only the D1' isoform out of the three studied isoforms. Four strains contain the D1:2 and D1' isoform, four strains contain all the three isoforms and only one strain contains the D1' together with the D1r isoform.

The D1r isoform is present in five cyanobacterial genomes, from which four genomes have all the three isoforms and one have the D1r and D1' isoforms.

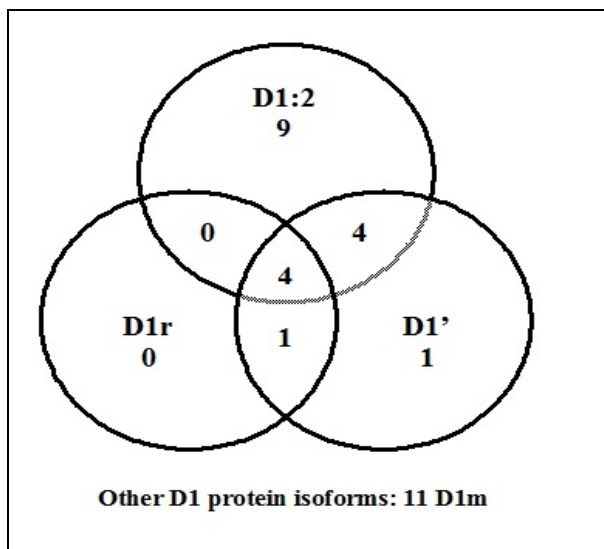


Figure 1. Venn diagram summarizing the distribution of D1 protein isoforms among the cyanobacterial strains analyzed.

Based on the alignment done with the sequences imported in the CLC sequence Viewer 6.9.1 software, a phylogenetic tree including all 91 sequences was constructed by using Neighbour Joining method, (Fig. 2).

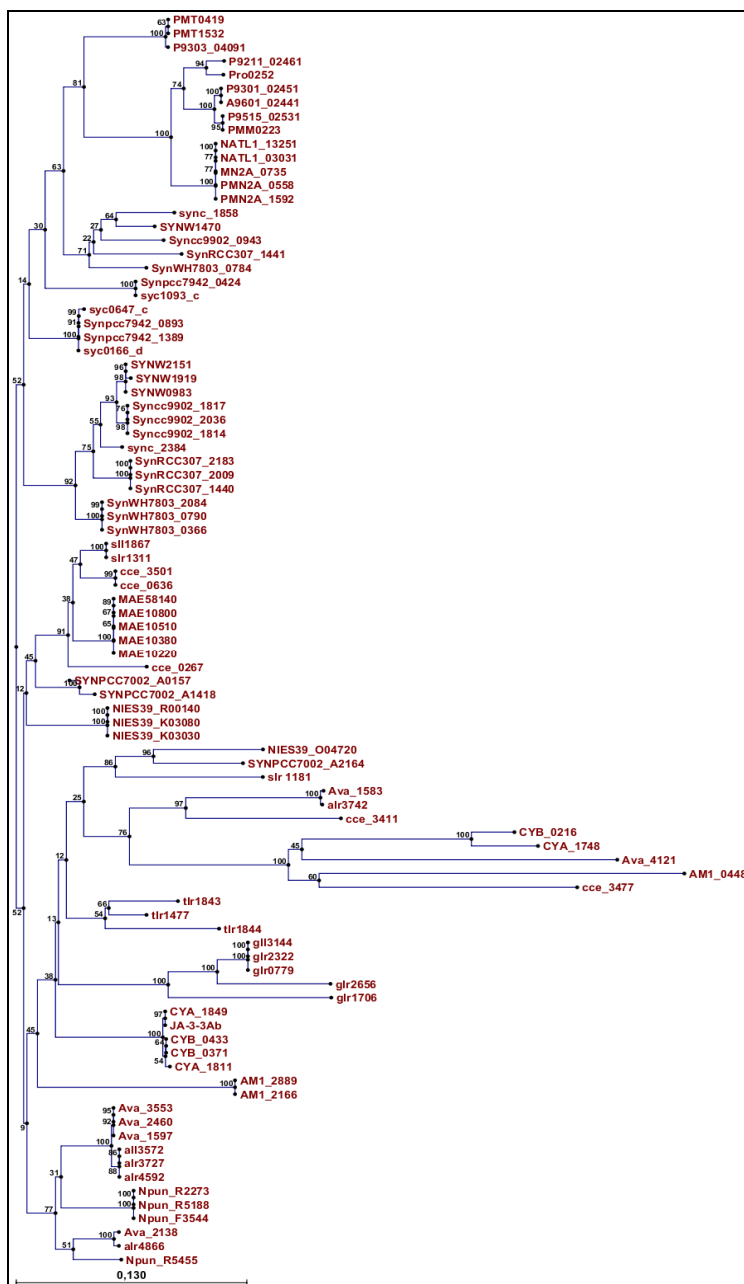


Figure 2. Phylogenetic tree of the 91 protein sequences from Cyanobase, made with Neighbour Joining method, 1000 bootstrap replicates. The scale bar represents number of insertions, deletions/time.

Analyzing the tree it becomes evident that specific isoforms are grouped together independent of the cyanobacterial strain they belong to. D1' protein from *Arthrospira platensis* NIES-39 (NIES39_O04720) is grouped with the one from *Synechococcus* sp. PCC 7002 (SYNPCC7002_A2164), *Synechocystis* sp. PCC 6803 (Slr1181), *Anabaena variabilis* ATCC 29413 (Ava_1583), *Anabaena* sp. PCC 7120 (Alr3742) and *Cyanothece* sp. ATCC 51142 (Cce_3411). There are, however, several D1 protein families grouped after the strain they belong. Examples for this case are the D1 protein family from *Gloeobacter violaceus* PCC 7421 (glr0779, gll3144, glr2322, glr2656, glr1706) or *Thermosynechococcus elongatus* BP-1 (tlr1843, tlr1477, tlr1844).

Based on our results, we can conclude that there are two possible ways in which the *psbA* genes and the D1 protein encoded by them developed.

One strategy is by horizontal transfer, when the proteins are more closely related to each other than to the other members of the gene family from the same strain, it can be observed that the D1' protein from different species that group together (Fig. 2). An explanation can be the fact that the strains sharing the same environment conditions managed to adapt by the importing a gene that will give them an evolutionary edge. The transfer can be intermediated by common phages infecting a specific community.

The second strategy is by intraspecific duplication of the genes. In this way the strains create new forms of D1 protein by duplication from the existing ones as a response to the need to adapt to changes in the environment (Sajjaphan *et al.*, 2002).

Conclusions

Herewith, we reviewed the available new information regarding the D1 protein isoforms from newly sequenced cyanobacteria genomes and we presented a possible functional distribution of these forms based upon the available knowledge. This information can trigger further functional studies leading to characterization of new D1 protein families. We also processed these data into an evolutionary tree that gives indications regarding the evolutionary strategies of the *psbA* gene family from cyanobacteria. The presented data bring, original interpretation of updated information regarding the *psbA* gene families and the encoded D1 protein isoforms available from the newly sequenced cyanobacterial genomes.

Acknowledgements. This work was supported by a grant of the Romanian National Authority for Scientific Research, CNCS-UEFISCDI, project number PN-II-ID-PCE-2011-3-0765

REFERENCES

- Campbell, D., Eriksson, M.-J., Oquist, G., Gustafsson, P., Clarke, A.K. (1998) The cyanobacterium *Synechococcus* resists UV-B by exchanging photosystem II reaction-center D1 proteins, *Proc. Natl. Acad. Sci. USA* **95**, 364-369
- Clarke, A.K., Hurry, V.M., Gustafsson, P., Oquist, G. (1993) Two functionally distinct forms of the photosystem II reaction-center protein D1 in the cyanobacterium *Synechococcus* sp. PCC 7942, *Proc. Natl. Acad. Sci. USA* **90**, 11985-11989
- Dwivedi, U., Bhardwaj, R. (1995) D1 protein of photosystem II: The light sensor in chloroplasts, *J. Biosci.* **20**, 35-47
- Loll, B., Kern, J., Saenger, W., Zouni, A., Biesiadka, J. (2005) Towards complete cofactor arrangement in the 3.0 Å resolution structure of photosystem II, *Nature*, **438**, 1040-1044
- Mate, Z., Sass, L., Szekeres, M., Vass, I., Nagy, F. (1998) UV-B induced differential transcription of *psbA* genes encoding the D1 protein of photosystem II in the cyanobacterium *Synechocystis* 6803, *J. Biol. Chem.* **273** 17439-17444
- Mulo, P., Sicora, C., Aro, E.-M. (2009) Cyanobacterial *psbA* gene family: optimization of oxygenic photosynthesis, *Cell. Mol. Life. Sci* **66**: 3697-3710
- Murray, J.W. (2012) Sequence variation at the oxygen-evolving centre of photosystem II: a new class of 'rogue' cyanobacterial D1 proteins, *Photosynth Res* **110**, 177-184
- Sajjaphan, K., Shapir, N., Judd, A.K., Wackett, L.P., Sadowsky, M.J. (2002) Novel *psbA1* Gene From a Naturally Occuring Atrazine-Resistant Cyanobacterial Isolate, *Appl. Environ. Microbiol.*, **68**, 1358-1366
- Sane, P.V., Ivanov, A.G., Sveshnikov, D., Huner, N.P.A., Oquist, G. (2002) A transient exchange of the photosystem II reaction center protein D1:1 with D1:2 during low temperature stress of *Synechococcus* sp. PCC 7942 in the light lowers the redox potential of Q (B), *J. Biol. Chem.* **277**, 32739-32745
- Sicora, C.I., Ho, F.M., Salminen, T., Styring S., Aro, E.-M. (2009) Transcription of a "silent" cyanobacterial *psbA* gene is induced by microaerobic conditions, *Biochim.Biophys. Acta.* **1787**, 105-112
- Singhal, G.S., Renger, G., Sopory, S. K., Irrgang, K. D., Govindjee. (1999) Introduction to Photobiology, Photosynthesis and Photomorphogenesis, In: Concepts in photobiology, Photosynthesis and Photomorphogenesis, *Narosa Publishing House*, 1-7
- Summerfield, T. C., Toepel, J., Sherman, L. A. (2008) Low-oxygen induction of Normally Cryptic *psbA* genes in Cyanobacteria, *Biochemistry*, **47**, 12939-12941