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**STUDIA UBB EDITORIAL OFFICE:** B.P. Hasdeu no. 51, 400371 Cluj-Napoca, Romania,  
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#### **CUPRINS – CONTENT – SOMMAIRE – INHALT**

PETRE FLORINEL NENU, LUISA IZABEL DUNGAN, ADRIAN EUGEN CIOABLĂ, GERLINDE RUSU, GEZA NICOLAE BANDUR, ALINA GABRIELA DUMITREL, VASILE PODE, Biomass Analysis for Combustion Applications – Case Study Scenarios .....	7
KATARZYNA PSTROWSKA, JAN KACZMARCZYK, HANNA CZAPOR-IRZABEK, MAREK KUŁAŻYŃSKI, Characterization of Activated Beech Wood Char – Methane Storage Application.....	21
ADRIAN PATRUT, ROXANA T. PATRUT, LASZLO RAKOSY, ILEANA-ANDREEA RAȚIU, DANIEL A. LOWY, JENŐ BODIS, KARL F. VON REDEN, Radiocarbon Dating of the Old Ash of Aiton, Romania ..	41
IOAN PETEAN, GERTRUD ALEXANDRA PALTINEAN, AURORA MOCANU, DANA FLORINA MUNTEAN, LIANA MURESAN, GEORGE ARGHIR, MARIA TOMOAI-COTISEL, Micro and Nano Organization of Atmospheric Particulate Matter in Grigorescu District of Cluj-Napoca.....	49

SIMION DRĂGAN, Macrokinetic Investigations of Dry Flue-Gas Desulfurization Process Using TGA Technique .....	59
SORIN RAPUNTEAN, REKA BALINT, GERTRUD ALEXANDRA PALTINEAN, GHEORGHE TOMOAIA, AURORA MOCANU, CSABA-PAL RACZ, OSSY HOROVITZ, MARIA TOMOAIA-COTISEL, Antibacterial Activity of Silver Nanoparticles Obtained by Co-Reduction with Sodium Citrate and Tannic Acid .....	73
ILDIKO LUNG, MARIA-LOREDANA SORAN, OCSANA OPRIȘ, MANUELA STAN, CONSTANTIN BELE, Microwave Irradiation Effect on Polyphenol Content and Antioxidant Activity of Basil .....	87
AHMED. A. M. ELNOUR, MOHAMED E. S. MIRGHANI, N. A. KABBASHI, MD Z. ALAM, KHALID HAMID MUSA, Gum Arabic: An Optimization of Ultrasonic- Assisted Extraction of Antioxidant Activity .....	95
DAN-ALEXANDRU TĂTARU, DAN-MIRCEA OLINIC, ANDRADA URDĂ, MARIA OLINIC, REMUS ORĂSAN, Oxidative Stress Markers in Acute Myocardial Infarction Treated by Primary Percutaneous Coronary Intervention .....	117
CRISTIAN T. MATEA, TEODORA MOCAN, FLAVIU TABARAN, TEODORA POP, OFELIA MOSTEANU, LUCIAN MOCAN, CLAUDIU ZDREHUS, Synthesis and Characterization of MUC-1 Functionalized Gold Nanoparticles .....	129
RÉKA BARABÁS, MELINDA RIGÓ, MARGIT ENISZNÉ-BÓDOGH, CORINA MOISA, OANA CADAR, Preparation and Characterization of Hydroxyapatite Based Nano-Composite Biomorphic Implants.....	137
SILVIA BURCĂ, CERASELLA INDOLEAN, Ionic Exchange Studies for Correcting Water Quality Indicators.....	155

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## BIOMASS ANALYSIS FOR COMBUSTION APPLICATIONS – CASE STUDY SCENARIOS

PETRE FLORINEL NENU<sup>a</sup>, LUISA IZABEL DUNGAN<sup>\*a</sup>,  
ADRIAN EUGEN CIOABLĂ<sup>a</sup>, GERLINDE RUSU<sup>b</sup>,  
GEZA NICOLAE BANDUR<sup>b</sup>, ALINA GABRIELA DUMITREL<sup>b</sup>,  
VASILE PODE<sup>b</sup>

**ABSTRACT.** Nowadays the need for renewable resources becomes more and more stringent. Biomass remains one of the most important sources of energy, being used directly or by chemical conversion processes in order to produce clean energy. Relative to this aspect, the present paper proposes the direct approach, by analyzing biomass type materials (topinambur and Romanian hemp) in order to determine their physical and chemical characteristics and their potential use in combustion processes.

**Keywords:** *biomass, combustion, thermal and chemical analysis*

### INTRODUCTION

The worldwide socio-economical context imposes an economical growth in the sense of obtaining heat, electricity and biofuels, but the main objective for safe energy distribution has to be overlapped with the one of sustainable development of society, which represents an essential component of the 2009/28/EC Directive regarding Renewable Energy. Energetic durability is recognized as a decisive factor in reaching a global scale sustainable development of any country [1-3].

The EU has established ambitious objectives regarding climate and energy until 2020, such as a reduction of 20% for the GHG gases (greenhouse effect) and a quota of 20% from the energetic consumption in the UE should be produced by renewable energy sources. At global level there exists an urgent demand for bioenergy and the renewable energy sources are sufficient

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<sup>a</sup> Politehnica University Timisoara, Faculty of Mechanical Engineering, 1 Mihai Viteazu Blv, Timisoara, RO-30022, Timisoara, Romania

<sup>b</sup> Politehnica University of Timisoara, Faculty of Industrial Chemistry and Environmental Engineering, 6 V. Parvan blvd., RO-300223, Timisoara, Romania

\* Corresponding author: [luisa.dungan@upt.ro](mailto:luisa.dungan@upt.ro)

for offering potential to all energetic needs worldwide necessary in the next century [4-5]. The residual materials obtained from forestry, agriculture and organic waste (for example municipal solid waste) would be enough to ensure between 50 and 150 EJ/year and the difference can be obtained from energetic crops, excessive forest growing and optimized agricultural productivity [6]. Short rotation coppice (SRC) can be a factor in reduction of energetic dependence of a country and GHG and can contribute as a lever to regional durable development, especially in rural areas, where there is need for new working places and to create a durable economy in order to attract young population [7].

Aside solar and wind energy, bioenergy represents one of the most used renewable energy technologies [8], and at global level the main concern is to obtain independence from fossil fuels while identifying potential sources and increased usage on a large scale of “fuel mix”.

The concept “bioenergy” (energy from biomass), included in the term “green energy” (renewable energy), describes an alternative solution, ecological, apart from the classical form of using carbon based sources. If biomass is directly used for heat and / or energy generation, by being just a fuel (or an energy source), the energy generated from it is called bioenergy [9,10].

“The ratio between the mass of organic substance versus a certain surface and time unit” [11], as biomass can be defined, including all the organic matter produced through metabolic processes of living organisms in chemical form, constitutes in fact the resource with the highest potential usage for man. Biomass is in essence the ensemble for non-fossil organic matter, which includes: wood, animal residues, oils and vegetable residues from forestry, agriculture and industry but also cereals and fruits from which ethanol can be produced. Biomass reserves are especially wood residues, agricultural residues, municipal waste and energetic crops [12, 13].

Biomass can also be used for the production of biochemicals or biomaterials [14-16].

It is one of the most promising sources of renewable energy because of its attractive properties like low production cost, low emissions, storage capacity and availability throughout the year from different sources [17-20].

It is the only renewable energy source which can be transformed in solid, liquid or gaseous fuels [21, 22].

The main forms of capitalizing biomass are:

- Direct combustion for thermal energy production;
- Pyrolysis firing with syngas production ( $\text{CO} + \text{H}_2$ );
- Fermentation with biogas production ( $\text{CH}_4$ ) or bioethanol ( $\text{CH}_3\text{-CH}_2\text{-OH}$ );
- Chemical conversion of vegetal oils by alcoholic treatment and obtaining esters (eg. biodiesel and glycerol);
- Enzymatic decay with ethanol or biodiesel production [12].

So it can be considered that biomass has an important role to play in the energy production [13], especially in the developing countries [24].

Presently the use of biomass produces about 5% from the total energy consumption at European level and in countries like Finland, Sweden and Austria it assures over 15-20%. In Romania, the electric energy production in 2016 was accomplished at a level of around 42.38% from renewable energy sources and the rest through fossil fuels according to existing data given by the National Authority for Energetic Regulation (ANRE) from monitoring the electric energy market.

According to the presented document, from all the renewable energy sources, the hydroelectric part assured 28.86% from the total produced electric energy, followed by wind installations (10.13%), solar (2.60%) and biomass (0.75%) [24].

Until 2020, the ENERO study recommends local heating and cogeneration through using wood biomass, straw and biogas obtained from anaerobic digestion of organic residues as being the main effective technologies for Romania and biomass (including biofuels used for transportation) will be the main contributor at RES quota (Renewable Energy Sources) with more than 65% from the total amount [25].

Both presently and also as estimation for 2020, the main biomass contribution in our country is represented by wood and agricultural residues, used for heating the traditional rural stoves, which is about 40% from the living population [25].

Using biomass in firing processes still presents technical issues because of the difficulty to model the combustion behavior due to the large number of biomass types with different chemical compositions, which can vary significantly.

Biomass has certain disadvantages as solid fuel: high water and Oxygen content, hydrophilic nature, low calorific value, high bulk density and energetic density, and also posing problems regarding transportation, storage and usage. Transportation costs are generally high, which can affect the process economical viability [26].

In order to partially overcome those problems, biomass can be pretreated and transformed into products like pellets and briquettes. Through this type of processing it can be obtained a homogenous and solid product with high density, thus having an improvement in terms of biomass quality and reduction of the supply chain [27, 28].

Knowing the physical and chemical characteristics for the used products is very important in order to better understand the burned fuel behavior in order to better consolidate the biomass supply chain viability. The process of obtaining pellets / briquettes is improved in order to be used for capitalization of agro-industrial residues [29].

It is a known fact that the primary matter has a great influence over the qualities of pellets and briquettes. For forestry biomass, the leaves have a superior heating value than the wood fractions [30,31], but they also present a high ash content which implies more industrial problems like abrasive wear in firing boilers / ovens.

## RESULTS AND DISCUSSION

There were conducted both physical and chemical analysis and thermal analysis in order to better determine the potential applications for the studied materials in combustion processes.

Next tables will underline the obtained results for the laboratory determinations.

For comparison reasons there will be added two other different materials previously used for analysis purposes in order to observe a basic comparison relative to chemical properties [32] but having in mind the fact that the main focus is made on the first two materials used for laboratory determinations (positions 1 and 2 in the next tables).

**Table 1.** Experimental results (part 1)

No.	MATERIAL	Moisture content [%]	Ash content (db) [%]	Gross calorific value (db) [J/g]	Net calorific value (db) [J/g]
1	Hemp RO	15,5	2,87	19334	17940
2	Topinambur	11,3	5,94	18277	16903
3	Wheat bran	9,7	5,54	19034	17520
4	Two row barley	10,7	2,22	18354	16763

**Table 2.** Experimental results (part 2)

No.	MATERIAL	Carbon content (db) [%]	Hydrogen content (db) [%]	Nitrogen content (db) [%]	Volatile matter content (db) [%]
1	Hemp RO	48,3	5,4	0,45	79,1
2	Topinambur	45,6	5,59	1,46	74,3
3	Wheat bran	41,3	6,2	2,06	78,4
4	Two row barley	40,1	6,5	1,38	82,4

It can be observed from table 2 that the ash content of the two studied materials is relatively low, which represents a good indicator of combustion use due to the fact that there are low quantities of residual materials after the process. By comparison with other materials it can be observed that there can be lower values (two row barley in our case) that are better suited for further use in firing processes. Also the calorific values are relatively high, which implies high energetic levels for all materials.

**Table 3.** Experimental results (part 3)

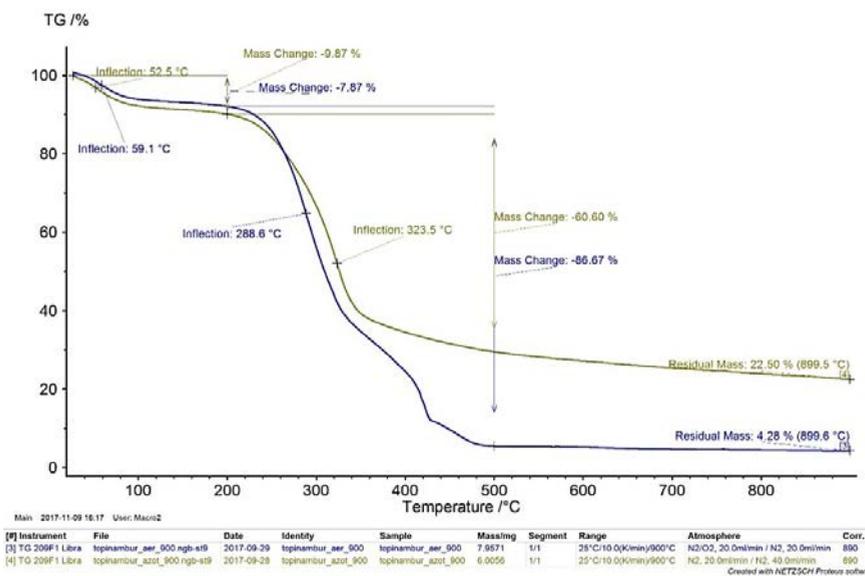
No.	MATERIAL	Sulphur content (db) [%]	Chlorine content (db) [%]	Oxygen Content (db) [%]
1	Hemp RO	0.008	0.001	43.0
2	Topinambur	0.117	0.258	41.0
3	Wheat bran	0.014	0.004	42.0
4	Two row barley	0.014	0.009	43.0

From tables 2 and 3 an overall idea is that there are low levels of sulphur and chlorine, which for combustion processes indicate low potential of degrading the furnace which uses as fuel one of the studied materials.

**Table 4.** Experimental results (part 4)

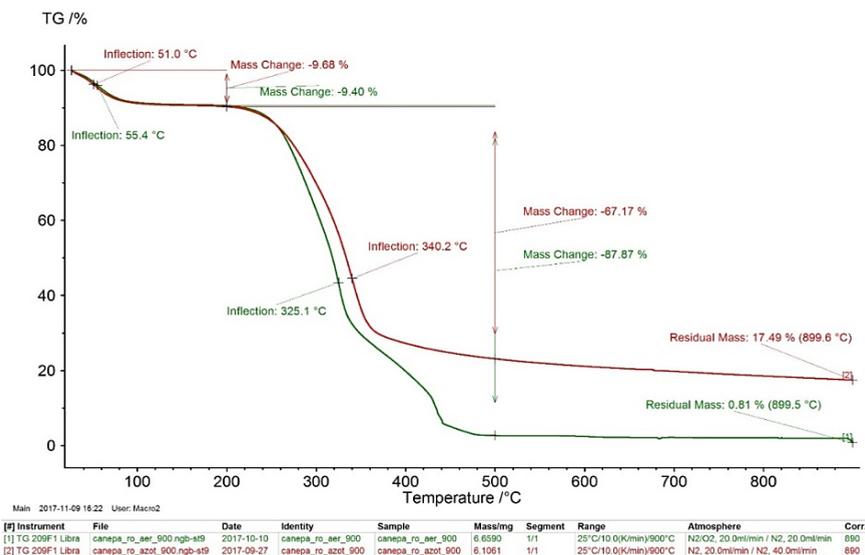
No.	MATERIAL	Shrinking temperature [° C]	Deformation temperature [° C]	Hemisphere temperature [° C]	Flow temperature [° C]
1	Hemp RO	960	1240	1300	1320
2	Topinambur	610	1280	1490	>1540
3	Wheat bran	640	670	730	740
4	Two row barley	830	1120	1300	1350

From table 4 one can determine that topinambur has a higher flow temperature than hemp, making the first material much more suitable in terms of behavior during the combustion process. By comparison with the other two materials, it can be observed that its flow temperature is in the same range as two row barley.



**Figure 1.** TG curves for topinambur under air and nitrogen

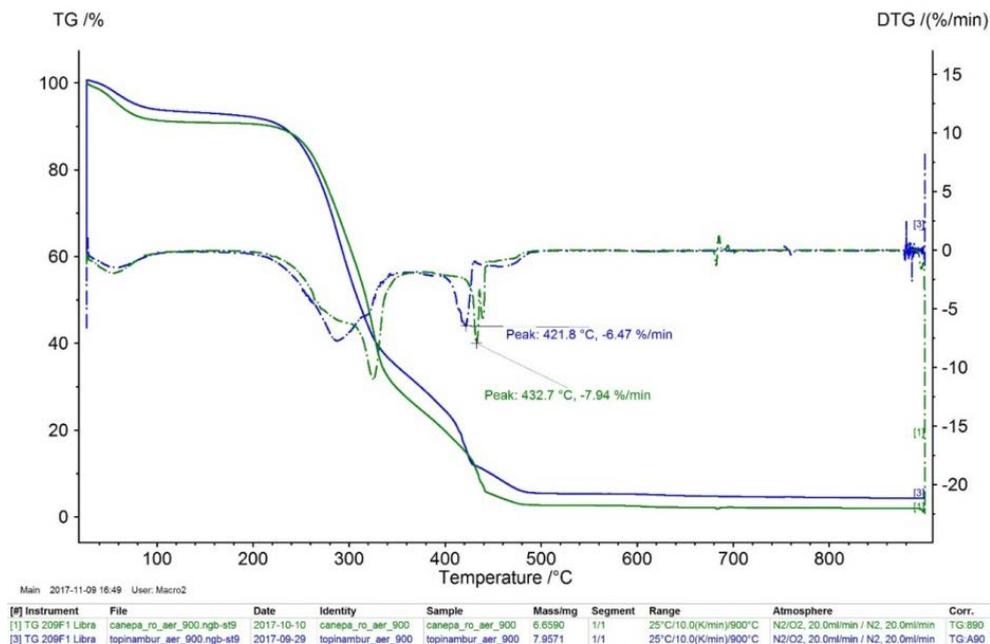
The TG curves present two degradation steps, the first from 25 to 200°C, associated with water loss, and the second from 200 to 500 °C associated to thermal decomposition of the samples.



**Figure 2.** TG curves for hemp under air and nitrogen

The mass loss for the first degradation step is under 10% (air or nitrogen) and for the second degradation step is under 90% for the samples under air and around 60% for the samples under nitrogen. The residual mass is under 5% for the sample in air and around 20% for the samples under nitrogen.

The thermal decomposition behavior under air and under nitrogen is similar for the studied samples. Due to the complex reaction, that take place under air, both samples presents around 425 °C another degradation step (figure 3), step that is missing in the analysis recorded under nitrogen (figure 4).



**Figure 3.** TG curves of topinambur and hemp under air

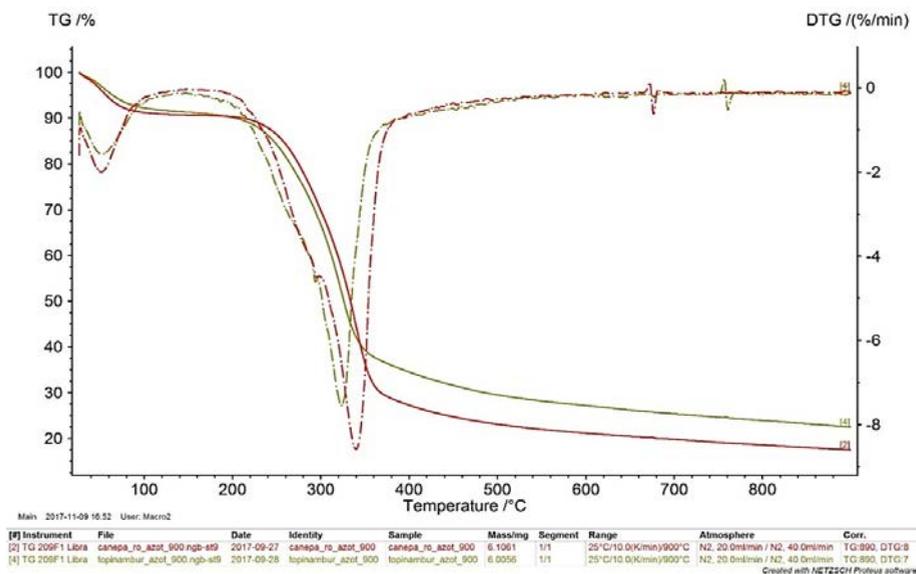


Figure 4. TG curves of topinambur and hemp under nitrogen

Table 5 presents the overall characteristics determined during the thermogravimetric analysis.

Table 5. Thermogravimetric analysis

Sample	Inflexion		Mass loss (%)		Residual mass (%)
	First step	Second step	25-200°C	200-500°C	
Topinambur air	59.1	288.6	7.87	86.67	4.28%
Topinambur N <sub>2</sub>	52.5	323.5	9.87	60.60	22.50
Hemp air	55.4	325.1	9.40	87.87	0.81
Hemp N <sub>2</sub>	51.0	340.2	9.68	67.17	17.49

The first mass loss is not influenced by the medium in which the analysis is carried out (nitrogen or air) thus the temperatures values are between 50 and 60 °C, this being physical process (dehydration).

The second step is the chemical decomposition of the samples and in this case the temperatures are lower in air medium as compared in nitrogen. There is a 20 °C difference in the inflexion temperatures of the decomposing processes. Also, the mass loss is with 20% higher for the second decomposing step in air as compared in nitrogen medium.

The residual mass has lower value for the analysis that was carried out in air, fewer than 5%. Under nitrogen atmosphere the residual mass is around 20%; the difference is that in nitrogen atmosphere the carbon is decomposed as carbon black which is not oxidized to CO<sup>2</sup>.

## CONCLUSIONS

From the chemical analysis made on the two studied material, it can be considered that both of them have a relatively high energetic potential and can further be used in firing processes.

The potentially corrosive elements inside the studied materials are in low quantities and pose no risks for the burning chamber in terms of deterioration on the long run.

Topinambur presents a higher flowing temperature which makes it more difficult to liquefy and from this point of view, hemp is easier to be evaluated and approached relative to its heating properties.

Topinambur and hemp are valuable raw materials that can be used in combustion processes. These materials were submitted for physical, chemical and thermal analysis in order to determine the potential applications in combustion processes.

From the thermogravimetric analysis can be concluded that the chemical decomposition take place between 200 and 500°C. Under air, the decomposition processes have complex reactions, which take place in two steps. The residual mass was found to be under 5% under air and around 20% under nitrogen atmosphere.

## EXPERIMENTAL SECTION

In the present paper there are studied 2 materials in terms of physical, chemical and thermo analytical properties relative to their potential use in combustion processes, namely topinambur and Romanian hemp.

The used standards for laboratory determinations were:

- EN ISO 18134 – Solid biofuels – Determination of moisture content – Oven dry method (3);
- EN ISO 18122- Solid biofuels - Determination of ash content;
- EN 14918 - Solid biofuels –Determination of calorific value;

- EN ISO 16948 – Solid biofuels – Determination of total content of carbon, hydrogen and nitrogen
- EN ISO 16994 - Solid biofuels — Determination of total content of sulfur and chlorine
- EN ISO 18123 – Solid biofuels – Determination of the content of volatile matter.
- CEN/TS 15370 - Determination of ash melting behavior.

*Determination of moisture content.*

- inside each vessel it was weighed approximately 1 g of material (balance precision 0.0001),
- there were made 3 time determinations for each sample,
- the samples were introduced inside the furnace at 105 °C for a period until constant mass,
- after the drying period, the samples were weighed again and the data were introduced in the specified protocol,
- for precision determinations, for samples with a high moisture content the determinations were repeated once more.

*Ash content determination.*

- before introducing the samples inside the furnace, the crucibles are let inside for about 2 hours at 550 °C and after this operation they are let to cool down for 10 – 15 minutes,
- after the crucibles are cooled, weighed and inside each of them it will be weighed minimum 1 g of material (balance precision 0,0001),
- the furnace is switched off after this step in order to cool down until a temperature lower than 100 °C,
- the process of ash content determination has 2 steps: first, the crucibles with the weighed samples are introduced inside the furnace and let at least 1 hour at 250 °C to allow volatile matter elimination before ignition (the temperature is raised up constantly to 250 °C over a period of 30 – 50 minutes),
- after this initial step, the temperature inside the furnace will be raised to 550 °C and these samples will reside for a minimum of two hours
- after the determination the samples are put down from the furnace and let to cool for 2 -3 -5 minutes outside, and the cooled samples are then weighed and the results are introduced inside the corresponding protocol,
- for precision determinations, for samples with a high ash content the determinations were repeated once more
- because of the high temperatures it is difficult to number the crucibles, hence the numbered position inside the protocols,
- the ash content was corrected to dry basis by using the moisture content.

*Calorific value determination.*

- before calorific value determination, the samples are prepared for the analysis inside the calorimetric bomb,
- the preparation part is related to using a hydraulic press in order to make a small pill like pellet of 0.3 – 0.5 g which is going to be inserted into the bomb, which in its turn will be introduced inside a calorimeter (Model IKA C 6000), because the expected heating value is in the calibrated range,
- after the material data is introduced in the bomb protocol (weighed quantity, identification number for the sample and the user) the determination starts,
- for each material there were made at least 2 determinations, in order to verify if the difference in calorific value between both determinations is lower than 120 J /g,
- after the bomb finishes the digestion process, its interior is washed with ultra pure water and the washing liquid was filled into a 100 ml flask until mark with ultra pure water for each sample with the corresponding identification number on it,
- the obtained values are then introduced inside the corresponding protocol for obtaining the calorific value.

*Ion chromatography*

- this type of chromatography is used in general for liquid analysis. the used apparatus is a Metrohm 930 model equipped with calibration for fluoride, chloride, nitrites, nitrates, bromides, sulfates and phosphates,
- the obtained values are presented in form of peaks on a chromatogram from which are selected the retention times according to the used standard (reference standard solutions inside the carousel), measured and compared with the existing etalon (for the used samples there were determined just the nitrates and sulphates for correcting the obtained calorific value),
- inside the carousel are introduced samples (vessels with a volume of approximately 10 ml) and after this step there are made system check-ups for all the components

*Carbon, Hydrogen and Nitrogen determinations*

- before the analysis, a general system check takes place using “blank” tests and reference materials for equipment calibration,
- those results are used also for recalculating the results using the new calculated value which is made after obtaining constant values for the three presented elements,
- between the samples are put standard materials in order to keep the equipment calibrated and the measurement precise

- the sample is weighed (at the maximum possible amount of material for a domain between 0.1 and 0.3 g / sample, because of the volume of the material) and is introduced inside a Zinc foil and into the carousel of the equipment in parallel with introducing the sample data inside the protocol,
- for analysis is used Oxygen of high purity, for a complete combustion of the resulting combustion gas and Helium for Nitrogen content determination.

*Determination of the volatile matter content*

- the empty crucibles are introduced inside the furnace at  $900\text{ }^{\circ}\text{C} \pm 10\text{ }^{\circ}\text{C}$  for about 60 minutes and after this period are put out and let to cool down for 20 minutes
- after the crucibles are cooled down it is weighed a quantity of minimum  $1\text{g} \pm 0.1\text{ g}$  of material,
- the crucibles are then inserted into the furnace for exactly 7 minutes and after this time period are let out to cool to room temperature
- after the crucibles are cooled down, are weighed again and the results are inserted into the corresponding protocol,
- through mass difference the volatile matter content is determined for the considered samples (three times determination),
- the volatile matter was corrected to dry basis by using the moisture content.

*Determination of ash melting behavior - CEN/TS 15370 - Determination of ash melting behavior*

For this type of determination there is used a thermal microscope – model IRF 1600 F

First, the materials have to be prepared: the ash is obtained using a combination of temperatures starting at  $200\text{ }^{\circ}\text{C}$  in order to degas the sample, for about 20 hours, after which the temperature is increased slowly for about 1 hour to  $250\text{ }^{\circ}\text{C}$  and another hour from  $250\text{ }^{\circ}\text{C}$  to  $300\text{ }^{\circ}\text{C}$ , after this step, the temperature is risen from  $300\text{ }^{\circ}\text{C}$  to  $550\text{ }^{\circ}\text{C}$  during a period of 6 hours and in the last period the material is let to preash at  $550\text{ }^{\circ}\text{C}$  for about 2 hours.

After the ash is obtained, the next step is to prepare the sample.

A part of the ash is mixed with ethanol and introduced in a special device in order to obtain a small pellet which will be introduced inside the thermal microscope.

The pellet is introduced and kept inside the thermal microscope for three hours.

During this time, there can be observed the material modifications because of temperature influence. The main points of interest are: shrinking point, deformation point, hemisphere point and flow point.

At the end of the process, the materials are put out of the thermal equipment and readings are made with the help of specific software in order to determine the correct temperature values.

Thermogravimetric analysis (TGA) were performed on TG 209 F1 Libra equipment (Netzsch) under nitrogen and air atmosphere and under dynamic conditions 10 K/min from 20 to 900 °C.

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## REFERENCES

1. M.A. Rosen, *Sustainability*, **2009**, *1*, 55.
2. E. Toklu, M.S. Güney, M. Işık, O. Comaklı, K. Kaygusuz, *Renewable & Sustainable Energy Reviews*, **2010**, *14*, 1172.
3. K. Hakeem, M. Jawaid, U. Rashid, “Biomass and bioenergy: applications”, Springer-Verlag, New York, **2014**.
4. K. Bilen, O. Ozyurt, K. Bakırcı, S. Karşlı, S. Erdogan, M. Yılmaz, et al., *Renewable & Sustainable Energy Reviews*, **2008**, *12*, 1529.
5. O. Edenhofer, R. Pichs Madruga, Y. Sokona, *Climate Policy*, **2012**, *6*, 1088.
6. J. Popp, Z. Lakner, M. Harangi-Rakos, M. Fari, *Renewable & Sustainable Energy Reviews*, **2014**, *32*, 559.
7. S. Pereira, M. Costa, M.G. Carvalho, A. Rodrigues, *Energy Conversion and Management*, **2016**, *125*, 242.
8. W.H. Chen, W.Y. Cheng, K.M. Lu, Y.P. Wuang, *Applied Energy*, **2011**, *88*, 3636.
9. U.S. Energy Information Administration. Biomass. Glossary, **2016**, available at: <https://www.eia.gov/tools/glossary/index.cfm? Id=B>, (accessed 03 January 2018).
10. DOE. Biomass Feedstocks, **2016**, available at: <http://www.energy.gov/eere/bioenergy/biomass-feedstocks>, (accessed 03 January 2018).
11. Dicționar explicativ al limbii române, available at: <https://dexonline.ro/definitie/biomas%C4%83/4403>, (accessed 03 January 2018).
12. A. Balázsi, „Biomasa ca sursă de energie regenerabilă, analiza tehnologiilor de obținere a energiei din aceasta”, **2013**, available at: <http://www.urbanaodorhei.ro/dokumentumok/Informaciok /Biomasa.pdf>, (accessed 03 January 2018).

13. Renewables 2014 Global Status Report. Renewable Energy Policy Network for the 21 St Century, REN21 Secretariat, Paris, **2014**, EN21.
14. D. Pleissner, W.C. Lam, W. Han, K.Y. Lau, L.C. Cheung, M.W. Lee, *BioMed Research International*, **2014**, 8, available at: <http://dx.doi.org/10.1155/2014/819474>.
15. W. Han, W.C. Lam, M. Melikoglu, M.T. Wong, H.T. Leung, C.L. Ng, *ACS Sustainable Chemistry & Engineering*, **2015**, 3, 2043.
16. M. Melikoglu, C.S.K. Lin, C. Webb, *Food and Bioproducts Processing*, **2015**, 95, 63.
17. C.S. Chou, S.H. Lin, C.C. Peng, W.C. Lu, *Fuel Processing Technology*, **2009**, 90, 1041.
18. M. Main-Knorn, W.B. Cohen, R.E. Kennedy, W. Grodzki, D. Pflugmacher, P. Griffiths, *Remote Sensing of Environment*, **2013**, 139, 277.
19. A. Nansaior, A. Patanothai, A.T. Rambo, S. Simaraks, *Biomass Bioenergy*, **2013**, 52, 113.
20. A. Aslan, *Renewable & Sustainable Energy Reviews*, **2016**, 57, 362.
21. F. Rosillo-Calle, P. de Groot, S.L. Hemstock, J. Woods, "The Biomass Assessment Handbook: Bioenergy for a Sustainable Environment", Earthscan, London, **2007**.
22. E. Ntona, G. Arabatzis, G.L. Kyriakopoulos, *Renewable & Sustainable Energy Reviews*, **2015**, 46, 1.
23. M.F. Demirbas, M. Balat, H. Balat, *Energy Conversion and Management*, **2009**, 50, 1746.
24. C. Zaharia, Mix-ul perfect în energie, available at: <http://www.green-report.ro/mix-ul-perfect-in-energie-42-din-productia-de-energie-electrica-a-romaniei-asigurata-din-surse-regenerabile/>, (accessed 03 January 2018).
25. Ministerul Economiei, Comertului si Mediului de Afaceri – Romania, NL Agency – Olanda, ENERO – Romania, „Master Plan Biomasă pentru România”, București, **2010**, available at: [http://www.minind.ro/biomasa/Plan\\_de\\_Actiune\\_pentru\\_Biomasa.pdf](http://www.minind.ro/biomasa/Plan_de_Actiune_pentru_Biomasa.pdf), (accessed 03 January 2018).
26. M. Campoy, A. Gómez-Barea, P. Ollero, S. Nilsson, *Fuel Processing Technology*, **2014**, 121, 63.
27. A. Demirbas, Chapter 2: "Fuels from Biomass", In: Demirbas, A. (Ed.), "Biorefineries: for biomass upgrading facilities". Springer, London, **2010**, 33.
28. N. Kaliyan, R.V. Morey, *Biomass and Bioenergy*, **2009**, 33, 337.
29. F.F. Felfli, P.J.M. Mesa, J.D. Rocha, D. Filippetto, C.A. Luengo, W.A. Pippo, *Biomass and Bioenergy*, **2011**, 35, 236.
30. L. Cuiping, W. Chuangzhi, Yanyongjie, H. Huang, *Biomass and Bioenergy*, **2004**, 27, 119.
31. S. Pérez, C.J. Renedo, A. Ortiz, M. Mañana, *Biomass and Bioenergy*, **2011**, 35, 4657.
32. A.E.Cioablă, N. Pop, G. Trif-Tordai, D.G. Călinoiu, *Journal of Thermal Analysis and Calorimetry*, **2017**, 127, 515.

## CHARACTERIZATION OF ACTIVATED BEECH WOOD CHAR – METHANE STORAGE APPLICATION

KATARZYNA PSTROWSKA<sup>a,\*</sup>, JAN KACZMARCZYK<sup>a</sup>,  
HANNA CZAPOR-IRZABEK<sup>b</sup>, MAREK KUŁAŻYŃSKI<sup>a</sup>

**ABSTRACT.** Commercial beech wood char characterized by low volatile matter (5.3%) and high carbon content (94.5 %) was initially used as a potential feedstock to obtain the activated carbon for methane sorption. Micro- and mesopores volumes were determined to be at the level of  $0.21 \text{ cm}^3\cdot\text{g}^{-1}$ , while the specific surface area of the material was determined to be at the level of  $416 \text{ m}^2\cdot\text{g}^{-1}$ . Carbonization carried out at the temperature range of 750-900 °C resulted in slight increase of the pore volume (up to the value of  $0.23 \text{ cm}^3\cdot\text{g}^{-1}$ ) and the specific surface area (up to the value of  $480 \text{ m}^2\cdot\text{g}^{-1}$ ). Upon the steam activation (up to the 50 % loss of the organic mass) the specific surface area increased substantially and was within the range of 887 - 943  $\text{m}^2\cdot\text{g}^{-1}$  together with micro- and mesopores volumes that was within the range of 0.54 - 0.58  $\text{cm}^3\cdot\text{g}^{-1}$ . The maximum methane adsorption was determined for the sample activated at a temperature of 800 °C –  $17.7 \text{ g}\cdot\text{kg}^{-1}$  with a process reversibility of 84.2%. Considering the beech wood char price, its availability and well-known methods of its surface structure improvement, the obtained active carbons are interesting for their testing on a larger scale.

**Keywords:** adsorption; biomass; active carbon; steam activation; carbonization

### INTRODUCTION

Natural gas, whose main component is methane, is more environmentally friendly and cheaper than traditionally used crude oil-origin fuels [1]. Among all hydrocarbons, methane has the highest hydrogen-to-carbon ratio and consequently a higher research octane number (RON = 107).

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<sup>a</sup> Wrocław University of Science and Technology, Faculty of Chemistry, Division of Fuels Chemistry and Technology, Gdańska 7/9, 50-344 Wrocław, Poland

<sup>b</sup> Wrocław Medical University, Faculty of Pharmacy, Laboratory of Elemental Analysis and Structural Research, Borowska 211A, 50-556 Wrocław, Poland

\* Corresponding author: [katarzyna.pstrowska@pwr.edu.pl](mailto:katarzyna.pstrowska@pwr.edu.pl)

Among all hydrocarbon fuels, combustion of methane produces the smallest amount of CO<sub>2</sub> for each unit of heat that is released [2].

Four different methods were proposed for the storage of natural gas, i.e., liquefied natural gas (LNG), compressed natural gas (CNG), adsorbed natural gas (ANG) and natural gas hydrate (NGH). Among these, ANG technology enables the greatest efficiency in the storage of natural gas [3]. In a conventional high-pressure storage tank, gas is forced into the tank under pressure. Greater pressures enable larger volumes of gas to be stored in the tank. The maximum pressure, and therefore the volume of gas held in the tank, is limited by the physical properties of the tank and its valve. Microporous materials – methane adsorbents, which are used as a filling of a gas tank – may perform two functions: reducing the pressure of gas storage (reasons of the tank user safety) and increasing the volume of the gas storage in the tank (economic reasons - a less frequent exchange/tank filling). Development of efficient and cost-effective adsorbent materials is the key to a successful adsorbed natural gas technology. The most important attributes of commercial ANG sorbent is to have a high surface area per unit mass with regular porous structure, high adsorption capacity, mechanical resistance to pressure variations, and high reversibility of the CH<sub>4</sub> adsorption.

The search for suitable porous material in terms of improving ANG storage volumetric energy density and lowering the adsorbent cost to the end user is currently an active area of research. Among different porous materials, metal organic frameworks (MOFs), covalent organic frameworks (COFs), zeolites, and active carbons (ACs) have been intensively examined. MOF chemistry offers potential challenging issues typical to CH<sub>4</sub> storage due to their unique modularity, unprecedented tunability, and high degree of porosity combined with diverse framework functionalities. However, the commercial applications of MOFs are still restricted due to their limited mechanical robustness, low density, low volumetric adsorption in a synthesized powder form [4-5]. A comprehensive understanding on stability and performance is required against the typical operating conditions to realize these materials for commercial applications. Also, from the commercial point of view, high production costs strongly limit the possibility of their use. Comparing to all types of other potential sorbents, MOFs are also characterized by other factors such as open metal sites, porosity, topologies and framework densities, which can strongly improve methane adsorption capacity [5-9]. Covalent organic frameworks (COFs) are another class of crystalline materials built from organic linkers, which are made up of light elements (C, O, B, and Si) and held together by strong covalent bonds (–O, C–C, B–C, C–N and C–Si) [10-11]. Compared to MOFs, COF materials are more favorable when accounting for gravimetric uptake of natural gas compounds. Both MOFs and COFs are characterized as

crystalline, porous types of materials. Nevertheless, the absence of metal elements in the COF structure strongly reduces the synthesis costs of the material. One of the major advantages compared to MOFs is the possibility of obtaining COFs of similar porous structure characterized with a much lower density [5, 10, 12]. Still, a comprehensive understanding on material stability and large-size methods of production is required before the commercial application of COFs can be possible. The other disadvantage of these materials as methane sorbents is the small number of published experimental studies available (calculated vs. experimental results). To a large extent, the methane sorption capacity and the possibility of achieving extensive surface of the COFs is analyzed by computational methods [13-15]. Practical application of these materials therefore requires a considerable intensification of laboratory tests.

Several materials created by nature have a regular structure that can be modified by simple, well-known physico-chemical methods and lead to materials having a well-developed porous structure. Among them, zeolites and carbonaceous materials (as selected biomass materials) play the most important role. As potential methane sorbents, molecular sieves have strong advantages, such as being naturally regular, stable, and controllable channel structures and having similar surface chemical structures [16]. Methane adsorption on natural zeolite adsorbents has been widely investigated, as in case of the use of calculation methods as well as experimental studies, e.g. [17-19]. It was proven that zeolites have relatively high packing densities (compared to ACs) but have lower micropore volumes. In addition, zeolites are extremely hydrophilic and can lose their adsorption capacity for methane with time due to preferential moisture adsorption. On the other hand, it has been shown that ACs are very good adsorbents, presenting the highest ANG energy densities and thus the highest storage capacities. ACs can be essentially microporous materials with low mesoporosity and absence of macroporosity if they are properly prepared. This leads to a high packing density and an important volumetric capacity for methane storage. ACs obtained from biomass are characterized primarily by low price and high availability. Regardless of the above, the primary criterion for the selection of biomass raw material as active carbon is a high carbon content and low volatile matter, which provides stable structure and high production capacity in the process of thermal treatment. Among the many methods used, the most common are chemical activation (mainly with the use of KOH as activating agent [1,20-22]) and physical activation, where steam and/or CO<sub>2</sub> is used as activation agent, e.g. [23-26]. Textural properties (i.e., adsorption capacity [surface area and micropore volume] or the micropore size distribution) are the main factors responsible for gas uptake. The pores' shape, size, surface area, and mechanical resistance to pressure changes of the material is affected not only by the activation

method but the reactivity of the material. So far, several biomass materials have been used as AC precursors for methane storage. Coconut shells were used by Azevedo et al. [27] for the preparation of ACs with chemical activation ( $\text{ZnCl}_2$  as the activation agent) followed by physical activation. Depending on the amount of the activating agent, the authors obtained materials characteristic of 1090-2270  $\text{m}^2/\text{g}$  BET surface and 80-92% microporosity. A maximum adsorption of the methane at 25 °C and 40 bar was demonstrated at the level of ca. 122 mg/g with almost 94 % reversibility. KOH-activated sugarcane molasses was used by Sreńscek-Nazzal et al. [22]. The highest methane adsorption was measured at the level of ca. 197 mg/g at 20 °C and 50 bar. BET surface area of the materials obtained at the temperature range of 400-800 °C was determined at the level of 413-1202  $\text{m}^2/\text{g}$ . To the authors' knowledge, ACs were both prepared chemically, as physical activation is mainly used for water purification and removal of specific organic compounds from water or gaseous mixtures. The abovementioned examples of the coconut shells [27] and sugarcane molasses [22] indicate the possibility of obtaining a good sorbent with high sorption methane capacity at a low cost of material production.

This study sought to verify the possibility for using low-cost biomass material as a methane sorbent precursor. High carbon content and low volatile matter were decisive for the use of beech wood char as a commercially available and low-cost material. According to the above comments, the present paper focuses on (1) analysis and discussion of the potential use of beech wood char as an active carbon precursor, (2) analysis of the influence of carbonization and activation parameters on the porous structure of beech wood char and the possibility of using the obtained materials as methane sorbents, including adsorption capacity and reversibility of the process.

## **RESULTS AND DISCUSSION**

### **Beech wood char characterization**

The commercial beech wood char material (KB) is characterized by ca. 17 % content of the ballast (the total amount of moisture and ash) and a small amount of volatile matter (5.3% *daf*, Table 1). A low content of volatile matter shows that the industrial producer of the char used a higher carbonization temperature than the temperature commonly used in case of commercial carbonization (usually used in the range of 500 - 600 °C [28]). A higher beech wood carbonization temperature was also confirmed by elemental composition of the material. Such high content of carbon in case of the beech wood char

was previously observed by Zeng et al. [29] (94.4 wt. % at the pyrolysis final temperature of 1600 °C), and Guizani et al. [30] (ca. 91 wt. % calculated on dry basis, pyrolysis temperature at the level of 900 °C). KB material containing a small amount of volatile matter and built mainly of carbon meets two basic criteria regarding the proper raw material for the activated carbon production process.

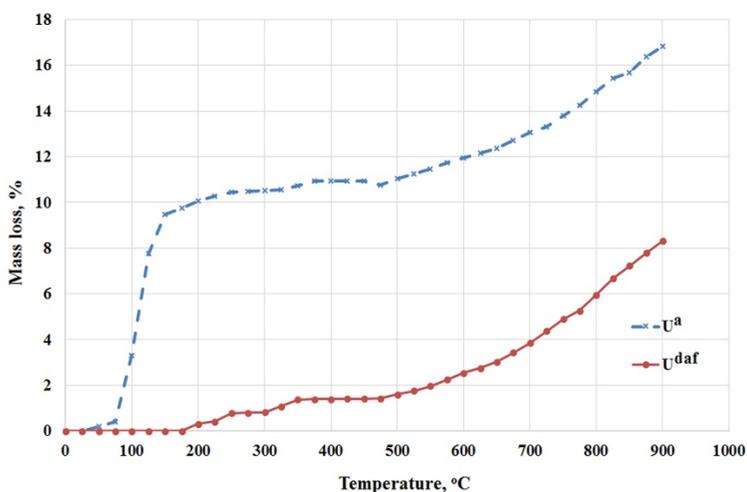
**Table 1.** Characterization of the beech wood char (KB)

Proximate analysis					Ultimate analysis				
Moisture, %	Ash content, %		Volatile matter, %		C%	H%	N%	S%	O%
analytical	analytical	dry	analytical	dry ash free	dry ash free				
9.8	5.7	6.3	4.5	5.3	94.5	0.5	1.2	0.5	3.3
Porous structure									
Pore width, nm	Micropores		Mesopores					Micro- & Mesopores	
	< 0.4	0.4 - 2	2 - 3	3 - 5	5 - 10	10 - 50	2 - 50		
Volume, cm <sup>3</sup> ·g <sup>-1</sup>	0.009	0.171	0.021	0.005	0.003	0.001	0.030	0.210	
Surface, m <sup>2</sup> ·g <sup>-1</sup>	24	472	17.6	2.9	0.9	0.1	21.5	518	

For the determination of the porous structure, CO<sub>2</sub> and C<sub>6</sub>H<sub>6</sub> sorption experiments were performed. Characteristic pore volume and pore surface size of the KB material is presented in Table 1. The total surface area of the KB calculated with the BET method was determined at the level of 416 m<sup>2</sup>·g<sup>-1</sup>. Volume and surface areas of the micropores (V<sub>MIKCO<sub>2</sub></sub> and S<sub>CO<sub>2</sub></sub>) available for carbon dioxide and inaccessible for benzene were calculated at the level of 0.180 cm<sup>3</sup>·g<sup>-1</sup> and 474 m<sup>2</sup>·g<sup>-1</sup>, respectively. The average mesopore diameter (d<sub>MES</sub>) was calculated as 2.79 nm.

Beech wood char is built mainly of large-width micropores available to the benzene molecules. The micropore volumes (0.4 - 2.0 nm width) were calculated at the level of 0.171 cm<sup>3</sup>·g<sup>-1</sup>, which is ca. 81.4% of the volume of all pores accessible for the adsorption. In a much smaller part, the material consists of mesopores (14.3%) and sub-micropores (4.3%). A significant share of micropores of a large volume predicts that the activation process will provide active carbons characterized with large pore volumes and surfaces.

For the thermogravimetric analysis (TG) of KB, the char was heated in argon in the temperature range of 25 - 900 °C at a gas flow rate of 5 °C·min<sup>-1</sup>. The resulting mass loss (U) depending on the temperature is shown in Fig. 1. The curve shapes of the TG results (Figure 1) confirm earlier observations that the producer of the beech wood char during the carbonization process used a higher temperature than is usually used. At the initial heating (25 - 200 °C), only the material moisture was eliminated. In a rather wide temperature range (200 - 650 °C), weight loss of the sample was small (weight loss at the level of ca. 3% of organic matter). A gradual and small weight loss with further increases in temperature up to the value of 900 °C and after 30 min at the 900 °C was reached, and 16.8% of the sample with respect to the initial weight and 8.3% of the weight of the sample calculated on a dry, ash-free state was observed.



**Figure 1.** Thermograms obtained under an argon atmosphere and beech wood char (KB) heating

### **Influence of the carbonization temperature on the porous structure of the beech wood char**

The size and design of the pores in activated carbon is highly dependent on the pore system of the material formed in the carbonization step, which in turn is largely dependent on the carbonization temperature. In the process of char activation, the original porous system is only developed and modified but it is initiated already in the process of carbonization. The temperatures of 750, 800, 850 and 900 °C were applied for optimization of the carbonization final temperature of KB.

KB was carbonized in the TG apparatus (Figure 3) under an argon atmosphere with a flow rate of  $30 \text{ dm}^3 \cdot \text{h}^{-1}$  and heating rate of  $10 \text{ }^\circ\text{C} \cdot \text{min}^{-1}$  up to the desired final temperature. At the final carbonization temperature, the material was conditioned for 30 min. The weight loss and yield of the products depending on the temperature of the process is summarized in Table 2.

Naturally, the thermal vulnerability of commercial beech wood char gradually decreases on a scale of rising carbonization temperature. An increase of temperature (from 750 up to 900  $^\circ\text{C}$ ) results in only a 2.5% weight loss (Table 2), which shows good thermal stability of the KB material. It is worth noting that this small loss contributes no decomposition of organic matter resulting from the supply to the increasing energy but also the inequalities associated with the industrial production cycle of the beech wood.

**Table 2.** Carbonization of the beech wood char – mass balance of the process

Sample	Final carbonization temperature, $^\circ\text{C}$	Mass loss, $U^a$ , %	Yield, %
KB <sub>750</sub>	750	14.2	85.8
KB <sub>800</sub>	800	15.3	84.7
KB <sub>850</sub>	850	16.5	83.5
KB <sub>900</sub>	900	16.7	83.3

Using the obtained carbon dioxide adsorption isotherms of the adsorption/desorption isotherms of benzene at 25  $^\circ\text{C}$ , the volume and surfaces of the micro- and mesopores were calculated and summarized in Table 3. The average pore diameter of the mesopores was determined in the range of 3.30 - 3.51 nm. With increasing carbonization temperature, the average pore diameter decreased. The volume and surface area of the micropores ( $V_{\text{MIKCO}_2}$  and  $S_{\text{CO}_2}$ ) available for carbon dioxide and inaccessible to the benzene were as follows:

KB <sub>750</sub>	$V_{\text{MIKCO}_2} = 0.155 \text{ cm}^3 \cdot \text{g}^{-1}$	$S_{\text{CO}_2} = 408 \text{ m}^2 \cdot \text{g}^{-1}$
KB <sub>800</sub>	$V_{\text{MIKCO}_2} = 0.190 \text{ cm}^3 \cdot \text{g}^{-1}$	$S_{\text{CO}_2} = 499 \text{ m}^2 \cdot \text{g}^{-1}$
KB <sub>850</sub>	$V_{\text{MIKCO}_2} = 0.175 \text{ cm}^3 \cdot \text{g}^{-1}$	$S_{\text{CO}_2} = 460 \text{ m}^2 \cdot \text{g}^{-1}$
KB <sub>900</sub>	$V_{\text{MIKCO}_2} = 0.178 \text{ cm}^3 \cdot \text{g}^{-1}$	$S_{\text{CO}_2} = 468 \text{ m}^2 \cdot \text{g}^{-1}$

**Table 3.** Porous structure of the materials obtained in the beech wood char carbonization

		Micropores		Mesopores					Micro- & Mesopores	S <sub>BET</sub>
		<0.4	0.4-2	2-3	3-5	5-10	10-50	2-50		
Volume, cm <sup>3</sup> ·g <sup>-1</sup>	KB <sub>750</sub>	0	0.173	0.009	0.006	0.005	0.003	0.023	0.196	-
	KB <sub>800</sub>	0	0.201	0.015	0.006	0.004	0.003	0.028	0.229	-
	KB <sub>850</sub>	0	0.193	0.016	0.009	0.004	0.005	0.034	0.227	-
	KB <sub>900</sub>	0	0.192	0.019	0.007	0.006	0.005	0.037	0.229	-
Surface, m <sup>2</sup> ·g <sup>-1</sup>	KB <sub>750</sub>	0	476	7.9	3.5	1.4	0.3	13.1	489	427
	KB <sub>800</sub>	0	555	11.9	3.6	1.1	0.3	16.9	572	496
	KB <sub>850</sub>	0	534	13.6	5.3	1.1	0.4	20.4	554	479
	KB <sub>900</sub>	0	529	16.3	3.8	1.8	0.5	22.4	551	480

The data summarized in Table 3 indicate that increasing the temperature of carbonization had a relatively limited effect on the pore structure. Minor differences can be seen in the slightly increased values of the micro- and mesopore volume. As a result of raising the temperature from 750 to 900 °C, the total volume and surface area of the pores were increased slightly in the range of 0.19 - 0.23 cm<sup>3</sup>·g<sup>-1</sup> and 490 - 570 m<sup>2</sup>·g<sup>-1</sup>, respectively. The obtained materials are highly microporous materials from their 84 - 88% share in the volume of the adsorption pores. It is expected that the char obtained at the carbonization temperature of 800 °C, which exhibited the highest micropore volumes, may be the best material to obtain activated carbon for gas storage. At the same time, KB<sub>800</sub> characterized by the largest BET surface area (496 m<sup>2</sup>·g<sup>-1</sup>) of all obtained carbonization products is a promising sorbent material.

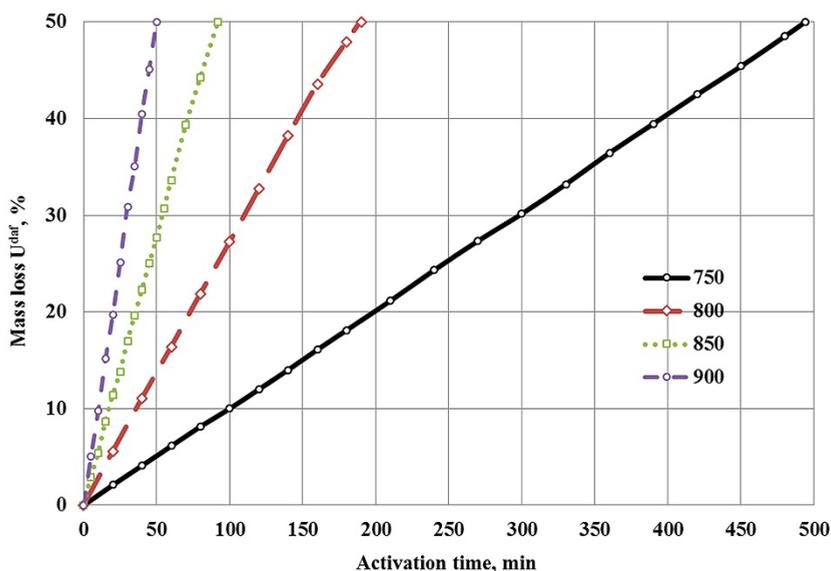
### Reactivity of the carbonized beech wood char and porous structure modification in the steam activation process

Isothermal conditions at the temperatures of 750, 800, 850 and 900 °C were applied for the KB<sub>750</sub>, KB<sub>800</sub>, KB<sub>850</sub>, and KB<sub>900</sub> samples for the partial gasification with the use of steam as the activation agent. Preparation of active carbons was conducted up to the organic mass loss at the level of 50 % (calculated to the *daf* basis). Materials obtained in the physical activation were designated as the following: KB<sub>750ac</sub>, KB<sub>800ac</sub>, KB<sub>850ac</sub>, KB<sub>900ac</sub>.

In Figure 2, reactivity of the carbonized samples in the steam activation process are presented. Table 4 contains some parameters that describe the course of these processes. The curves of isothermal steam activation processes shown in Figure 2 are similar to a straight line, which indicates a constant and similar speed of reaction of carbon material with steam in time. Increase in the temperature accelerates the rate of activation heterogeneous reaction over 9-times; the time to reach 50% weight loss reduced from 492 minutes (KB<sub>750ac</sub> activated at a temperature of 750 °C) to 53 min (KB<sub>900ac</sub> sample activated at a temperature of 900 °C –Table 4).

**Table 4.** Steam activation parameters up to 50% of the organic mass loss of the carbonized beech wood chars

Sample	Activation temperature, °C	Activation time, min	Average reactivity $R^{daf}$ , $g \cdot g^{-1} \cdot h^{-1}$
KB <sub>750ac</sub>	750	492	0.061
KB <sub>800ac</sub>	800	185	0.162
KB <sub>850ac</sub>	850	91	0.330
KB <sub>900ac</sub>	900	53	0.566



**Figure 2.** Changes in the samples mass loss during the steam activation of carbonized beech wood char

Characteristic parameters describing the porous structure of activated carbons, which are calculated on the basis of carbon dioxide and benzene adsorption isotherms, are summarized in Table 5. The volume and surface area of the micropores ( $V_{\text{MIK CO}_2}$  and  $S_{\text{CO}_2}$ ) available for carbon dioxide compound and inaccessible for benzene were as follows:

<b>KB<sub>750ac</sub></b>	$V_{\text{MIK CO}_2} = 0.177 \text{ cm}^3 \cdot \text{g}^{-1}$	$S_{\text{CO}_2} = 466 \text{ m}^2 \cdot \text{g}^{-1}$
<b>KB<sub>800ac</sub></b>	$V_{\text{MIK CO}_2} = 0.172 \text{ cm}^3 \cdot \text{g}^{-1}$	$S_{\text{CO}_2} = 453 \text{ m}^2 \cdot \text{g}^{-1}$
<b>KB<sub>850ac</sub></b>	$V_{\text{MIK CO}_2} = 0.171 \text{ cm}^3 \cdot \text{g}^{-1}$	$S_{\text{CO}_2} = 450 \text{ m}^2 \cdot \text{g}^{-1}$
<b>KB<sub>900ac</sub></b>	$V_{\text{MIK CO}_2} = 0.173 \text{ cm}^3 \cdot \text{g}^{-1}$	$S_{\text{CO}_2} = 461 \text{ m}^2 \cdot \text{g}^{-1}$

Compared to the chars, the main result of the steam activation process was almost two times higher pore volume and a surface area in the obtained active carbons (Table 3, Table 5). The average pore diameter of the active carbon mesopores was determined in the range of 3.54 – 4.07 nm. Conversely, compared to the carbonization, increase in the activation temperature corresponded to an increase in the average pore diameter of the mesopores. The obtained active carbons are characterized with a relatively large micropore volume ( $0.34 - 0.35 \text{ cm}^3 \cdot \text{g}^{-1}$ ) and well-developed system of mesopores ( $0.20 - 0.23 \text{ cm}^3 \cdot \text{g}^{-1}$  – Table 5). Considering total surfaces of micro- and mesopores, which vary in the range of  $1044 - 1092 \text{ m}^2 \cdot \text{g}^{-1}$  (Table 5), the obtained carbon materials may be considered as high-performance adsorbents.  $S_{\text{BET}}$  level of the active carbons obtained in the steam activation of biomass by other authors can be noted as similar or higher, eg. ACs from giant knotweed,  $S_{\text{BET}}$  below  $800 \text{ m}^2 \cdot \text{g}^{-1}$  [26], millet stalks ACs:  $1324 \text{ m}^2 \cdot \text{g}^{-1}$ , rice husk ACs:  $384 \text{ m}^2 \cdot \text{g}^{-1}$ , cashew shells ACs:  $858 \text{ m}^2 \cdot \text{g}^{-1}$  [31], coconut shell based ACs:  $1499 \text{ m}^2 \cdot \text{g}^{-1}$ , wood-based carbon  $970 \text{ m}^2 \cdot \text{g}^{-1}$  [32].

**Table 5.** Porous structure of the activated carbons obtained in the carbonized beech wood char steam activation

Pore width	Micropores		Mesopores					Micro- & Mesopores	$S_{\text{BET}}$	
	<0.4	0.4-2	2-3	3-5	5-10	10-50	2-50			
Volume, $\text{cm}^3 \cdot \text{g}^{-1}$	KB <sub>750ac</sub>	0	0.338	0.073	0.060	0.049	0.015	0.197	0.535	-
	KB <sub>800ac</sub>	0	0.352	0.078	0.065	0.066	0.020	0.229	0.581	-
	KB <sub>850ac</sub>	0	0.341	0.066	0.057	0.055	0.031	0.209	0.550	-
	KB <sub>900ac</sub>	0	0.346	0.060	0.059	0.058	0.032	0.209	0.550	-
Surface, $\text{m}^2 \cdot \text{g}^{-1}$	KB <sub>750ac</sub>	0	933	61.4	33.2	15.2	1.5	111.3	1044	887
	KB <sub>800ac</sub>	0	971	64.6	35.7	19.3	1.8	121.4	1092	943
	KB <sub>850ac</sub>	0	942	54.7	30.8	16.7	3.2	105.4	1047	919
	KB <sub>900ac</sub>	0	956	49.5	32.7	17.3	3.3	120.8	1059	928

Increase in the activation temperature by 150 °C resulted in a significant reduction of the activation time (Table 4). A shorter contact time between the activation agent and the carbonized char did not strongly influence the porous structure of the obtained active carbons. Taking into account the development of the material surface and the amount of specific pore volumes/width (Table 5), it can be noticed that in the range of 750-900 °C the carbonized beech wood char retains a similar structure. That is why, considering the activation time for the assumed 50% of organic mass loss, the most economically preferable would be the activation temperature of 900 °C (the shortest activation time). On the other hand, considering the methane storage, the KB<sub>800ac</sub> material of the most developed surface area (943 m<sup>2</sup>·g<sup>-1</sup>) and the most developed micro-mesoporous structure (1092 m<sup>2</sup>·g<sup>-1</sup>), obtained at the activation temperature of 800 °C, is the most preferable.

### **Beech wood char as methane sorbent**

To evaluate the obtained carbons as methane storage materials, tests of methane adsorption in the sorption apparatus (Fig. 4) were conducted. The following parameters of the test were applied: the temperature of 25 °C and methane pressure in the range of 0-700 mm Hg. Based on the determined adsorption/desorption isotherms, the following parameters of the prepared carbons were calculated: the amount of methane adsorbed at the pressure of 700 mm Hg ( $a_{ad,700}$ , mmol·g<sup>-1</sup>; g·kg<sup>-1</sup>); the amount of methane remaining in the pores after desorption process at a pressure of 0 mm Hg ( $a_{des,0}$ , mmol·g<sup>-1</sup>; g·kg<sup>-1</sup>); and the amount of methane desorbed at a pressure of 0 mm Hg ( $a_{ad,700} - a_{des,0}$ , mmol·g<sup>-1</sup>; g·kg<sup>-1</sup>). All the obtained results are summarized in Table 6. The calculation of the CH<sub>4</sub> mmol·g<sup>-1</sup> per g·kg<sup>-1</sup> was prepared with the assumption of methane molar mass equal 16.043 g·mol<sup>-1</sup>.

Large quantities of methane (about 18 g·kg<sup>-1</sup>) were adsorbed on the KB surface at a pressure of 700 mmHg. Nevertheless, the reversibility of this process is not satisfactory. It can be assumed that the cause of approx. 26% loss of methane in the desorption process is the geometry of pores (closed or narrow). As expected, the carbonization caused the expansion of the preferred size pores and a greater share of the reversible adsorption. Carbonization at the temperature range of 750-900 °C resulted in a slight increase in the volume and surface micro- and mesopores (mesopores mainly of the smallest widths: 2 - 3 nm; Table 3). As is already known, these pores play the most significant role in the methane sorption process (micro- and small mesopores). A significant improvement in the properties of materials caused by carbonization process is evident in the case of reversible

storage of methane. As the temperature increases, there is an increase in reversibility in the range of 83-85 %. The positive effect of higher temperature carbonization was not observed with an amount of methane adsorbed (Table 6). One of the obtained carbonization products, KB<sub>800</sub>, is characterized with the methane adsorption at the level of ca. 19.1 g·kg<sup>-1</sup> and a process reversibility of 83 % (Table 6). It can be considered as a commercial material for methane storage. Assuming that in the gas storage adsorption a crucial role is played by the presence of only micropores and mesopores of the smallest width, the results obtained in this work do not fully confirm such findings. Active carbons obtained in the steam activation are characterized by almost a two-fold higher micropore volume (0.338-0.352 cm<sup>3</sup>·g<sup>-1</sup> - Table 5) than carbonization products (0.173-0.201 cm<sup>3</sup>·g<sup>-1</sup> - Table 3). Narrow mesopores (2-3 nm) were highly developed in the steam activation process and were also involved in the methane sorption.

**Table 6.** Methane adsorption/desorption results with a use of carbon materials

Sample	Amount of the methane				Reversibility of adsorption
	Adsorption p = 700 mm Hg		Desorption p = 0 mm Hg		
	mmol·g <sup>-1</sup>	g·kg <sup>-1</sup>	mmol·g <sup>-1</sup>	g·kg <sup>-1</sup>	%
<b>Beech wood char</b>					
KB	1.125	18.05	0.296	4.75	73.7
<b>Carbonization products</b>					
KB <sub>750</sub>	1.010	16.20	0.172	2.76	83.0
KB <sub>800</sub>	1.190	19.09	0.203	3.26	82.9
KB <sub>850</sub>	1.095	17.57	0.188	3.02	82.8
KB <sub>900</sub>	1.096	17.58	0.163	2.62	85.1
<b>Active carbons</b>					
KB <sub>750ac</sub>	1.089	17.47	0.159	2.55	85.4
KB <sub>800ac</sub>	1.102	17.68	0.174	2.79	84.2
KB <sub>850ac</sub>	1.060	17.01	0.166	2.66	84.3
KB <sub>900ac</sub>	1.042	16.72	0.099	1.59	90.5

Activated carbons are characterized with 3-4 times higher volume of 2-3 nm width mesopores (up to the value of  $0.078 \text{ cm}^3 \cdot \text{g}^{-1}$  in case of  $\text{KB}_{800\text{ac}}$  sample – Table 5) than carbonized beech wood char samples. Nevertheless, the level of methane adsorbed is similar, as in the case of carbonized samples ( $16.2\text{--}19.1 \text{ g} \cdot \text{kg}^{-1}$ ) and in the case of activated carbons ( $16.7\text{--}17.7 \text{ g} \cdot \text{kg}^{-1}$  - Table 6). These results confirm the findings of other researchers that the storage of methane in the porous material is not only controlled by the micropore volume of the material but that their geometry also plays an important role [33, 34].

## CONCLUSIONS

Commercial beech wood char was initially treated as a potential substrate for receiving the activated carbon for methane sorption.

Commercial beech wood char is characterized by low volatile matter (5.3%) and high carbon content (94.5%). The original porous structure of the material is well developed. Micro- and mesoporous volumes were determined at the level of  $0.21 \text{ cm}^3 \cdot \text{g}^{-1}$ , while  $S_{\text{BET}}$  of the material was determined at the level of  $416 \text{ m}^2 \cdot \text{g}^{-1}$ . This material meets all the basic conditions required for precursors of activated carbons. Carbonization conducted at the temperature range of  $750\text{--}900 \text{ }^\circ\text{C}$  did not significantly affect the pore system. The improvement resulted in slightly increasing pore volumes in the range of  $0.19\text{--}0.23 \text{ cm}^3 \cdot \text{g}^{-1}$ . The most favorable system for methane storage was developed in the carbonization step at a final process temperature of  $800 \text{ }^\circ\text{C}$ . The methane adsorption level was determined as  $19 \text{ g} \cdot \text{kg}^{-1}$  with 83% reversibility of the process. Steam activation of chars were characterized with  $S_{\text{BET}}$  within the range of  $1044\text{--}1092 \text{ m}^2 \cdot \text{g}^{-1}$  and micro- and mesoporous volumes within the range of  $0.54\text{--}0.58 \text{ cm}^3 \cdot \text{g}^{-1}$ . The maximum methane adsorption was determined for the sample activated at a temperature of  $800 \text{ }^\circ\text{C}$  –  $17.7 \text{ g} \cdot \text{kg}^{-1}$  with a process reversibility of 84.2%.

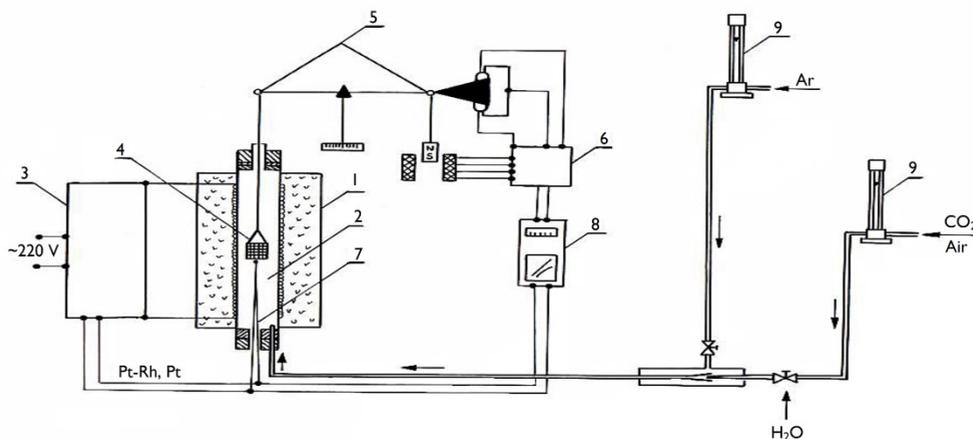
## EXPERIMENTAL SECTION

### Beech wood char and active carbons characterization

As the raw material, a commercial beech wood char (KB) with different grain sizes was used. For the following studies, a  $0.5\text{--}3.15 \text{ mm}$  sieve fraction was isolated. The following analyses were utilized with the carbon material: elemental analysis (CHNS EA 1110 CE Instruments), proximate analysis

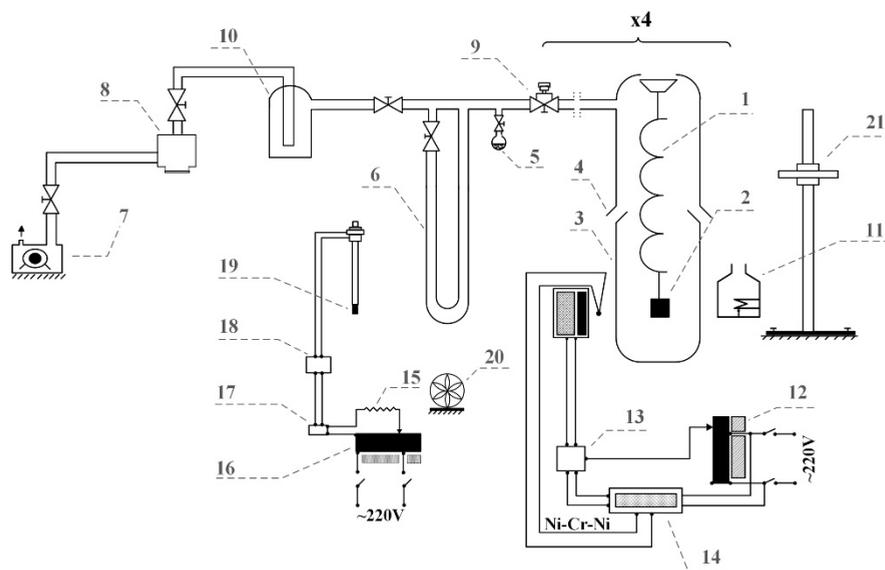
(moisture content according to: ASTM D3173–11, ash content according to: ASTM D3174–12, volatile matter according to: ASTM D3175 – 11), and thermogravimetric studies.

Thermogravimetric studies of the raw materials as well as activated carbon materials were performed using the thermogravimetric apparatus shown in Fig. 3. The apparatus consists of a tubular resistance furnace (1) with the quartz tube (2) positioned coaxially. The furnace is powered from the AC mains by a programmable temperature controller (3). 1 g of the tested sample was placed in a measuring cell (4) made of a heat-resistant net inside the quartz tube through the hole in the top stopper. The measuring cell was suspended from one of the arms of an analytical balance (5) using a heat-resistant wire (5). For automatic compensation and determination of the changes of sample weight during the test, the second arm of balance was controlled with the use of an electronic photoelectric-electromagnetic starter (6). Changes in the sample temperature were measured using a thermocouple (7) and recorded on a tape using a multi-electronic compensator (8). A determined amount of inert gas (argon) was introduced into the lower part of the furnace using rotameters (9). Argon was purified from traces of moisture and remaining oxygen before entering the furnace. The obtained thermograms showed the loss of weight expressed as a percentage plotted versus the temperature within the range of 20–900 °C with a heating rate at the level of 5 °C/min. At the final carbonization temperature of 900 °C, a char was stabilized for 30 minutes.



**Figure 3.** The thermogravimetric apparatus (TG) scheme. (1) tubular resistance furnace, (2) quartz tube, (3) programmable temperature controller, (4) measuring cell, (5) analytical balance, (6) electronic photoelectric-electromagnetic starter, (7) thermocouple, (8) multi-electronic compensator, and (9) rotameter

The porous structure of the samples was determined at the temperature of 25 °C in the sorption apparatus (Fig. 4) using the isotherms of carbon dioxide adsorption in the pressure range of the 0 - 700 mm Hg and the adsorption/desorption isotherms of C<sub>6</sub>H<sub>6</sub> in the relative pressure range of  $p/p_0$  0-1.



**Figure 4.** Sorption apparatus scheme. (1) suspended in the glass tube McBain – Bakr weight equipment in the form of a quartz spring; (2) quartz flask; (3)(4) glass tubes; (5) adsorbate container; (6) mercury manometer measuring 0-1013 kPa (0-760 mmHg); (7) rotary oil pump; (8) diffusion vacuum pump; (9) vacuum gauge; (10) freezer; (11) electric resistance furnace; (12) autotransformer; (13) (14) thermocouples connected to the thermoregulatory system; (15) heat exchanger; (16) thermoregulatory system; (17) contactor; (18) electrical contact; (19) contact thermometer; (20) fan; (21) cathetometer

For the interpretation of the CO<sub>2</sub> adsorption isotherms, the theory and equation of Dubinin – Radushkevich (DR) was used [35]. For the DR equation calculations, coefficient  $\beta$  affinity was adopted at the level of 0.37. Using the DR equation, micropore volumes available for carbon dioxide compounds was calculated assuming that each molecule of CO<sub>2</sub> at the temperature of 25 °C is laid flat on the surface of micropores and covers an area of 0.185 nm<sup>2</sup> [36].

Brunauer, Emmet, and Teller theory (BET) was used for the interpretation of benzene adsorption isotherm obtained at the relative pressure  $p/p_0 = 0.01 - 0.30$  [37]. The BET method was used for the calculation of the

material surface ( $S_{\text{BET}}$ ). For the calculations, it was assumed that the surface of the benzene molecule at 25 °C occupies 0.41 nm<sup>2</sup> [38]. Based on the benzene desorption curve in the range of the relative pressure  $p/p_0 = 0.96 - 0.175$ , the mesopores volume ( $V_{\text{MES}}$ ) and mesopores size distribution ( $S_{\text{MES}}$ ) as a function of width (2-3; 3-5; 5-10 and 10-50 nm) was calculated. The calculations were made according to the Pierce method [39] modified by Orr and Dallavalle [40], considering the amendments on the thickness of the adsorbed layer [41]. The calculations assumed that the pores have the shape of a slot. The average mesopores diameter ( $d_{\text{MES}}$ ) was calculated from the following equation:  $d_{\text{MES}} = 2V_{\text{MES}} : S_{\text{MES}}$ .

Micropore volumes available for benzene vapors ( $V_{\text{MIK}}$ ) were calculated as the difference of volume of benzene adsorbed at  $p/p_0 = 0.96$  (Gurvich volume  $V_{0.96}$ ) [42] and a previously calculated volume of mesopores:  $V_{\text{MIK}} = V_{0.96} - V_{\text{MES}}$  [cm<sup>3</sup>·g<sup>-1</sup>]. The micropore volumes of the widths below 0.4 nm (sub-micropores,  $V_{\text{SUB}}$ ) available to smaller molecules of carbon dioxide and inaccessible to larger molecules of benzene was calculated. Sub-micropore volumes were calculated as the difference of the micropore volume calculated from the CO<sub>2</sub> adsorption curve ( $V_{\text{MIK CO}_2}$ ) and the micropore volume calculated from the benzene adsorption curve ( $V_{\text{MIK}}$ ) at  $p/p_0 = 0.96$ .  $V_{\text{SUB}} = V_{\text{MIK CO}_2} - V_{\text{MIK}}$ .

### Carbonization and physical activation

Carbonization was operated in the apparatus described in Figure 3 under an argon atmosphere with a flow rate of 30 dm<sup>3</sup>·h<sup>-1</sup> and heating rate of 10 °C·min<sup>-1</sup> up to the final temperature of 750, 800, 850 and 900 °C. At the final carbonization temperature, the experiment was held at constant (final) temperature for 30 min till the carbonization was completed. The obtained materials were cooled in the argon flow up to room temperature. Carbonized materials were designated as the following: KB<sub>750</sub>, KB<sub>800</sub>, KB<sub>850</sub>, KB<sub>900</sub>.

Carbonized materials were used for the preparation of the active carbon. Physical activation with steam was conducted at temperatures of 750, 800, 850 and 900 °C up to the 50% organic mass loss of the material (calculated to *daf* – dry, ash free). After reaching the scheduled weight loss, the activating agent was replaced with argon, and the sample was cooled to room temperature. Active carbons were designated as KB<sub>750ac</sub>, KB<sub>800ac</sub>, KB<sub>850ac</sub>, and KB<sub>900ac</sub>. The average reactivity of the samples ( $R^{\text{daf}}$ ) was calculated by dividing the weight of the gasified char ( $\Delta m^{\text{daf}}$ ) by the product of the mass of char applied for the process ( $m^{\text{daf}}$ ) and the activation time expressed in hours (t):

$$R^{\text{daf}} = \frac{\Delta m^{\text{daf}}}{m^{\text{daf}} t}, [g \cdot g^{-1} \cdot h^{-1}].$$

Both carbonization and physical activation of the beech wood were conducted in the TG apparatus (Fig. 3).

### Determination of methane sorption capacity

To evaluate the obtained carbons as methane storage materials, tests of methane adsorption and desorption were conducted in the sorption apparatus presented in Fig. 4. The following parameters of the test were applied: temperature of 25 °C and methane pressure range 0-700 mm Hg. Based on the determined adsorption and desorption isotherms of methane, the following parameters of the prepared carbons were calculated:

- $a_{ad,700}$  - the amount of methane adsorbed at the pressure of 700 mm Hg ( $\text{mmol}\cdot\text{g}^{-1}$ ;  $\text{g}\cdot\text{kg}^{-1}$ );
- $a_{des,0}$  - the amount of methane remaining in the pores after desorption process, at the pressure of 0 mm Hg ( $\text{mmol}\cdot\text{g}^{-1}$ ;  $\text{g}\cdot\text{kg}^{-1}$ );
- $a_{ad,700} - a_{des,0}$  - the amount of methane desorbed at a pressure of 0 mm Hg ( $\text{mmol}\cdot\text{g}^{-1}$ ;  $\text{g}\cdot\text{kg}^{-1}$ ).

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### REFERENCES

1. D. Lozano-Castello, J. Alcaniz-Monge, M.A.M de la Casa-Lillo, D. Cazorla-Amoros, A. Linares-Solano, *Fuel*, **2002**, *81*, 1777.
2. S.E. Manahan, "Environmental Science and Technology: A Sustainable Approach to Green Science and Technology", Second Edition, Taylor & Francis, Boca Raton/USA, **2006**, chapter **18**.
3. T. Düren, L. Sarkisov, O.M. Yaghi, R.Q. Snurr, *Langmuir*, **2004**, *20*, 2683.
4. D. Bazer-Bachi, L. Assie, V. Lecocq, B. Harbuzaru, V. Falk, V., *Power Technology*, **2014**, *255*, 52.
5. M.I. Nandasiri, S.R. Jambovane, B.P. McGrail, H.T. Schaefer, S.K. Nune, *Coordination Chemistry Reviews*, **2016**, *311*, 38.
6. J.A. Mason, M. Veenstra, J.R. Long, *Chemical Science*, **2014**, *5*, 32.
7. Y. He, W. Zhou, G. Qian, B. Chen, *Chemical Society Reviews*, **2014**, *43*, 5657.

8. J. Shen, J. Sulkowski, M. Beckner, A. Dailly, *Microporous and Mesoporous Materials*, **2015**, 212, 80.
9. O.M. Yanghi, M. O'Keefe, N.W. Ockwig, H.K. Chae, M. Eddaoudi, J. Kim, *Nature*, **2003**, 423, 705.
10. H. Furukawa, O.M. Yaghi, *Journal of the American Chemical Society*, **2009**, 131, 8875.
11. J.H. Guo, H. Zhang, Y. Miyamoto, *Physical Chemistry Chemical Physics*, **2013**, 15, 8199.
12. H.M. El-Kaderi, J.R. Hunt, J.L. Mendoza-Cortes, A.P. Cote, R.E. Taylor, M. O'Keefe, O.M. Yanhi, *Science*, **2007**, 316, 268.
13. J.H. Hu, J.F. Zhao, T.Y. Yan, *Journal of the Physical Chemistry*, **2015**, 119 (4), 2010.
14. J.L. Mendoza-Cortes, T.A. Pascal, W.A. Goddard III, *Journal of the Physical Chemistry A*, **2011**, 115, 13852.
15. J. Zhao, T. Yan, *RSC Advances*, **2014**, 4, 15542.
16. X.H. Zhang, Z.Q. Song, X.L. Zhou, C.L. Li, L.Y. Xu, *Energy Sources, Part A: Recovery, Utilization, and Environmental Effects*, **2016**, 38(2), 202.
17. T.J. Grey, K.P. Travis, J.D. Gale, D. Nicholson, *Microporous and Mesoporous Materials*, **2001**, 48, 203.
18. R. Hernandez-Huesca, L. Diaz, G. Aguilar-Armenta, *Separation and Purification Technology*, **1999**, 15, 163.
19. M. Sakizci, O.L. Tanriverdi, *Turkish Journal of Chemistry*, **2015**, 39 (5), 970.
20. M.E. Casco, M. Martinez-Escandell, K. Kaneko, J. Silvestre-Albero, F. Rodriguez-Reinoso, *Carbon*, **2015**, 93, 11.
21. D. Lozano-Castello, D. Cazorla-Amoros, A. Linares-Solano, D.F. Quinn, *Carbon*, **2002**, 40, 989.
22. J. Sreńscek-Nazzal, W. Kamińska, B. Michalkiewicz, Z.C. Koren, *Industrial Crops and Products*, **2013**, 47, 153.
23. J. Alcaniz-Monge, M.A. De la Casa-Lillo, D. Cazorla-Amoros, A. Linares-Solano, *Carbon*, **1997**, 35 (2), 291.
24. H.M.B.M. Herath, A.A.P. De Alwis, MERCon 2015 - Moratuwa Engineering Research Conference, article number: 7112310, **2015**, available online: <http://ieeexplore.ieee.org/stamp/stamp.jsp?tp=&arnumber=7112310>, Accessed 11 March 2018
25. J. Sun, M.J. Rood, M. Rostam-Abadi, A.A. Lizzio, *Gas Separation & Purification*, **1996**, 10 (2), 91.
26. H. Fałtynowicz, J. Kaczmarczyk, M. Kułazyński, *Open Chemistry*, **2015**, 13, 1150.
27. D.C.S. Azevedo, J.C.S. Araujo, M. Bastos-Neto, E.B. Torres, E.F. Jaguaribe, C.L. Cavalcante, *Microporous and Mesoporous Materials*, **2007**, 100, 361.
28. S. Meyer, B. Glaser, P. Quicker, *Environmental Science & Technology*, **2011**, 45, 9473.
29. K. Zeng, D.P. Minh, D. Gauthier, E. Weiss-Hortala, A. Nzihou, D. Flamant, *Bioresource Technology*, **2015**, 182, 114.
30. C. Guizani, F.J. Escudero Sanz, M. Jeguirim, R. Gadiou, S. Salvador, *Fuel Processing Technology*, **2015**, 138, 687.

31. M.M. Dieme, A. Villot, C. Gerente, Y. Andres, S.N. Fiop, C.K. Diawara, *Environmental Technology*, **2017**, 38 (3), 353.
32. E. Lorenc-Grabowska, *Adsorption*, **2016**, 22, 599.
33. K. Sapag, A. Vallone, G.A. Blanco, C. Solar, "Adsorption of Methane in Porous Materials as the Basis for the Storage of Natural Gas, Natural Gas", InTech, Primož Potocnik (Ed.), **2010**, Available from:  
<http://www.intechopen.com/books/natural-gas/adsorption-of-methane-in-porous-materials-as-the-basis-for-the-storage-of-natural-gas>
34. K. Wu, Z. Chen, X. Li, X. Dong, *Scientific Reports*, **2016**, 6, 33461.
35. M.M. Dubinin, *Carbon*, **1989**, 27, 457.
36. Y. Toda, M. Hatami, Y. Toyoda, Y. Yoshida, H. Honda, *Fuel*, **1971**, 50, 187.
37. S. Brunauer, P.H. Emmet, E. Teller, *Journal of the American Chemical Society*, **1938**, 60, 309.
38. T. Siemienińska, K. Tomków, J. Kaczmarczyk, A. Albinia, E. Broniek, A. Jankowska, Y. Grillet, M. François, *Studies in surface science and catalysis*, **1994**, 87, 695.
39. C. Pierce, *Journal of Physical Chemistry*, **1953**, 57, 149.
40. C. Orr, J.M. DallaValle, "Fine particle measurement: Size, Surface, and Pore Volume", The McMillan Company, New York, **1959**, chapter 8.
41. S.J. Gregg, K.S. Sing, "Adsorption, Surface Area and Porosity", Academic Press London - New York, **1982**, chapter 4.
42. A. Filozof-Nieścior, J. Kaczmarczyk, T. Siemienińska, Conference on Catalysis and Adsorption in Fuels Processing and Environmental Protection, Kudowa, pp. 69 – 74 (2002).



## RADIOCARBON DATING OF THE OLD ASH OF AITON, ROMANIA

ADRIAN PATRUT<sup>a,\*</sup>, ROXANA T. PATRUT<sup>a,b</sup>, LASZLO RAKOSY<sup>b</sup>,  
ILEANA-ANDREEA RAȚIU<sup>a</sup>, DANIEL A. LOWY<sup>c</sup>,  
JENŐ BODIS<sup>a</sup>, KARL F. VON REDEN<sup>d</sup>

**ABSTRACT.** The article reports the AMS (accelerator mass spectrometry) radiocarbon investigation results of the old common ash of Aiton. Five wood samples were collected from the trunk of the tree. The deepest ends of the samples were analysed by AMS radiocarbon. We found radiocarbon dates between  $165 \pm 20$  BP and  $240 \pm 18$  BP, which correspond to calibrated ages of 230 – 360 years. These results, combined with a ring counting estimate, indicate an age of  $330 \pm 30$  years for the ash of Aiton. By this value, the ash of Aiton becomes the oldest known common ash with accurate dating results.

**Keywords:** AMS radiocarbon dating, *Fraxinus excelsior*, dendrochronology, age determination.

### INTRODUCTION

The common ash, European ash or simply ash (*Fraxinus excelsior* L.) is a deciduous flowering tree that belongs to the Oleaceae family. The ash is native throughout mainland Europe occurring in 64% of European territories; its natural range largely coincides with that of the pedunculate oak. It is typically 20-30 m tall, exceptionally reaching 40-50 m, and has a diameter up to 2 m [1-3].

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<sup>a</sup> Babeş-Bolyai University, Faculty of Chemistry and Chemical Engineering, 11 Arany Janos, RO-400028, Cluj-Napoca, Romania.

<sup>b</sup> Babeş-Bolyai University, Faculty of Biology and Geology, 44 Republicii, RO-400015, Cluj-Napoca, Romania.

<sup>c</sup> Dept. of Science and Innovation, Valor Hungariae, 4 Nagysándor József, 1054 Budapest, Hungary.

<sup>d</sup> NOSAMS Facility, Dept. of Geology & Geophysics, Woods Hole Oceanographic Institution, Woods Hole, MA 02543, U.S.A.

\* Corresponding author: [apatrut@gmail.com](mailto:apatrut@gmail.com)



**Figure 1. a.** General view of the ash of Aiton.  
**b.** Detail of sample 3, showing obvious and faint growth rings.

In 2005, we started an in-depth research to elucidate several controversial or poorly understood aspects concerning the architecture, growth and age of the African baobab. The research is based on our new approach which enables to investigate and date standing live specimens, as well. This approach consists of AMS radiocarbon dating of tiny wood samples collected from different areas of such trees [4-9]. We extended our research by starting to date individuals which belong to other angiosperm species, including trees from Romania.

Romania hosts several monumental trees which have reached large dimensions and old ages. We investigated and dated by AMS radiocarbon some remains of the historic oak of Tebea [10], the large pedunculate oak of Cajvana [11], as well as the old black poplar of Mocod and the big grey poplar of Rafaila [12]. In each case we found very old ages for these species.

Here we disclose the AMS radiocarbon dating results of another representative Romanian tree, namely the old ash of Aiton.

## RESULTS AND DISCUSSION

*The old ash of Aiton and its area.* The old ash is located in the garden of a privately-owned house (No. 420) in the village and commune Aiton, Cluj county, at 18 km SE of Cluj-Napoca, in the northern part of Romania. Its GPS coordinates are 46°40.949' N, 023°44.056' E and the altitude is 636 m. The mean annual rainfall in the area is around 650 mm.

The ash of Aiton has today a maximum height of 29.2 m, the circumference at breast height (cbh; at 1.30 m above mean ground level) is 6.27 m, while the circumference on slope at ground level is 11.65 m. The overall wood volume is 32 m<sup>3</sup>, out of which the trunk/stem has a volume of 26 m<sup>3</sup> (**Figure 1a**).

The quasi-cylindrical trunk forks at a height of 9.6–10.0 m into four large primary branches, with diameters up to 0.8 m. Two other very large branches fell in 2005 and one in 2015 during storms, while other two branches burned down in 1994 in a big fire, after the tree was hit by lightning. The fire was only extinguished after 8 hours by firefighters; the fire damage reduced the canopy to the half and decreased the height of the ash. Now, the horizontal dimensions of the remaining canopy are 27.6 (NS) x 23.6 (WE) m.

*Wood samples.* A number of five wood samples were extracted from the outer part of the tree around the circumference. The five samples (labelled 1 to 5) have lengths between 0.61 and 0.75 m. The heights of the

sampling points were between 1.70 and 2.01 m. The deepest end of each sample (marked as a) was extracted for radiocarbon dating.

*AMS results and calibrated ages.* Radiocarbon dates of the deepest segments extracted from the five samples are listed in Table 1. Radiocarbon dates and errors were rounded to the nearest year. The radiocarbon dates are expressed in  $^{14}\text{C}$  yr BP (radiocarbon years before present, i.e., before the reference year AD 1950).

Calibrated (cal) ages, expressed in calendar years, are also shown in Table 1. The 1- $\sigma$  probability distribution was selected to derive calibrated age ranges. For all sample segments, the 1- $\sigma$  distribution corresponds to two, three or four ranges of calendar years. For these segments, the confidence interval of one range is considerably greater than that of the others; therefore, it was selected as the cal AD range of the segment for the purpose of this discussion. For obtaining single calendar age values of sample segments, we derived a mean calendar age of each segment from the selected range (marked in bold). Calendar ages of segments represent the difference between AD 2018 and the mean value of the selected range, with the corresponding error. Calendar ages and errors were rounded to the nearest 5 yr.

**Table 1.** AMS radiocarbon dating results and calibrated calendar ages of samples/segments collected from the Aiton ash.

Sample (Segment) code	Depth <sup>1</sup> [height <sup>2</sup> ] ( $10^{-2}$ m)	Radiocarbon date [error] ( $^{14}\text{C}$ yr BP)	Cal AD ranges 1- $\sigma$ [confidence interval]	Sample age [error] (cal yr)
1a	64 [195]	170 [ $\pm$ 17]	1670-1682 [12.4%] <b>1736-1780 [42.8%]</b> 1798-1804 [5.6%] 1936-1944 [7.4%]	260 [ $\pm$ 20]
2a	71 [185]	229 [ $\pm$ 19]	<b>1653-1666 [40.2%]</b> 1784-1794 [28.0%]	360 [ $\pm$ 5]
3a	75 [201]	240 [ $\pm$ 18]	<b>1647-1665 [55.1%]</b> 1786-1794 [13.1%]	360 [ $\pm$ 10]
4a	62 [170]	205 [ $\pm$ 21]	1659-1675 [18.9%] <b>1777-1799 [32.1%]</b> 1941-... [17.3%]	230 [ $\pm$ 10]
5a	61 [177]	165 [ $\pm$ 20]	1673-1682 [8.2%] <b>1735-1778 [42.9%]</b> 1799-1806 [6.3%] 1930-1942 [10.7%]	260 [ $\pm$ 20]

<sup>1</sup> Depth in the wood from the sampling point.

<sup>2</sup> Height above ground level.

*Dating results of samples (segments).* The deepest part/end of the collected samples have radiocarbon dates between  $165 \pm 20$  BP and  $240 \pm 18$  BP. These values correspond to calibrated ages between  $230 \pm 10$  and  $360 \pm 10$  yr.

One should mention that the calibration, i.e., the conversion of radiocarbon dates into calendar ages, can be discussed in particular cases. Thus, the calibration of wood samples which have calendar/dendro ages in the time frame AD 1640 – 1950 shows some uncertainties. In such cases, the calibration of one radiocarbon date may result in up to five or six calendar age ranges. This is due to the so-called “Suess effect”, which expresses the high variation of the atmospheric radiocarbon concentration produced by the combustion of fossil fuels [13,14].

*Ring counting.* On the collected wood samples, the growth/annual rings are obvious in certain areas, while in other areas they are very faint (**Figure 1b**). Therefore, an accurate ring counting is not possible. The longest sample 3 had a length of 0.75 m and was collected at a height of 2.01 m. At this height, the circumference of the trunk is 5.50 m. This value corresponds to a diameter of 1.72 m without bark. The distance from the sample end to the centre/theoretical pith is 0.11 m.

We estimate that sample 3 contains around 300 – 320 rings. By considering that the ash grows faster in the first 40 years [3], the number of missing rings from the sample end to the pith must be close to 20. Therefore, this estimate for the ash of Aiton suggests a number of 320 – 340 growth rings.

We investigated and measured the tree in 2009, 2010, 2013, 2015 and 2018. The mean growth rate over this period, namely the increase in radius, was of  $1.9 \times 10^{-3} \text{ m yr}^{-1}$  ( $0.19 \text{ cm yr}^{-1}$ ) at the height of 2 m.

*Age of the ash of Aiton.* The radiocarbon dating results and the ring counting estimate indicate an age of 300 – 360 yr, i.e.,  $330 \pm 30$  yr for the investigated ash. By this value, the ash of Aiton becomes the oldest known common ash with accurate dating results and certainly one of the oldest common ash in Europe.

## CONCLUSIONS

Our research discloses the AMS radiocarbon dating results of the old common ash of Aiton. The main aim of the research was to determine the age of tree. Five wood samples were collected from its trunk. The deepest

ends of each sample were processed and dated by AMS radiocarbon. The five segments have radiocarbon dates between  $165 \pm 20$  BP and  $240 \pm 18$  BP, which correspond to calibrated ages between  $230 \pm 10$  and  $360 \pm 10$  yr. These results, combined with a ring counting estimate, indicate an age of  $330 \pm 30$  yr for the ash of Aiton which has started growing around the year 1690.

## EXPERIMENTAL SECTION

*Metric measurements.* The height of the tree was measured using a Bosch DLE 70 Professional laser rangefinder. The circumference at different heights was measured with graduated tapes. The volume was determined from laser measurements of the trunk and of large branches at different heights.

*Sample collection.* The five wood samples were collected with a Hagl6f CH 800 increment borer (80 cm length, 0.54 cm inner diameter). A number of five tiny pieces/segments of the length of 0.1 cm were extracted from the deepest end of each wood sample. The segments were processed and investigated by AMS radiocarbon dating.

*Sample preparation.* The standard acid-base-acid pretreatment method was used for removing soluble and mobile organic components. The pretreated samples were combusted to CO<sub>2</sub> by using the closed tube combustion method. Next, CO<sub>2</sub> was reduced to graphite on iron catalyst, under hydrogen atmosphere [15]. Finally, the resulting graphite samples were analysed by AMS.

*AMS measurements.* AMS radiocarbon measurements were performed at the NOSAMS Facility of the Woods Hole Oceanographic Institution (Woods Hole, MA, U.S.A.) by using the Pelletron® Tandem 500 kV AMS system. The obtained fraction modern values, corrected for isotope fractionation with the normalized  $\delta^{13}\text{C}$  value of  $-25 \text{ ‰}$ , were ultimately converted to a radiocarbon date.

*Calibration.* Radiocarbon dates were calibrated and converted into calendar ages with the OxCal v4.3 for Windows [16], by using the IntCal13 atmospheric data set [17].

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## REFERENCES

1. D. Dobrowolska, S. Hein, A. Oosterbaan, S. Wagner, J. Clark, J.P. Skovsgaard, *Forestry*, **2011**, *84*(2), 133.
2. P.A. Thomas, *Journal of Ecology*, **2016**, *104*, 1158.
3. K. Giagli, J. Baar, M. Fajstavr, V. Gryc, H. Vavrcik, *BioResources*, **2018**, *13*(1), 804.
4. A. Patrut, K.F. von Reden, D.A. Lowy, A.H. Alberts, J.W. Pohlman, R. Wittmann, D. Gerlach, L. Xu, C. Mitchell, *Tree Physiology*, **2007**, *27*, 1569.
5. A. Patrut, K.F. von Reden, R. Van Pelt, D.H. Mayne, D.A. Lowy, D. Margineanu, *Annals of Forest Science*, **2011**, *68*, 993.
6. A. Patrut, S. Woodborne, K.F. von Reden, G. Hall, M. Hofmeyr, D. Lowy, R.T. Patrut, *PLoS ONE*, **2015**, *10*(1), e0117193, doi: 10.1371/journal.pone.0117193.
7. A. Patrut, L. Rakosy, R.T. Patrut, I.A. Ratiu, E. Forizs, D.A. Lowy, D. Margineanu, K.F. von Reden, *Studia UBB Chemia*, **2016**, *LXI*, *4*, 7-20.
8. A. Patrut, S. Woodborne, K.F. von Reden, G. Hall, R.T. Patrut, L. Rakosy, P. Danthu, J-M. Leong Pock-Tsy, D.A. Lowy, D. Margineanu, *Radiocarbon*, **2017**, *59*(2), 435.
9. A. Patrut, S. Woodborne, R.T. Patrut, L. Rakosy, D.A. Lowy, G. Hall, K.F. von Reden, *Nature Plants*, **2018**, *4*, 423.
10. A. Patrut, K.F. von Reden, D.A. Lowy, S. Pasca, L. Kekedy-Nagy, I. Sovago, *Studia UBB Chemia*, **2010**, *55*, *1*, 113.
11. A. Patrut, K.F. von Reden, V. Savu, D.A. Lowy, R. Mitea, I. Barbul, *Studia UBB Chemia*, **2011**, *56*, *1*, 145.
12. A. Patrut, K.F. von Reden, D.A. Lowy, R.T. Patrut, D.L. Vaida, D. Margineanu, *Nuclear Instruments and Methods in Physics Research Section B*, **2013**, *294*, 616.
13. P.P. Tans, A.F.M. de Jong, W.G. Mook, *Nature*, **1979**, *280*, 826.

14. M. Worbes, *Dendrochronologia*, **2002**, 20 (1-2), 217-231.
15. A. Patrut, S. Woodborne, R.T. Patrut, L. Rakosy, G. Hall, I.A. Ratiu, K.F. von Reden, *Studia UBB Chemia*, **2017**, LXII, 2, 347.
16. C. Bronk Ramsey, *Radiocarbon*, **2009**, 51, 337.
17. P.J. Reimer, E. Bard, A. Bayliss, J.W. Beck, P.G. Blackwell, C. Bronk Ramsey, C.E. Buck, H. Cheng, R. Lawrence Edwards, M. Friedrich, P.M. Grootes, T.P. Guilderson, H. Hafliðason, I. Hajdas, C. Hatte, T.J. Heaton, D.L. Hoffman, A.G. Hogg, K.A. Hughen, K.F. Kaiser, B. Kromer, S.W. Manning, M. Niu, R.W. Reimer, D.A. Richards, E.M. Scott, J.R. Southon, R.A. Staff, C.S.M. Turney, J. van der Plicht, *Radiocarbon*, **2013**, 55(4), 1869.

## MICRO AND NANO ORGANIZATION OF ATMOSPHERIC PARTICULATE MATTER IN GRIGORESCU DISTRICT OF CLUJ-NAPOCA

IOAN PETEAN<sup>a</sup>, GERTRUD ALEXANDRA PALTINEAN<sup>a\*</sup>,  
AURORA MOCANU<sup>a</sup>, DANA FLORINA MUNTEAN<sup>b</sup>, LIANA MURESAN<sup>b</sup>,  
GEORGE ARGHIR<sup>c</sup>, MARIA TOMOAI-COTISEL<sup>a,d</sup>

**ABSTRACT.** The paper is focused on the micro and nano organization of atmospheric particulate matter in Grigorescu district of Cluj-Napoca in April and May 2017. Floating particles (FP) collected from atmosphere have an increased amount of minerals such as: quartz, calcite, clays (e.g. kaolinite and muscovite), and traces of lepidocrocite. Their size varies from less than 1 to over 120  $\mu\text{m}$ , established by Scherer method on an XRD pattern. PM<sub>10</sub> contains mainly quartz and calcite followed by fractions of kaolinite and muscovite. PM<sub>2.5</sub> contains clay in large amount and less quartz particles. Certain nanoparticles are found among PM<sub>2.5</sub>. The obtained values were in good agreement to TEM and AFM microscopy. The diameters are 42 nm for clay and 92 nm for quartz particles. Nanostructural features are organized around fine micro particles such as PM 2.5. PM<sub>10</sub> is often found as particles around FP.

**Keywords:** floating particles, PM<sub>10</sub>, PM<sub>2.5</sub>, micro and nano organization

### INTRODUCTION

The composition and distribution of particulate matter (PM) depend on the particularities of soil, weather, and human activities. Neglected and eroded soils lead to an increased amount of mineral particles in PM [1-4].

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<sup>a</sup> Babeş-Bolyai University, Faculty of Chemistry and Chemical Engineering, Arany Janos Str., No. 11, RO-400084 Cluj-Napoca, Romania.

<sup>b</sup> Environmental Protection Agency Cluj, Dorobanţilor Str., No. 99, RO-400609, Cluj – Napoca, Romania.

<sup>c</sup> Technical University of Cluj - Napoca, Faculty of Materials and Environment Engineering, Muncii Ave., No. 103 -105, RO-400641 Cluj – Napoca, Romania.

<sup>d</sup> Academy of Romanian Scientists, 54 Splaiul Independentei, 050094, Bucharest, Romania

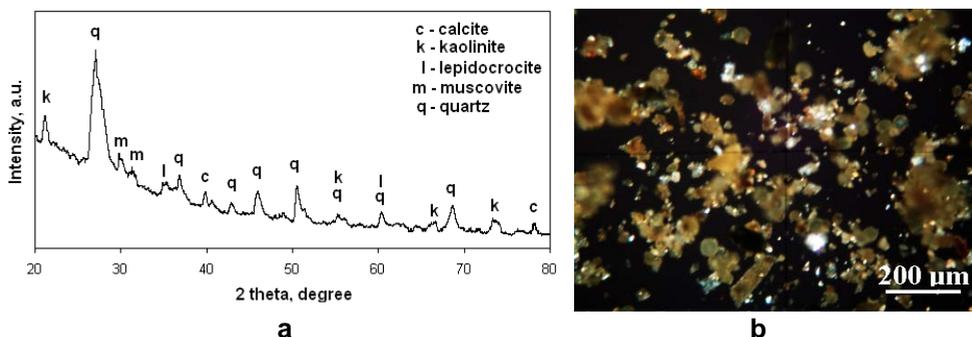
\*Corresponding author: pgertrud@gmail.com

Human activities, industrial and transport, lead to significant amount of anthropogenic PM such as acid falls, soot, and minerals resulted from industrial processes [5-9]. All these particles could be lifted into the atmosphere by weather conditions such as intense air currents related to dry environment. Humid weather as rainy days and snow during winter inhibit the spreading of particles into the atmosphere [10, 11]. Recent studies evidenced nanoparticles among small fraction of PM such as PM<sub>2.5</sub> [12, 13]. Fine micro-particles and nanoparticles dispersed into the atmosphere present high risk of acute or chronic lung affections [14-16].

Grigorescu district of Cluj-Napoca is situated far away from industrial facilities and is affected only by residential activities and transport traffic. Thus, it is expected to found among PM fractions mainly particles derived from soil decay than particles resulted from anthropogenic sources.

## RESULTS AND DISCUSSION

The first category of investigated PM was the floating particles (FP). These contain various particles lifted in atmosphere due to the weather condition. They fall back to the soil as sediment layer after their flight. XRD spectrum reveals FP sample composition, Figure 1a.



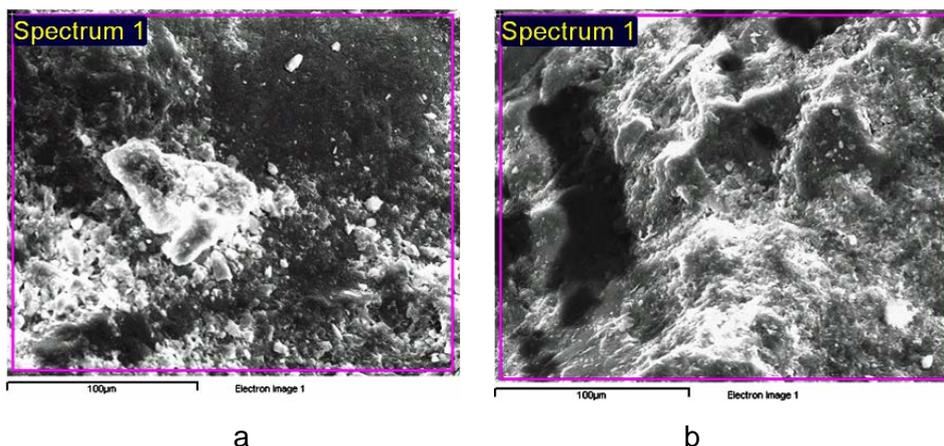
**Figure 1.** Mineralogical characterization of the FP sample collected from Grigorescu district with the Air Quality Monitoring Station: a) XRD pattern and b) cross polarized light microscopy image.

The dominant mineral is quartz, followed by calcite, clays (e.g. kaolinite, and muscovite), and traces of lepidocrocite. Cross polarized light microscopy, Figure 1b, allows determining the characteristics of particles from each mineral class data in Table 1.

**Table 1.** Components properties of FP sample

Component	Quartz	Kaolinite	Muscovite	Calcite	Lepidocrocite
Formula	SiO <sub>2</sub>	Al <sub>2</sub> Si <sub>2</sub> O <sub>5</sub> (OH) <sub>4</sub>	KAl <sub>2</sub> (AlSi <sub>3</sub> O <sub>10</sub> )(F,OH) <sub>2</sub>	CaCO <sub>3</sub>	γFeO(OH)
Particle size, μm	3 - 120	1 - 30	1 - 30	5 - 100	5 - 30
Color	Green - gray	White-blue	Pink	Yellow-brown	Reddish - brown
Shape	round	tabular	tabular	round	elongated

Data in Table 1 shows significant particle size ranges which match the category of PM<sub>10</sub> and PM<sub>2.5</sub>. These PM fractions were monitored by Automatic Air Quality Monitoring Station and representative samples were subjected to SEM imaging and SEM-EDX elemental analysis, Figure 2 and Table 2.

**Figure 2.** SEM images for particulate matters: a) PM<sub>10</sub> and b) PM<sub>2.5</sub>.

PM<sub>10</sub> sample, Figure 2a, present a mixture of well individualized particles having diameters between 2.6-10 μm surrounding a particle conglomerate of 60 μm. The coalescence tendency is proven as observed in literature [12, 13, 17]. PM<sub>2.5</sub> sample features a dense deposit of fine micro particle having diameters below 2.5 μm, Figure 2b.

**Table 2.** Elemental composition for PM10 and PM2.5, EDX analysis.

Element	PM10		PM2.5	
	weight %	atomic %	weight %	atomic %
C	11.27	16.52	32.52	43.47
O	59.40	65.37	42.53	42.67
Na	-	-	2.09	1.46
Mg	0.32	0.23	0.48	0.32
Al	1.15	0.75	5.26	3.13
Si	26.41	16.56	12.72	7.27
S	-	-	0.41	0.20
K	0.23	0.10	0.35	0.14
Ca	0.68	0.30	2.59	1.04
Fe	0.53	0.17	1.05	0.30
Total	100.00	100.00	100.00	100.00

The elemental composition for PM10 and PM2.5, given in Table 2, is in good agreement with XRD observation in Figure 1a and respectively in Figure 1b, all elements being specific to the identified minerals. Carbon is related to calcite and high amount of oxygen to large amount of silicate minerals. A trace of 0.41 wt. % of sulphur is an exception for PM2.5. Elemental distribution reveals that PM10 is rich in quartz and PM2.5 is rich in clay particles such as kaolinite and muscovite. These are flat and have small diameter being able to form compact layers as observed in Figure 2b.

PM10 and PM2.5 emissions were daily monitored during April and May 2017, measurement being performed by the Automatic Air Quality Monitoring Station from Grigorescu district. The data are compiled in Table 3, where minimum, maximum, and average values are given. A pattern was observed: the weekends with relative high humidity feature the lowest PM emission and the dry days of the middle week leads to high value of emission. However, the recorded emissions are below the limit. Grigorescu district has a friendly environment for the lungs and it worth to live there. The limit of 40  $\mu\text{g}/\text{m}^3$  was exceeded at PM10 only in a day per month where maximum values were recorded 46.59  $\mu\text{g}/\text{m}^3$  and 39.16  $\mu\text{g}/\text{m}^3$ .

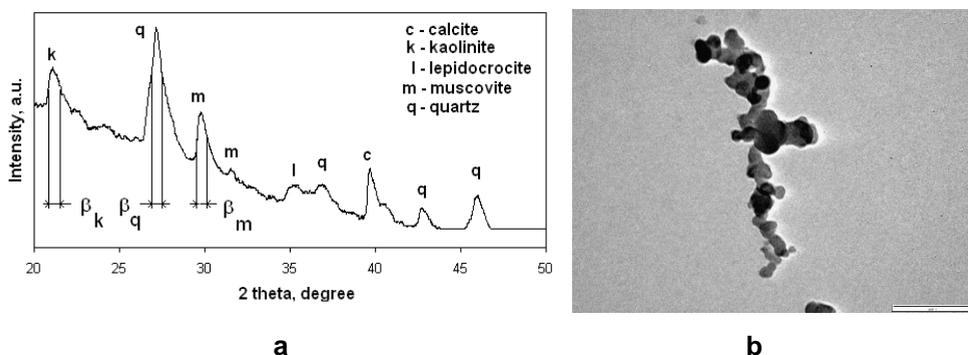
**Table 3.** PM emission level in Grigorescu district for April and May 2017

Emission type		April 2017	May 2017
PM2.5, $\mu\text{g}/\text{m}^3$	minim	7.07	12.32
	maxim	24.65	23.75
	average	11.45	18.05
PM10, $\mu\text{g}/\text{m}^3$	minim	2.92	9.79
	maxim	46.59	39.16
	average	21.14	22.61
FP, $\text{g}/\text{m}^2/\text{month}$	average	12.47	6.89

PM2.5 was dispersed in ultra-pure water, micro-particles sediment to the bottom of Berzelius glass beaker and the nanoparticles remains into dispersion. Liquid dispersed sample was subjected to TEM investigation. For XRD and AFM studies, the nanoparticles were deposited onto solid substrate (e.g. glass plate) by vertical adsorption according to the procedure [18 - 23].

XRD pattern, Figure 3a, shows less intense and broadened peaks than the ones resulted for FP sample, Figure 1. This is the effect of reducing the particles size. The sample does not have internal tensions which might affect the peak broadening, then Scherer method is suitable for particle size determination [24]. The Scherer method was applied to kaolinite, muscovite, and quartz peaks. The results are centralized in Table 4.

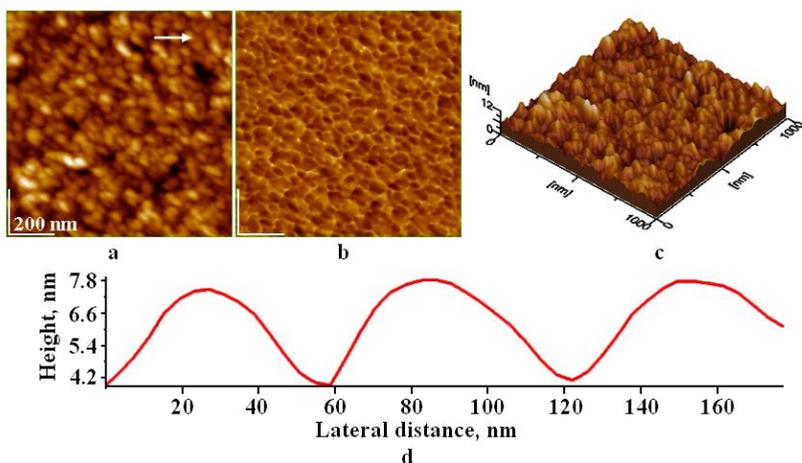
TEM image, Figure 3b, shows bigger quartz nanoparticle having a diameter of about 90 nm and clay particles ranging from 40-45 nm diameter. It is a good agreement with XRD results.



**Figure 3.** Characterization of PM2.5 thin layer from Grigorescu district collected with the Air Quality Monitoring Station:  
a) XRD pattern and b) TEM microscopy, scale bar 200 nm.

The AFM images presented in Figure 4, resulted for the thin film of nanoparticles, extracted from PM2.5 via aqueous dispersion. The topographic image shows a uniform nanoparticles film having a roughness of 1.38 nm, Figure 4a. Nanoparticles are well individualized as observed in phase image, Figure 4b. The nanoparticles appear in light brown and the border between them in light yellow.

Several profiles were taken to establish the nanoparticle diameter. It results two categories of nanoparticles: fewer around 90 nm comparable with value obtained for quartz and many having around 40-45 nm as observed in cross section presented in Figure 4d.



**Figure 4.** AFM images of PM<sub>2.5</sub> thin layer from Grigorescu district collected with the Air Quality Monitoring Station: a) topographic image, b) phase image, c) 3D image, and d) profile along white arrow in figure (a).

**Table 4.** Nano-particles diameter by several measurement techniques

Mineral	Particle diameter, nm			
	AFM	TEM	XRD	Rounded value
Kaolinite	40 ± 3	40	45.77	42
Muscovite	40 ± 5	45	41.32	42
Quartz	90 ± 6	90	95.21	92

All values obtained for nanoparticles size are centralized in Table 4. They are in good agreement for all used techniques, so we could establish a rounded value as a characteristic for each mineral involved. The clay nanoparticles are around 42 nm and quartz nanoparticles around 92 nm.

## CONCLUSIONS

The particulate matter contains mineral composition of quartz, calcite, clays (e.g. kaolinite and muscovite), and traces of lepidocrocite in Grigorescu district, of Cluj-Napoca, during April and May 2017. Their size varies from less than 1 to over 120 μm. PM<sub>10</sub> contains mainly quartz and calcite followed by

fractions of kaolinite and muscovite. PM<sub>2.5</sub> sample is abundant in clay particles and contains less quartz than PM<sub>10</sub>. Nanoparticles were found among PM<sub>2.5</sub>. XRD, TEM, and AFM allow establishing nanoparticles nature and size: kaolinite and muscovite 42 nm and quartz 92 nm. Air monitoring during April and May 2017 in Grigoresu District, Cluj – Napoca, shows that emission level is under the limit.

## EXPERIMENTAL SECTION

The experimental area is situated in Grigorescu district, of Cluj-Napoca. PM samples: FP, PM<sub>10</sub>, and PM<sub>2.5</sub> were collected using Automatic Air Quality Monitoring Station CJ-3 from Environmental Protection Agency of Cluj-Napoca, and their level of emission in the atmosphere was monitored during April and May 2017 and the results were centralized.

The X-ray diffraction analysis was performed on a DRON 3 diffractometer equipped with data acquisition module and MATMEC VI.0 soft. A monochrome Co<sub>K $\alpha$</sub>  radiation was used for all X-ray spectra. The mineral identification was effectuated using MATCH 1.0 X – ray standard data base from Crystal Impact Co.

Optical microscopy was performed on a Karl Zeiss Jena Laboval 2 microscope equipped with digital capture Kodak 10 Mpx camera. Quantitative analysis on the optical microphotographs was done using the Image J professional soft as freeware resource from National Institutes of Health, USA.

The AFM investigation was performed on a Jeol JSPM 4210 microscope in tapping mode [25-32] using NSC 15 cantilever. Samples for AFM investigation were prepared according to the appropriate data [33-38]. AFM images and cross profile were processed into the standard manner using Win SPM 2.0 soft. SEM images were achieved on a Jeol JSM 5600 LV microscope in secondary electrons imaging mode.

## REFERENCES

1. G.A. Hosu-Prack, I. Petean, G. Arghir, L.D. Bobos, M. Tomoaia-Cotisel, *Studia Univ. Babeş-Bolyai, Chemia*, **2010**, 55 (3), 93.
2. I. Petean, A. Mocanu, G. A. Paltinean, R. Tarcan, D. F. Muntean, L. Muresan, G. Arghir, M. Tomoaia-Cotisel, *Studia Univ. Babeş-Bolyai, Chemia*, **2017**, 62 (4), Tom I, 33.

3. W. De Poel, S. Pinte, J. Drnec, F. Carla, R. Felici, P. Mulder, J. Elemans, W. Van Enckevort, A.E. Rowan, E. Vlieg, *Surface Science*, **2014**, 619, 19.
4. M.S. Žbik, N.A. Raftery, R.S.C. Smart, R.L. Frost, *Applied Clay Science*, **2010**, 50, 299.
5. G.A. Hosu-Prack, I. Petean, G. Arghir, L.D. Bobos, I. Iurcut, M. Tomoaia-Cotisel, *Carpathian Journal of Earth and Environmental Sciences*, **2013**, 8 (4), 75.
6. J. Berger, B. Denby, *Atmospheric Environment*, **2011**, 45, 3692.
7. M. Giugliano, G. Lonati, P. Butelli, L. Romele, R. Tardivo, M. Grosso, *Atmospheric Environment*, **2005**, 39, 2421.
8. G. Lazar, C. Capatina, C.M. Simionescu, *Revista de Chimie (Bucharest)*, **2014**, 65 (10), 1215.
9. G. Damian, F. Damian, D. Năsui, C. Pop, C. Pricop, *Carpathian Journal of Earth and Environmental Sciences*, **2010**, 5 (1), 139.
10. D.F. Muntean, D. Ristoiu, G. Arghir, R.F. Campean, I. Petean, *Carpathian Journal of Earth and Environmental Sciences*, **2012**, 7 (3), 175.
11. D.F. Muntean, I. Ivan, L. Muresan, *Studia Univ. Babeş-Bolyai, Chemia*, **2015**, 60 (2), 207.
12. G.A. Paltinean, I. Petean, G. Arghir, D.F. Muntean, L.D. Bobos, M. Tomoaia-Cotisel, *Particulate Science and Technology*, **2016**, 34 (5), 580.
13. G.A. Paltinean, I. Petean, G. Arghir, D.F. Muntean, M. Tomoaia-Cotisel, *Revista de chimie (Bucharest)*, **2016**, 67 (6), 1118.
14. W.C. Lo, R.H. Shie, C.C. Chan, H.H. Lin, *Journal of the Formosan Medical Association*, **2017**, 116, 32.
15. Q. Xu, S. Wang, Y. Guo, C. Wang, F. Huang, X. Li, Q. Gao, L. Wu, L. Tao, J. Guo, W. Wang, X. Guo, *Environmental Pollution*, **2017**, 220, 317.
16. M. Watanabe, H. Noma, J. Kurai, H. Sano, D. Hantan, M. Ueki, H. Kitano, E. Shimizu, *Allergology International*, **2017**, 66, 52.
17. G.A. Paltinean, I. Petean, G. Arghir, D.F. Muntean, L. Muresan, M. Tomoaia-Cotisel, *Annals of the Academy of Romanian Scientists Series on Physics and Chemistry Sciences*, **2017**, 2 (2), 43.
18. O. Horovitz, G. Tomoaia, A. Mocanu, T. Yupsanis, M. Tomoaia-Cotisel, *Gold Bulletin*, **2007**, 40 (4), 295.
19. M. Tomoaia-Cotisel, A. Tomoaia-Cotisel, T. Yupsanis, G. Tomoaia, I. Balea, A. Mocanu, Cs. Racz, *Revue Roumaine de Chimie*, **2006**, 51 (12), 1181.
20. M.A. Naghiu, M. Gorea, E. Mutch, F. Kristaly, M. Tomoaia-Cotisel, *Journal of Material Science and Technology*, **2013**, 29 (7), 628.
21. G. Tomoaia, O. Soritau, M. Tomoaia-Cotisel, L.-B. Pop, A. Pop, A. Mocanu, O. Horovitz, L.D. Bobos, *Powder Technology*, **2013**, 238, 99.
22. G. Tomoaia, O. Horovitz, A. Mocanu, A. Nita, A. Avram, C.P. Racz, O. Soritau, M. Cenariu, M. Tomoaia-Cotisel, *Colloids and Surfaces B: Biointerfaces*, **2015**, 135, 726.
23. P.T. Frangopol, D.A. Cadenhead, G. Tomoaia, A. Mocanu, M. Tomoaia-Cotisel, *Revue Roumaine de Chimie*, **2015**, 60 (2-3), 265.
24. I. Petean, G. Arghir, M. Tomoaia-Cotisel, O. Horovitz, D.A. Pop, *Journal of Optoelectronics and Advanced Materials*, **2010**, 12 (10), 2119.

25. R.D. Pasca, G. Tomoaia, A. Mocanu, I. Petean, G.A. Paltinean, O. Soritau, M. Tomoaia-Cotisel, *Studia Univ. Babes-Bolyai, Chemia*, **2015**, 60 (3), 257.
26. R.D. Pasca, A. Mocanu, S.C. Cobzac, I. Petean, O. Horovitz, M. Tomoaia-Cotisel, *Particulate Science and Technology*, **2014**, 32 (2), 131.
27. G. Tomoaia, M. Tomoaia-Cotisel, L.B. Pop, A. Pop, O. Horovitz, A. Mocanu, N. Jumate, L.D. Bobos, *Revue Roumaine de Chimie*, **2011**, 56, 1039.
28. M. Tomoaia-Cotisel, A. Mocanu, *Revista de Chimie (Bucharest)*, **2008**, 59 (11), 1230.
29. G. Furtos, M.A. Naghiu, H. Declercq, M. Gorea, C. Prejmerean, O. Pana, M. Tomoaia-Cotisel, *Journal of Biomedical Materials Research Part B. Applied Biomaterials*, **2015**, 104 (7), 1290.
30. F. Goga, E. Forizs, A. Avram, A. Rotaru, A. Lucian, I. Petean, A. Mocanu, M. Tomoaia-Cotisel, *Revista de Chimie (Bucharest)*, **2017**, 68 (6), 1193.
31. I. Cojocaru, A. Tomoaia-Cotisel, A. Mocanu, T. Yupsanis, M. Tomoaia-Cotisel, *Revista de Chimie (Bucharest)*, **2017**, 68 (7), 1470.
32. O. Cadar, P.T. Frangopol, G. Tomoaia, D. Oltean, G.A. Paltinean, A. Mocanu, O. Horovitz, M. Tomoaia-Cotisel, *Studia Univ. Babes-Bolyai, Chemia*, **2017**, 62 (4), Tom I, 67.
33. I. Petean, G. Tomoaia, O. Horovitz, A. Mocanu, M. Tomoaia-Cotisel, *Journal of Optoelectronics and Advanced Materials*, **2008**, 10 (9), 2289.
34. G. Tomoaia, A. Mocanu, L.D. Bobos, L.B. Pop, O. Horovitz, M. Tomoaia-Cotisel, *Studia Univ. Babes-Bolyai, Chemia*, **2015**, 60 (3), 265.
35. G. Tomoaia, M. Tomoaia-Cotisel, A. Mocanu, O. Horovitz, L.D. Bobos, M. Crisan, I. Petean, *Journal of Optoelectronics and Advanced Materials*, **2008**, 10 (4), 961.
36. A. Mocanu, R.D. Pasca, G. Tomoaia, C. Garbo, P.T. Frangopol, O. Horovitz, M. Tomoaia-Cotisel, *International Journal of Nanomedicine*, **2013**, 8, 3867.
37. G. Tomoaia, P.T. Frangopol, O. Horovitz, L.D. Bobos, A. Mocanu, M. Tomoaia-Cotisel, *Journal of Nanoscience and Nanotechnology*, **2011**, 11 (9), 7762.
38. G. Tomoaia, L.B. Pop, I. Petean, M. Tomoaia-Cotisel, *Materiale Plastice*, **2012**, 49 (1), 48.



# MACROKINETIC INVESTIGATIONS OF DRY FLUE-GAS DESULFURIZATION PROCESS USING TGA TECHNIQUE

SIMION DRĂGAN<sup>a</sup>

**ABSTRACT.** In this paper, a kinetic study about of dry flue-gas desulfurization process was investigated. Experimental thermogravimetric measurements, employing a CAHN TG- 121 system, have been performed on the reaction of sulfur dioxide and oxygen with calcined limestone. The conversions versus time of calcined limestone, ranging in particle size from 25 to 450  $\mu\text{m}$ , were measured over the temperature range 973-1173 K and a gas rate of 0.046 m/s. The influence of kinetic parameters on the sulfation process has been tracked. A significant influence is the temperature at which the sulfation and the size of the adsorbent granules are achieved. Processing the experimental results in  $\ln k_{i,ii}-T^{-1}$  coordinates allowed the determination of the activation energy and the establishment of the corresponding mechanisms. The results,  $E_a > 42$  KJ/mol for the both indicate that the sulfation process is carried out according to the macrokinetic mass transformation model, only for small granules of the solid with  $d_p < 25$   $\mu\text{m}$  and temperatures  $T > 973$  K) are the limiting ones the process. At higher granulations of the solid particles, the activation energy values  $E_a < 42$  kJ/mol, emphasizes that limitative of the global process are the inner/outer diffusion phenomena shown that the sulfation process follows the transfer macrokinetic model.

**Keywords:** *dry desulfurization process, sulfation process, transfer and mass transformation macrokinetic model.*

## INTRODUCTION

Industrial development and the economic progress have been accompanied by a growing energy demand. Majority of energy production is obtained by burning fossil fuels with variable sulfur content. International Energy Agency estimates that fossil fuels will still play a key role in total energy production for the years to come. Has estimated that fossil fuels will continue to supply almost 80% of world energy use until 2040, and according to the

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<sup>a</sup> *Universitatea Babeş-Bolyai, Facultatea de Chimie și Inginerie Chimică, Str. Kogălniceanu, Nr. 1, RO-400084 Cluj-Napoca, Romania, sdragan@chem.ubbcluj.ro*

World Energy Council, this might be prolonged to 2050 [1]. The combustion of coal and oil supply a significant part of the world's present energy demand and caused emissions of gas pollutants which released into the atmosphere affects the quality of the environment. An important particular case, with a major environmental impact, is the emissions of  $\text{SO}_2$ , which is considered as one of the gravest chemical threats to the global environment [1-4].

The industrial sectors that contribute to the  $\text{SO}_2$  emissions are the metallurgical ore refining and the power industry that use fossil fuels as raw material. Sulfur dioxide emissions are harmful both to the environment and to human health, therefore the problem of desulfurization gaseous emissions from burning fossil fuels is very important. These industrial sectors continue to come under pressure from both national and local regulatory groups to reduce sulfur dioxide emissions. With trend in the industry, retrofit technologies are likely to play an important role in any  $\text{SO}_2$  emission reduction strategy. Emissions of  $\text{SO}_2$  can be controlled in several ways: switch to a fuel that has lower sulfur content, or improve the efficiency of the industrial process so that less fuel is required. The sulfur in the fuel can in principle be removed before use; however, in practice, it is uneconomic to remove more than a small percentage of the sulfur. In many applications, the most efficient means of controlling  $\text{SO}_2$  emissions is to remove the  $\text{SO}_2$  from the flue gases before they are released to the atmosphere.

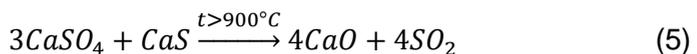
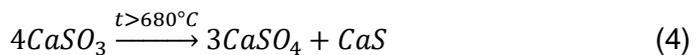
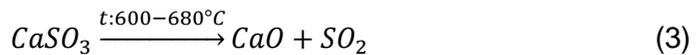
Several different flue gas desulfurization (FGD) technologies have been developed for this aim. This is the main reason why the issue of flue-gas desulfurization has become very topical and is the theme of numerous researches in this field [5-10].

In the last decade, technology of flue gas desulfurization has made considerable progress in terms of efficiency, reliability and those are classified into three main processes: dry, semi-dry and wet [5]. Independently of the classification, most of specialized literature [1-8] agrees on the fact that the wet limestone FGD system is the FGD process most widely used (87%) because of its high desulfurization performance and low operating cost [5]. In dry and/or semi-dry FGD systems,  $\text{SO}_2$ -containing flue gas reacts with an alkaline sorbent typically  $\text{Ca}(\text{OH})_2$  or  $\text{CaO}$ . Spray dry is the second most popular FGD system worldwide and mostly used for relatively small to medium capacity boilers using low to medium S (1.5 wt.%) coal. A comparative analysis of these shows that the process based on the solid-gas reaction using lime produced in situ by spraying fine grain limestone directly into the furnace (firing zone) is the process recommended for plants with a capacity less than 100 MWh in operation. It is simple to implement and exploited it does not require complex equipment. The main deficiency of the process consists in the low degree of use of lime, the large amount of waste that is difficult to recover, which poses problems of storage.

During the years, a number of investigations have been made on the kinetics of the reaction of  $SO_2$  with limestone in the dry desulfurization processes. Two major directions have been investigated: the effect of various process variables on the reaction rate and identification of the reaction mechanism with the help of a suitable model. The adsorption of  $SO_2$  by limestone involves two consecutive reaction steps: (a) dissociation of calcium carbonate and (b) reaction of  $CaO$  with  $SO_2$ . The rate of this second step is the important factor in all the pollution-control processes. Depending on the reactor temperature, reaction mechanisms of direct ( $CaCO_3$ ) or indirect ( $CaO$ ) sulfation of the limestone may occur [11]. It has been demonstrated that indirect sulfation reached the highest efficiency of desulfurization [12, 13].

Current research aims to establish the process mechanism and increase the use of lime and improve the  $SO_2$  retention efficiency. In this paper a kinetic study to show the influence of the main parameters (temperature and solids particle size) of the desulfurization process. Based on experimental measurements were calculated rate constants and activation energies. Numeric values of activation energy provide indications of the macro-kinetic mechanism of the global dry desulfurization process.

**Process chemistry:** The chemisorption process in the  $CaO-SO_2$  system is complex by the many transformation and transfer processes involved. The mass transformation processes that take place can be described by the reactions:



It is noticed that  $\text{CaSO}_4$  formation is possible in the absence of oxygen at temperatures above  $680^\circ\text{C}$ . At temperatures above  $900^\circ\text{C}$ ,  $\text{CaSO}_4$  can react with CaS when are recovered CaO and  $\text{SO}_2$ . The presence of excess oxygen in the combustion air promotes the chemisorption process as  $\text{CaSO}_3$  and it reacts with CaS according to the reactions:



## RESULTS AND DISCUSSION

The CaO sulfation degree was determined with the relationship deduced from the mass balance equations for the solid phase:

$$m_{\text{CaO}} = m_{\text{CaO}}^0 - m_{\text{CaO}}^0 \cdot \eta_{\text{CaO}} \quad (9)$$

$$m_{\text{CaSO}_4} = 0 + \frac{M_{\text{CaSO}_4}}{M_{\text{CaO}}} \cdot m_{\text{CaO}}^0 \cdot \eta_{\text{CaO}} \quad (10)$$

$$m_{\text{A}''} = m_{\text{A}''}^0 \quad (11)$$

$$m_{[\text{S}]} = m_{[\text{S}]}^0 \left( 1 + \frac{80}{56} \bar{x}_{\text{CaO}}^0 \cdot \eta_{\text{CaO}} \right) \quad (12)$$

$$\eta_{\text{CaO}} = \frac{56}{80} \cdot \frac{\Delta m_{[\text{S}]}}{m_{[\text{S}]}^0 \cdot \bar{x}_{\text{CaO}}^0} \quad (13)$$

The influence of temperature on the lime sulfation process for the three solid granulation classes used is shown in Figures 1-3.

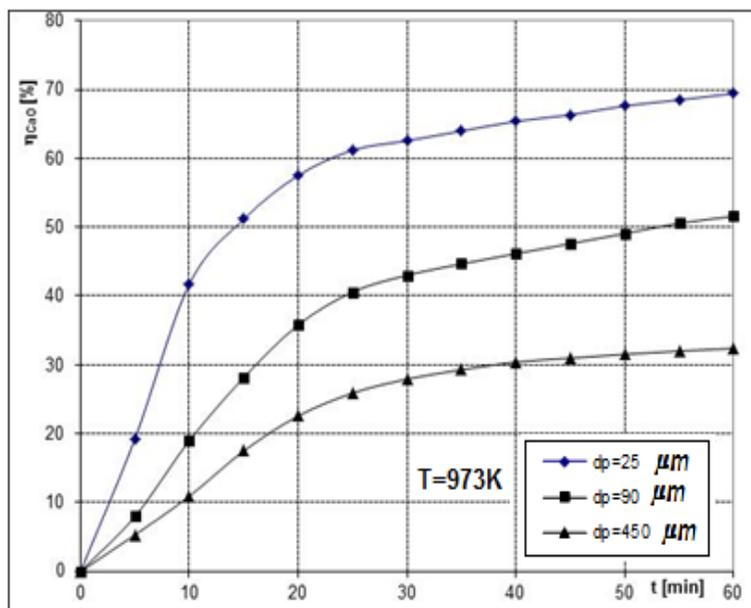


Figure 1. Influence of solid size on sulfation at temperature = 973 K

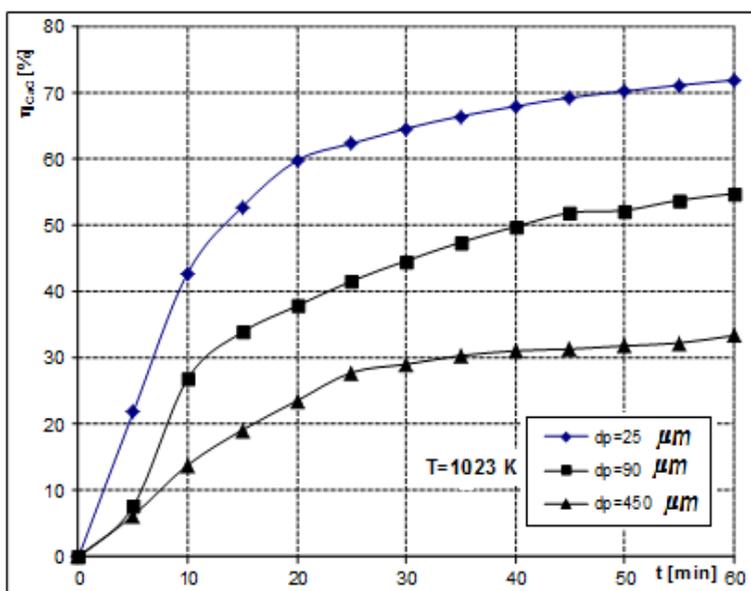
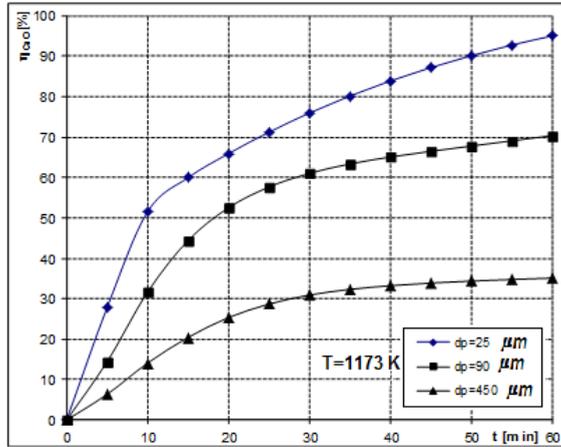


Figure 2. Influence of solid size on sulfation at temperature = 1023 K

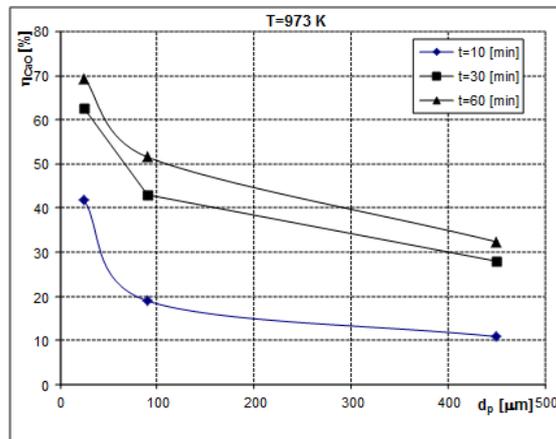


**Figure 3.** Influence of solid size on sulfation at T= 1173 K

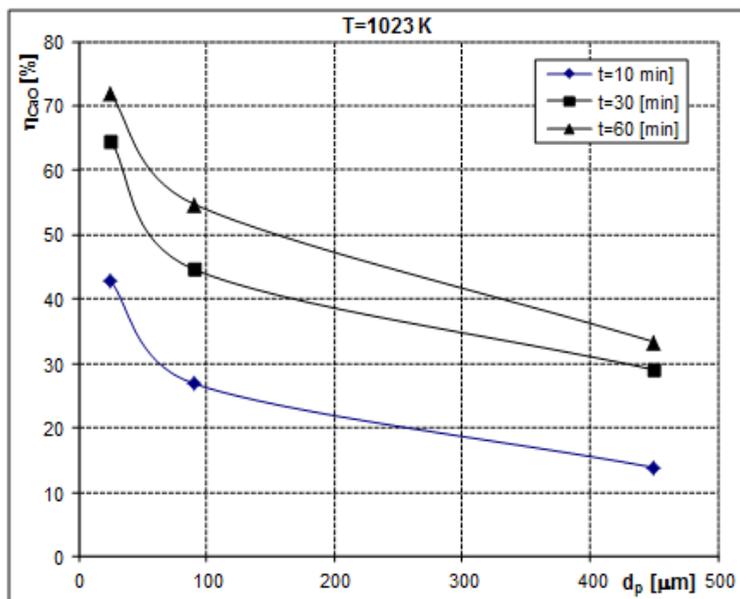
Temperature has a strong influence on the degree of sulfation in the case of the solids with 25 μm grains size. For solid particles with  $dp \leq 25 \mu\text{m}$ , the sulfation degree increases from 70% at T = 973K to 90% at T = 1173K.

From Figures 1-3 it can be seen that the rate of the sulfation process is greatly reduced after the first 15 minutes, for the grain size  $dp \geq 25 \mu\text{m}$ .

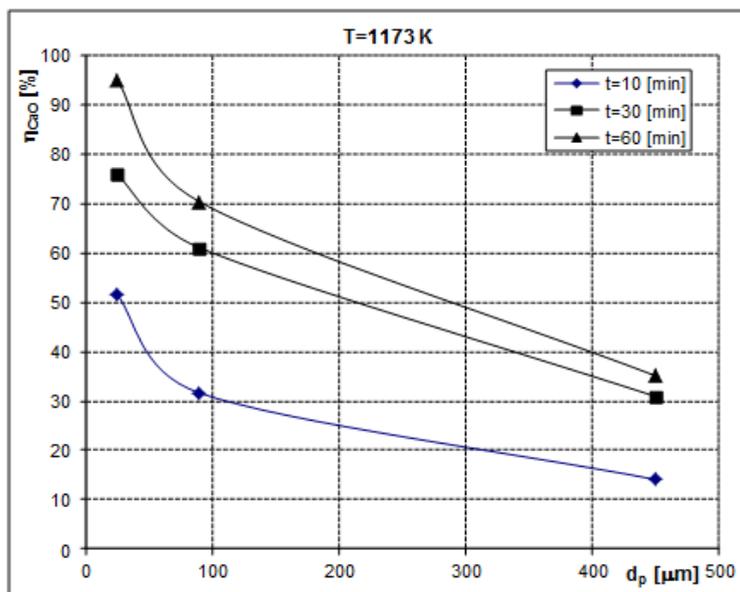
The influence of the solid particle size on the sulfation rate at different time and constant temperatures is shown in Figures 4-6.



**Figure 4.** Influence of solid size on sulfation at T= 973 K at different time intervals



**Figure 5.** Influence of solid size on sulfation at  $T = 1023$  K at different time intervals



**Figure 6.** Influence of solid size on sulfation at temperature = 1173 K at different time intervals

The diameter of the solid particles have a significant influence on the sulfation degree of lime, only for the solid particle size of  $d_p \leq 25\mu\text{m}$ , degree of sulfation is technologically acceptable when  $\eta_{\text{CaO}} = 95\%$  at  $T = 1173\text{ K}$ . For the grains solids with  $d_p \geq 90\mu\text{m}$ , the sulfation degree decreases significantly, so that for particles with  $d_p = 450\mu\text{m}$  the sulfation degree of lime does not exceed 35% even at  $T = 1173\text{ K}$ . Influence of the granulation of the solid on the sulfation degree of lime depends on the working temperature only for the solid particles with  $d_p \leq 25\mu\text{m}$ .

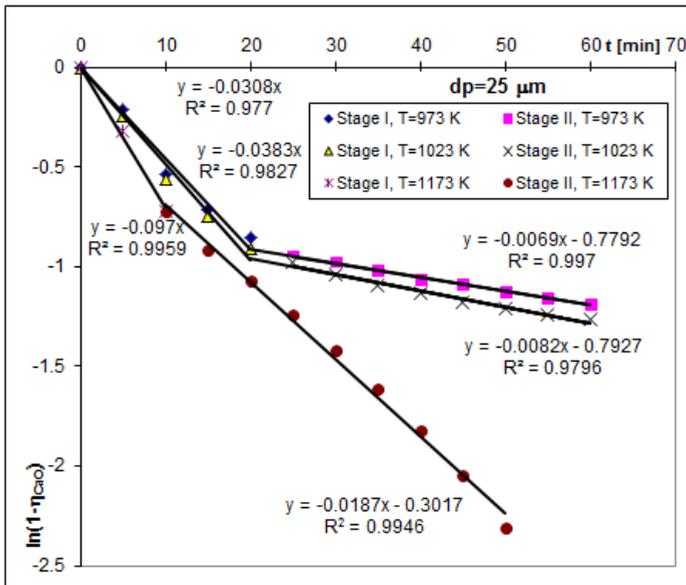
The experimental data obtained for evolution in time of the sulfation degree of lime shown in Figures 1-3 was used to calculate the rate constants of sulfation  $k$ , based on the following equation:

$$-\frac{dC_{\text{CaO}}}{dt} = k C_{\text{CaO}} \quad (14)$$

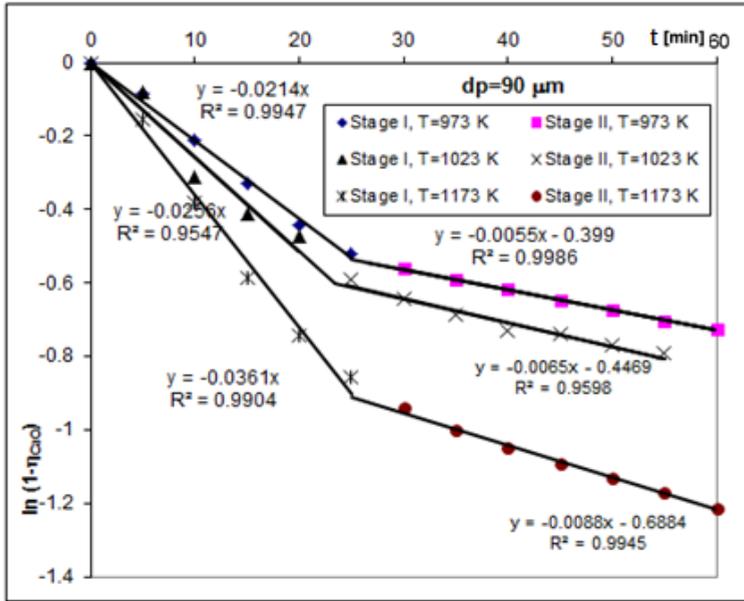
$$-\frac{dC_{\text{CaO}}}{C_{\text{CaO}}} = k dt \quad (15)$$

$$C_{\text{CaO}} = C_{\text{CaO}}^0(1 - \eta_{\text{CaO}}) \quad (16)$$

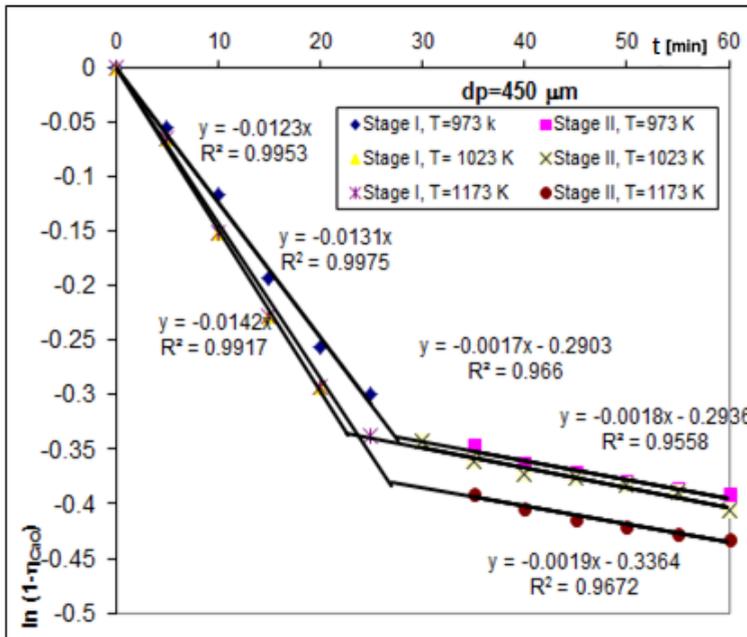
The integration of the equation for the boundary conditions leads to the expression specified:  $-\ln(1 - \eta_{\text{CaO}}) = k t$ . To determine the constants rate of the process of sulfation, have been represented the diagrams from Figures 7-9 in coordinate  $(1 - \eta_{\text{CaO}})$ -  $t$ .



**Figure 7.** Determination of the rate constants for grain solids with  $d_p = 25\mu\text{m}$



**Figure 8.** Determination of the rate constants for grain solids with  $dp = 90\mu\text{m}$



**Figure 9.** Determination of the rate constants for grain solids with  $dp = 450\mu\text{m}$

From the slope of the straight lines were obtained the values of the rate constants, that were used to determine the activation energies of by their graphic processing in the coordinates  $\ln k-1/T$ , according to the diagrams shown in figure 10-12.

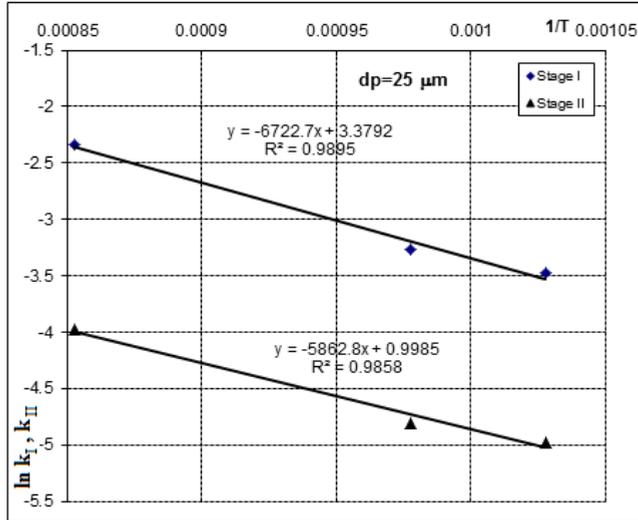


Figure 10. Determination the activation energies for solids with  $dp = 25 \mu m$

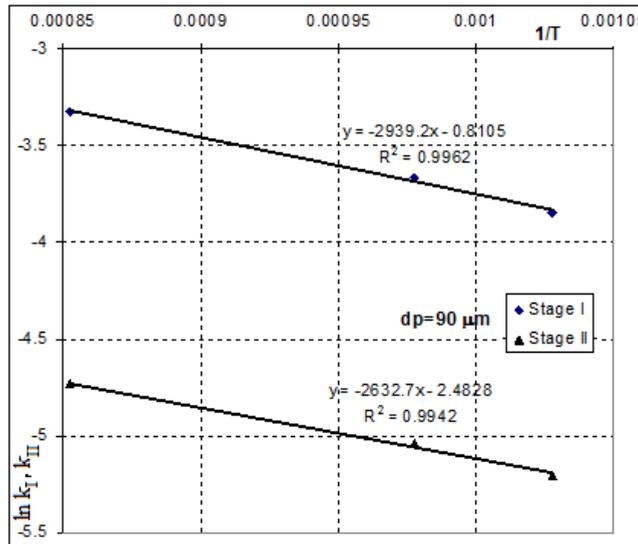
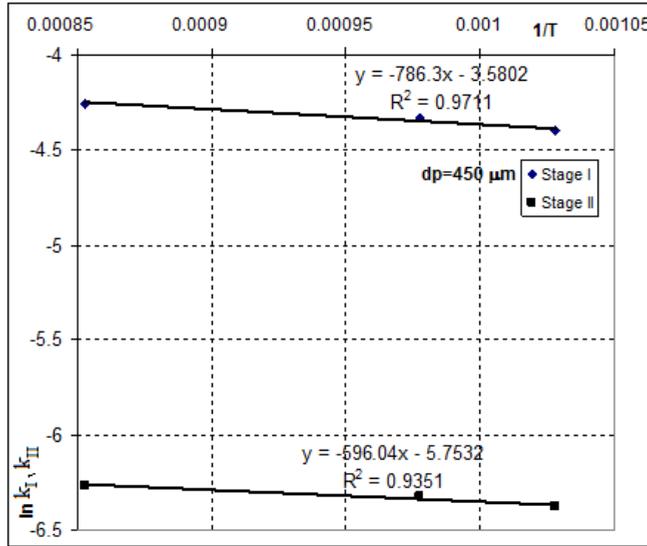


Figure 11. Determination the activation energies for solids with  $dp = 90 \mu m$



**Figure 12.** Determination the activation energies for solids with  $dp = 450\mu\text{m}$

The diagrams analysis suggests a change in the macrokinetic mechanism by which the global desulfurisation process takes place in time.

The numerical values of the rate constants and activation energies corresponding to the two stage of the sulfation process of under the working conditions are shown in Table1.

**Table 1.** The constants rate  $k$  and activation energies  $E_a$

dp [ $\mu\text{m}$ ]	k	T [K]			Ea [Kj/mol]	
		973	1023	1173	I	II
25	$k_I$	0.0308	0.0383	0.0971	55.92	48.76
	$k_{II}$	0.0069	0.0082	0.0187		
90	$k_I$	0.0214	0.0256	0.0361	21.9	21.51
	$k_{II}$	0.0055	0.0065	0.0088		
450	$k_I$	0.0123	0.0131	0.0142	6.54	4.95
	$k_{II}$	0.0017	0.0018	0.0019		

It can be seen that the values of the rate constants increase with increasing temperature and decrease with increasing solid grain size. For the stage I corresponding to the start-up of the process, the constants rate  $k_I$  are much higher than the constant rate  $k_{II}$  corresponding to the final stage of the process.

The analysis of the sulfation process based on the activation energies  $E_a$  shows that it is carried out according to the macrokinetic mass transformation model, only for small granules of the solid with  $d_p < 25 \mu\text{m}$  and temperatures  $T > 973 \text{ K}$ , where  $E_a$  has values higher than  $42 \text{ KJ/mol}$ . At higher granulations of the solid, the activation energy values shown that the sulfation process follows the pore-transfer macrokinetic model. The molar volume of  $\text{CaSO}_4$  that is formed is greater than the molar volume of the solid reagent, thus decreases the porosity of the solid. Thus, the Knudsen diffusion stage is passed, the share of mass transformation processes in the global process is decreasing, as evidenced by the strong decrease in the value of activation energies.

In the diffusion stage, the weight of mass transformation processes in the global process is decreasing, as evidenced by the strong decrease the activation energies.

## CONCLUSIONS

The dry injection of limestone fine particles in the power plant furnaces is the second most commonly used process for flue gas desulfurization and acid rain mitigation. The limestone is quickly converted to lime which further reacts with sulfur dioxide. The complete calcination is a key factor for higher desulfurization efficiency.

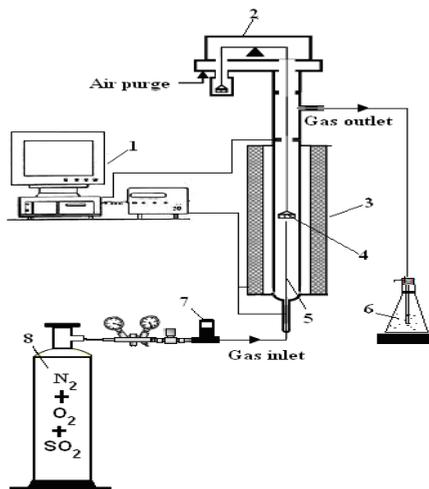
The effect of the principal variables affecting sulfur dioxide retention (temperature and solid particle size) and macrokinetic mechanism have been investigated with TGA technique. The influence of temperature on the conversion of solid particles, at a given time, is rather significant. This may suggest a transformation rate-determining step, with a great activation energy. Temperature has a strong influence on the degree of sulfation in the case of the solids with  $25 \mu\text{m}$  grains size. The diameter of the solid particles have a significant influence on the sulfation degree of lime. The sulfation reaction rate increase with decreasing sorbent particle size and increasing with temperature.

It can be observed that the sulfation process was performed by two stages. It is carried out according to the macrokinetic mass transformation model, only for small granules of the solid with  $d_p < 25 \mu\text{m}$  and temperatures  $T > 973 \text{ K}$ . where activation energy  $E_a$  has values higher than  $42 \text{ KJ/mol}$ .

The first stage was controlled by chemical reaction and/or diffusion through porous system of the particle. As the reaction proceeded, the pore volume of solid has decreased due to molar volume of the product ( $\text{CaSO}_4$ ) was higher than reagent solid ( $\text{CaO}$ ) and as a result the external pores were plugged. In this moment, the second stage began and it was controlled by diffusion through reaction product layer.

## EXPERIMENTAL SECTION

The experimental study was carried out with CAHN-TG 121, provided with a data acquisition and control DACS system, shown in Figure 13.



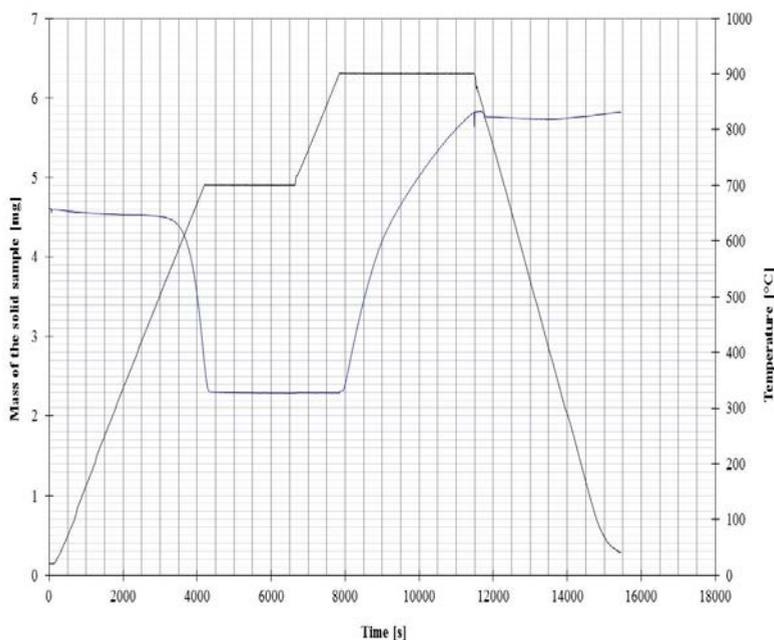
**Figure 13.** The schematic of the experimental setup for the SO<sub>2</sub> chemisorption on the solid CaO

The solid adsorbent used was CaO, obtained by calcination of Sandulesti (Turda) limestone, whose the chemical composition is shown in Table 2.

**Table 2.** Chemical composition of limestone

Component	CaCO <sub>3</sub>	Fe <sub>2</sub> O <sub>3</sub>	Al <sub>2</sub> O <sub>3</sub>	SiO <sub>2</sub>	MgCO <sub>3</sub>
(%)	96.5	1.8	0.86	0.33	0.5

Three granulation limestone classes were used, with:  $d_p = 0-50 \mu\text{m}$ ;  $d_p = 80-100 \mu\text{m}$ ;  $d_p = 400-500 \mu\text{m}$ . The carbonate decomposition was carried out by heating at  $T = 973 \text{ K}$  and pressure  $p = 0.34 \text{ bar}$  for 40 min to constant mass, then the resulting samples were brought into contact with the gaseous mixture with the concentration of 0.9% SO<sub>2</sub> and circulation rate of 0.046 m / s. The evolution of decomposition and sulfation processes was followed by sample mass variation and recorded in the form of thermogramme of the type shown in Figure 14.



**Figure 14.** The evolution of limestone decomposition and the sulfation process

## REFERENCES

1. R. del Valle-Zermeno, Formosa J. and Chimenos J.M., *Reviews in Chemical Engineering*, **2015**, 31 (4), 303.
2. A. Franco and A.R. Diaz , *Energy*, 2009, 34(3), 348.
3. T. Hlincik, P. Buryan, *Fuel* **2013a**; 104; 208.
4. T. Hlincik, P. Buryan *Fuel Process Technol.* **2013b**, 111, 62.
5. P. Córdoba, *Fuel*, **2015**, 144, 274.
6. S. Drăgan, *STUDIA Universitatis Babeş-Bolyai, Chemia*, **2017**, 62 (4), 283.
7. R. del Valle-Zermeno, J. Formosa, J.A. Aparicio, M. Guebbe, J.M. Chimenos. *Fuel Process Technol.*, **2015**, 138, 30.
8. L. Marocco, A. Mora, *Separation and Purification Technology*, **2013**, 108, 205.
9. S. Drăgan , Al. Ozunu, *Central European Journal of Chemistry*, **2012**, 10 (5), 1556.
10. Z. Zhu, Y. Ma, Z. Qu, L. Fang, W. Zhang, N Yan, *Fuel*, **2017**, 195, 33.
11. F. de Souza and S.R. Bragança, *Brazilian Journal of Chemical Engineering*, **2017**, 34 (1), 263.
12. D. Diego, L.F. Rufas, A. García-Labiano, F. de Las Obras-Loscertales, M. Abad, A. Gayán, P., Adánez, *Fuel*, **2013**, 114, 106.
13. F. de Souza, S.R. Bragança, *Thermochemica Acta*, **2013**, 561, 19.

## ANTIBACTERIAL ACTIVITY OF SILVER NANOPARTICLES OBTAINED BY CO-REDUCTION WITH SODIUM CITRATE AND TANNIC ACID

SORIN RAPUNTEAN<sup>a</sup>, REKA BALINT<sup>b</sup>, GERTRUD ALEXANDRA  
PALTINEAN<sup>b</sup>, GHEORGHE TOMOAI<sup>c,d</sup>, AURORA MOCANU<sup>b,\*</sup>,  
CSABA-PAL RACZ<sup>b</sup>, OSSY HOROVITZ<sup>b</sup>, MARIA TOMOAI<sup>a</sup>-COTISEL<sup>b,d</sup>

**ABSTRACT.** The wet chemical synthesis of silver nanoparticles (AgNPs) by reduction of silver nitrate with sodium citrate, tannic acid, and their mixture is reported. The obtained silver nanoparticles were investigated by UV-Vis spectroscopy, scanning transmission electron microscopy (STEM), and atomic force microscopy (AFM) images and their antibacterial effect against *Escherichia coli* was assessed.

**Keywords:** silver nanoparticles, sodium citrate, tannic acid, UV-Vis spectra, STEM, AFM, antibacterial effect

### INTRODUCTION

In the present war against antibiotic resistant bacterial strains, the old remedies with antimicrobial properties, such as certain metals or metal oxides were revived, but in the modern formula of nanoparticles (NPs). Most research is consecrated to silver nanoparticles (AgNPs), single or in association with antibiotics [1-12]. Nevertheless, the mechanism of the antibacterial action of

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<sup>a</sup> University of Agricultural Sciences and Veterinary Medicine of Cluj-Napoca, 3-5, Manastur Str., RO-400372, Cluj-Napoca, Romania

<sup>b</sup> Babes-Bolyai University of Cluj-Napoca, Faculty of Chemistry and Chemical Engineering, Physical Chemistry Centre, 11 Arany J. Str., RO-400028 Cluj-Napoca, Romania

<sup>c</sup> Iuliu Hatieganu University of Medicine and Pharmacy, Orthopedy and Traumatology Department, 47 Mosoiu T. Str., RO-400132 Cluj-Napoca, Romania

<sup>d</sup> Academy of Romanian Scientists, 54 Splaiul Independentei, RO-050094, Bucharest, Romania

\* Corresponding author [mocanu.aurora@gmail.com](mailto:mocanu.aurora@gmail.com)

AgNPs is still not fully understood; is it due to the attack of AgNPs themselves on bacterial cells, or to the silver ions released by the nanoparticles, or to the production of reactive oxidative species [12-15], induced by AgNPs.

For the preparation of AgNPs, three categories of methods are mentioned in literature [14, 16-18]: chemical methods based on the reduction of  $\text{Ag}^+$  ions in aqueous or in organic solution, physico-chemical methods (photo-physical and electrochemical) and biosynthesis. For the chemical methods in aqueous solution, various reduction agents were proposed, such as sodium citrate, sodium borohydride, hydrazine, glucose, hydroquinone, gallic acid,  $\beta$ -cyclodextrin, ascorbic acid, various plant extracts [7, 14-22]. Although, the excess of reduction agents acts sometimes also as stabilizer of the colloidal system of AgNPs, frequently supplementary capping agents are introduced, such as starch, gelatin, polyvinylpyrrolidone (PVP), polyvinyl alcohol (PVA), poly-L-lysine, cetyltrimethyl-ammonium bromide (CTAB) and other surfactants [18, 23].

Tannic acid (TA,  $\text{C}_{76}\text{H}_{52}\text{O}_{46}$ ) was also proposed, single or associated with another reduction agent, for the synthesis of metal nanoparticles [24], having many phenolic groups in the structure. Due to its large molecule (molar mass: 1700 g), TA appears itself as nanoparticle (hydrodynamic diameter 1.63 nm) [25]. It becomes unstable when temperature and pH increase, and suffers hydrolysis and oxidation reactions. The AgNPs modified with TA were proposed as microbicides and for wound healing [26-28]. TA was used for the preparation of AgNPs [22, 24, 29-34]. TA was also used together with sodium citrate to prepare AgNPs [35, 36] mainly for catalytic properties [35].

Our study is the first investigation to compare the synthesis of AgNPs using trisodium citrate (TSC) and TA independently and together at various molar ratios for the reduction of  $\text{Ag}^+$  ions in  $\text{AgNO}_3$  aqueous solutions to obtain AgNPs. Further, the characteristics of AgNPs realized by co-reduction for different mole ratios of the reagents and their biological effect on *Escherichia coli* were examined.

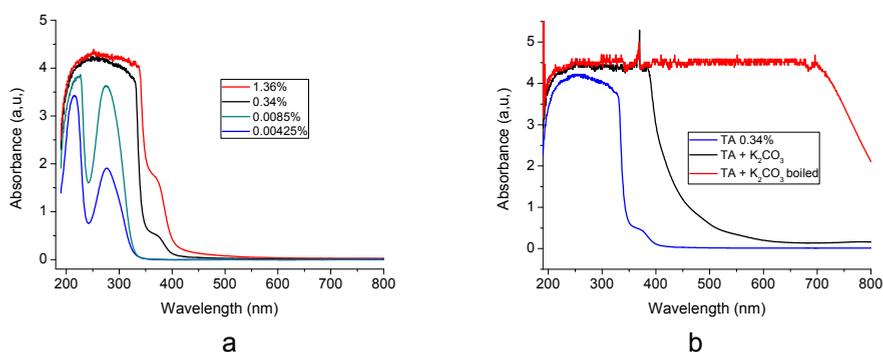
## RESULTS AND DISCUSSION

The samples prepared (colloidal silver solutions) from the various syntheses are presented in Table 1, along with the compositions of the mixtures of reactants (molar concentration and weight percentage), and the silver content in the final solution.

**Table 1.** The content of reactants in the syntheses of AgNPs and silver amount in the final colloidal solution

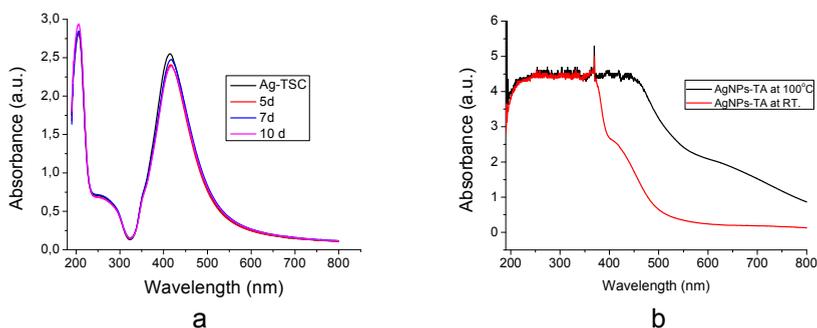
Sample Code	Content (mM/L)			Content (wt %)			Ag (mg/L)
	AgNO <sub>3</sub>	TSC	TA	AgNO <sub>3</sub>	TSC	TA	
Ag-TSC	0.96	1.49	-	0.0163	0.0384	-	104
Ag-TA	1	-	2	0.017	-	0.34	108
Ag-TSC-TA 1:7:2	1	7	2	0.017	0.1806	0.34	108
Ag-TSC-TA 1:7:0.2	0.25	1.75	0.05	0.00425	0.0452	0.0085	27
Ag-TSC-TA 1:3:0.2	0.25	0.75	0.05	0.00425	0.0193	0.0085	27
Ag-TSC-TA 1:20:0.1	0.25	5	0.025	0.00425	0.129	0.00425	27

**UV-Vis spectra.** Trisodium citrate and silver nitrate solutions present no absorption bands in the visible range of spectra. The light yellow colored tannic acid solutions (pH 6) present at higher TA concentrations a large absorption band in UV and at the lowest wavelengths in the visible spectrum. A shoulder is seen at about 370-380 nm. In diluted solution a deconvolution occurs, and two distinct bands are observed in the UV domain at 210 and 280 nm (Fig. 1a), in agreement with related data [22]. At higher pH values (alkaline medium, pH about 9), obtained by addition of potassium carbonate, the color turns rapidly to brown, due to the oxidation of phenolic groups by absorbed oxygen. The process is intensified by boiling (Fig. 1 b).



**Figure 1.** UV-Vis spectra: (a) aqueous solutions with different tannic acid content; (b) 0.34% (2 mM) tannic acid solution at pH 6, after alkalization with K<sub>2</sub>CO<sub>3</sub> (1 g/100 mL), and after boiling the alkaline solution for 15 min.

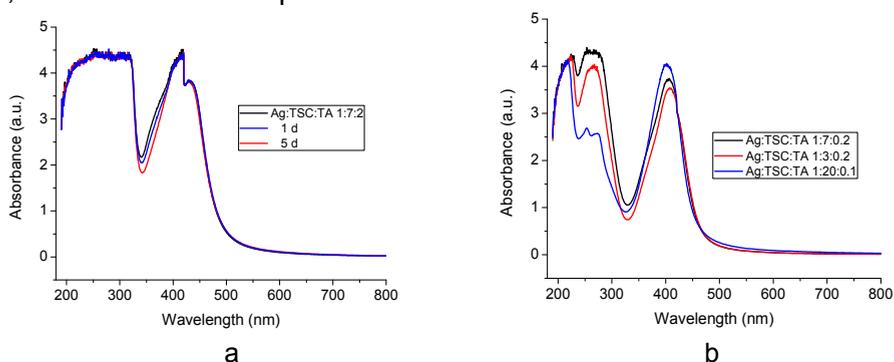
The UV-Vis spectra of the AgNPs obtained by reduction with TSC were measured in time (Fig. 2a) and the changes are minor for 10 days. The maximum absorption of the silver surface plasmon resonance (SPR) band is at 415-417 nm.



**Figure 2.** UV-Vis spectra of AgNPs obtained by reduction with TSC (a) and by reduction with TA at room temperature (RT) and at 100°C (b).

The solution obtained by reduction with TA (Ag 1 mM, TA 2 mM, as given in Table 1) at room temperature and at boiling temperature was diluted 8 fold with ultrapure water for the UV-Vis measurement (Fig. 2b). The characteristic SPR band of AgNPs cannot be distinguished because of its overlapping with the absorption band of TA and its oxidation products (resulted from the reaction with silver nitrate), the last one being more extended at higher temperature, as observed also for pure TA solutions (Fig. 1b).

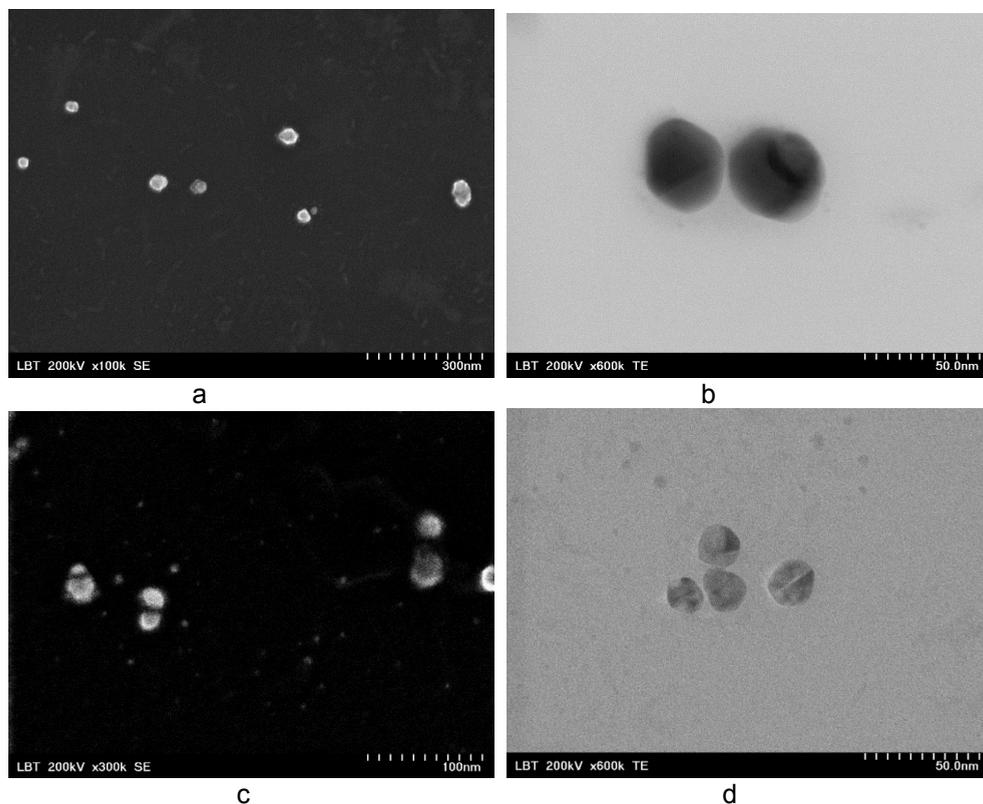
The AgNPs dispersion obtained by reduction with TSC and TA in the mole ratios Ag:TSC:TA for 1:7:2 (Table 1), with concentrations: Ag 1 mM, TSC 7 mM, tannic acid 2 mM (0.34%) had to be 4 fold diluted before measuring the UV-Vis spectra, in order to distinguish between the absorption bands for TA and the SPR band of AgNPs (Fig. 3a). The spectrum is rather unmodified for 5 days. The characteristic SPR band of AgNPs presents the maximum at 418 nm, while another lower peak is visible at 428 nm.



**Figure 3.** UV-Vis spectra of AgNPs prepared from AgNO<sub>3</sub> with trisodium citrate and tannic acid in molar ratio Ag:TSC:TA of 1:7:2, time evolution (a); comparison of UV-Vis spectra of AgNPs (b), prepared with different mole ratios of the reagents: Ag:TSC:TA, 1:7:0.2; 1:3:0.2; 1:20:0.1 (as given in Table 1); the Ag concentration in the final solutions was 0.25 mM.

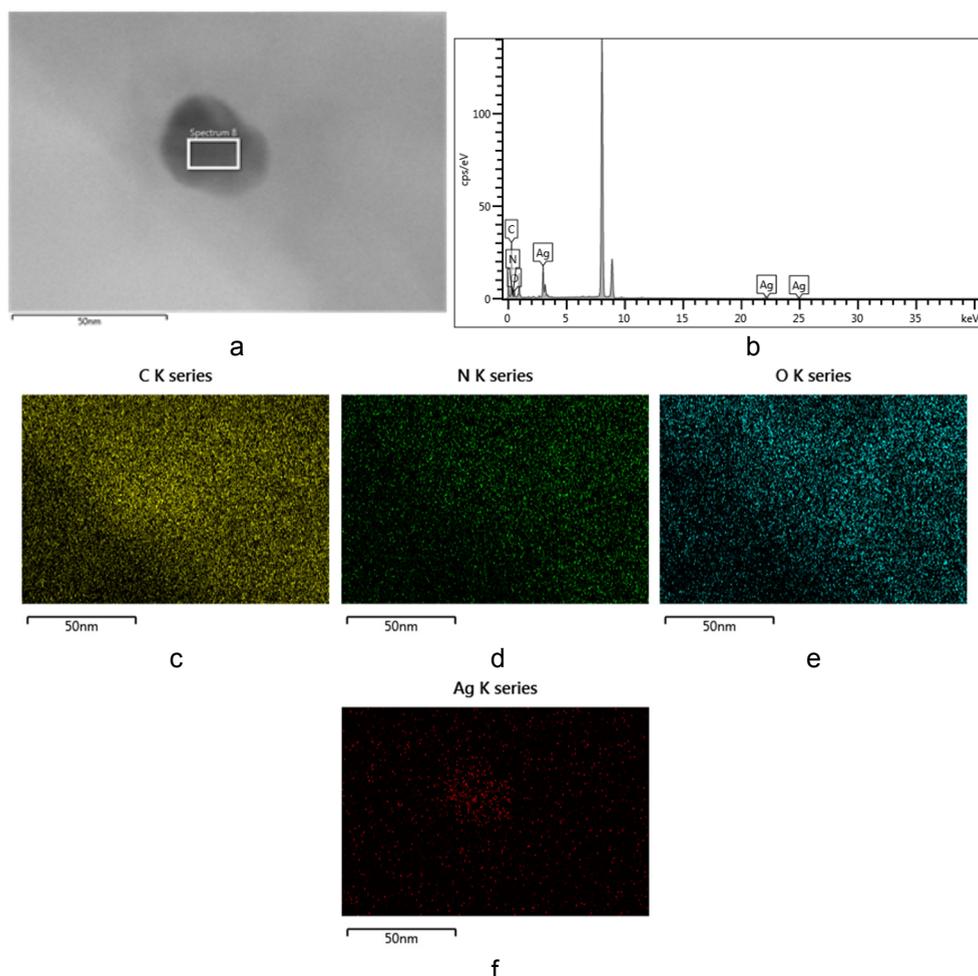
The spectra for the molar ratios Ag:TSC:TA 1:7:0.2 and 1:3:0.2, and 1:20:0.1 are compared in Fig. 3b. With a lowering of the TA content, the absorption band of TA and the SPR band of AgNPs become clearly separated; the maxima for AgNPs appear at 407 nm (1:7:0.2), 408 nm (1:3:0.2), and 402nm (1:20:0.1). The UV absorption band of TA in UV is split in two, as in Fig. 1a for low TA concentrations. In time, the AgNPs absorption bands remain almost unchanged for all samples (e.g. Fig. 3a).

**STEM images.** In the STEM images: Fig. 4a,b for AgNPs prepared with the Ag:TSC:TA 1:7:2 mole ratio, and Fig. 4c,d for those with 1:20:0.1 mole ratio, nearly spherical shaped AgNPs are observed. The sizes of the AgNPs in the samples were evaluated by measuring the diameters of a large number of particles on micrographs. For the different samples, the average diameter of particles was evaluated  $\pm$  the standard deviation going from  $10 \pm 5$  nm (Ag:TSC:TA for 1:20:0.1) to  $31 \pm 7$  nm (Ag:TSC:TA for 1:7:2).



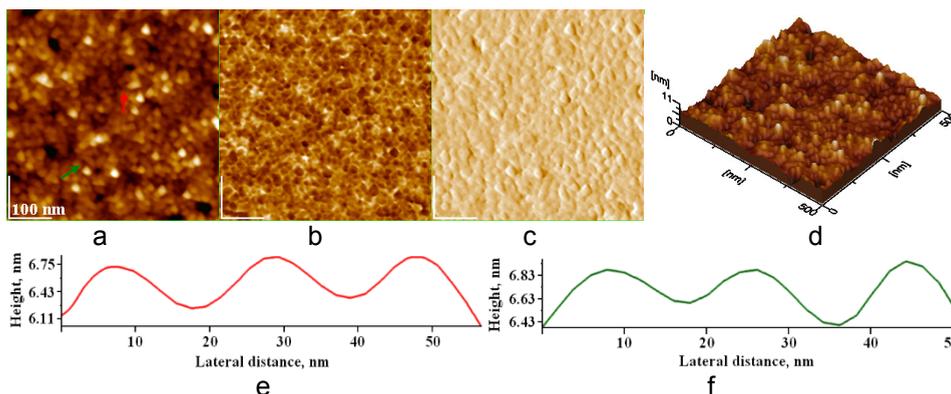
**Figure 4.** SE (a, c) and TE (b, d) images for AgNPs obtained with Ag:TSC:TA mole ratios 1:7:2 (a, b), and 1:20:0.1 (c, d). The scales in the STEM images correspond to 300 nm (a), 100 nm (c), and 50 nm (b, d).

In the energy dispersive X-ray spectroscopy, EDX spectra of AgNPs, obtained on STEM images (as in the example presented in Fig. 5a,b), the presence of Ag, as well of the elements from the organic compounds is evidenced (Fig. 5b). In Fig. 5c-f, the corresponding multicolor distribution maps for the individual elements (C, N, O and Ag) can be seen. The distribution maps confirm the presence of Ag mainly within the nanoparticle, while the other elements are distributed over the entire scanned area, due to the presence of the organic compounds (TSC, TA and their oxidation products) around the particles and among them, being adsorbed on STEM grid from deposited colloidal solution.



**Figure 5.** Electron image (a) and EDX spectrum (b), on AgNPs (within the white frame, shown in panel a) obtained for the Ag:TSC:TA sample at 1:3:0.2 mole ratio; distribution maps for C (c), N (d), O (e) and Ag (f); bars in the images are of 50 nm.

As a representative example, Fig. 6 presents **AFM images** on AgNPs obtained with the Ag:TSC:TA sample, having the 1:20:0.1 mole ratio. On AFM images, shape and size of nanoparticles can be estimated. The diameters of AgNPs, assessed from these images, are consistent with size obtained from STEM images, namely  $30 \text{ nm} \pm 5 \text{ nm}$  for Ag:TSC:TA at 1:7:2 molar ratio;  $22 \pm 4 \text{ nm}$  for Ag:TSC:TA at 1:3:0.2;  $16 \text{ nm} \pm 4 \text{ nm}$  for Ag:TSC:TA at 1:7:0.2; and  $10 \pm 3 \text{ nm}$  for Ag:TSC:TA at 1:20:0.1.



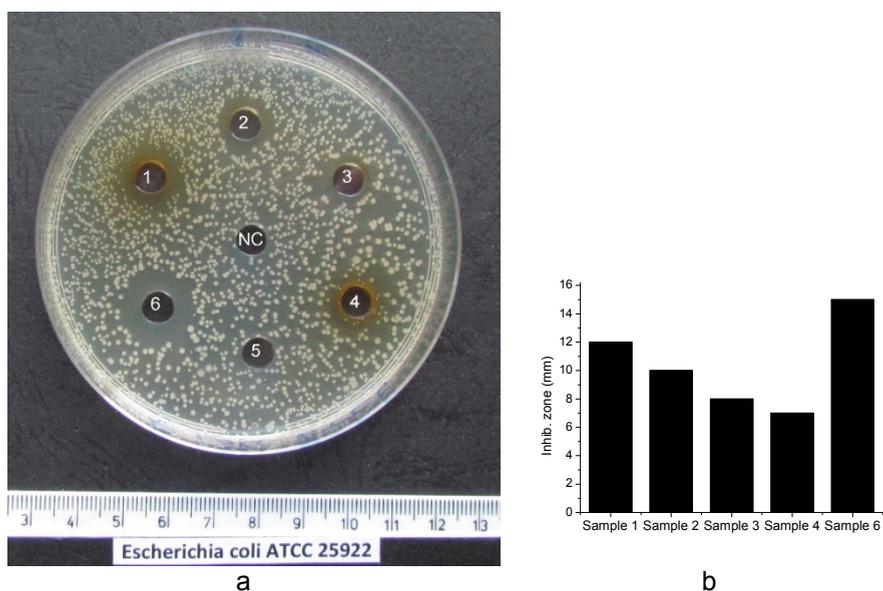
**Figure 6.** AFM images: 2D-topography (a), phase (b), amplitude (c) and 3D-topography (d) of AgNPs from Ag-TSC-TA sample at 1:20:0.1 mole ratio, deposited on glass by adsorption for 10 s; scanned area 500 nm x 500 nm; (e, f) cross section profiles along the arrows in panel a. Particles size is  $10 \text{ nm} \pm 3 \text{ nm}$ .

**Antimicrobial activity.** Figure 7a shows the agar plate with bacterial colonies of *Escherichia coli* 25922 ATCC after incubation at  $37^\circ\text{C}$  for 24 hours. The wells contained equal amounts ( $20 \mu\text{L}$ ) of the solutions to be tested. Among the four colloidal solutions, containing AgNPs with different Ag:TSC:TA mole ratios (Table 1), the largest inhibition zone were observed for the samples 1 and 2, corresponding to the mole ratios Ag:TSC:TA of 1:7:2 and 1:20:0.1 (Fig. 7b), respectively. The sample 2 had a lower silver content of  $0.25 \text{ mM}$  Ag in comparison with  $1 \text{ mM}$  Ag in the first sample, (Table 1), but showed a rather high antibacterial effect (Fig. 7b). This high antibacterial effect can be assigned to its smallest AgNPs (about  $10 \text{ nm}$  in average diameter).

The inhibition zone for the sample 3, corresponding to Ag( $0.25 \text{ mM}$ )-TSC-TA at 1:7:0.2 mole ratio is lower than for its value for samples 1 and 2. The lowest value of inhibition zone is for sample 4, corresponding to the Ag( $0.25 \text{ mM}$ )-TSC-TA at 1:3:0.2 mole ratio. For the last two samples, the size of particles is somehow bigger than the size of sample 2, but still smaller than the size of sample 1. Thus, antimicrobial activity of these samples appears to be related primarily with the average size of the AgNPs.

The inhibition zone for the 1 mM AgNO<sub>3</sub> solution is also important (sample 6), while the TSC solution (sample 5) and pure water (negative control, NC sample) presented no inhibition zones.

In addition, these results demonstrated that to increase the antibacterial activity of AgNPs, various reaction parameters should be considered, among which the mole ratio of reducing agent (TSC) and stabilizing (capping, TA) agent of NPs seems to be a crucial factor, to finally produce stable NPs of various sizes, and preferably small average diameters. These results are also consistent with our previous results, in which the antimicrobial activity of AgNPs was dependent on the silver amount [7] and on the size of nanoparticles.



**Figure 7.** (a) Inhibition zones for *Escherichia coli* 25922 ATCC in presence of AgNPs dispersions (20  $\mu$ L). Samples are numbered as follows: (1) - Ag (1 mM)-TSC-TA for 1:7:2 mole ratio; (2) - Ag (0.25 mM)-TSC-TA for 1:20:0.1; (3) - Ag (0.25 mM)-TSC-TA for 1:7:0.2 mole ratio; (4) - Ag (0.25 mM)-TSC-TA for 1:3:0.2 mole ratio; (5)- TSC (7 mM); (6) - AgNO<sub>3</sub> (1 mM); NC (negative control), pure water sample; (b) Comparison between inhibition zones for different samples.

Our results confirm the higher antibacterial activity of smaller particles, as previously observed by in other investigations, for instance [12, 37]. This effect can be explained by the theory which assigns the antibacterial effect to the AgNPs themselves [38], since smaller particles have a larger specific surface area, being able to stronger interact with the cell membrane [14], or

maybe penetrate into the cell [1]. On the other hand, if the antibacterial effect were due to the  $\text{Ag}^+$  ions [13, 39], it is again the higher specific surface area which would guarantee a higher release of silver ions. To these reasoning we could add that for the diffusion in agar plates, which is slower than in liquid medium, smaller particles have a higher mobility, thus increasing the size of inhibition zones.

## CONCLUSIONS

The co-reduction of silver nitrate in aqueous solution with TSC and TA proved to be a simple and rapid one-pot method to prepare AgNPs, with controlled size, from 30 to 10 nm, as shown by STEM and AFM images. The presence of elementary silver is evident from the UV-Vis spectra (characteristic SPR band) and from the EDX spectra and elements distribution maps. The antibacterial effect of the AgNPs was tested by measuring the inhibition zones on cultures of *Escherichia coli*. The effect was evident for all AgNPs samples, but the AgNPs with smallest size proved to be the most active. Thus AgNPs obtained by the investigated methods could be successfully used, as such or in association with antibiotics, against bacterial infections.

## EXPERIMENTAL SECTION

### *Synthesis of silver nanoparticles (AgNPs)*

The *reduction of silver nitrate with sodium citrate* [40, 41] was achieved by boiling for 30 min, under continuous stirring, a mixture of 5 mL 1% trisodium citrate, TSC ( $\text{Na}_3\text{C}_6\text{H}_5\text{O}_7$ , high purity, above 99%, from Sigma Aldrich) aqueous solution with 125 mL 1 mM  $\text{AgNO}_3$  (from Merck, high purity above 99.5%). The mole ratio TSC/Ag was 1.55. The pale yellow solution obtained was cooled on an ice-water mixture at room temperature. The colloidal solution (Ag content 104 mg/L, 0.96 mM) kept in brown bottles at 4°C, was stable for at least 2 months

*Tannic acid (TA)* was also tested as reducing agent for  $\text{AgNO}_3$ . The  $10^{-2}$  M  $\text{AgNO}_3$  solution (10 mL) with 80 mL water was mixed with 10 mL 3.4% tannic acid (from Merck) solution ( $2 \cdot 10^{-2}$  M) (molar ratio Ag/TA = 1:2) at room temperature and at boiling temperature. The yellow color due to the TA solution was intensified during the reaction, and shifted to brown, more intensely at high temperature. The resulting colloid solution had a silver content of 108 mg/L (1 mM).

The *co-reduction* of silver nitrate with *sodium citrate* and *tannic acid* for variable Ag/TSC/TA mole ratios (Table 1) was performed [35, 36], by heating up the AgNO<sub>3</sub> solution to boiling, and then adding the calculated amount of TSC and TA mixture. The solutions were kept boiling, under continuous stirring, for 15 min. The colloidal silver obtained by complete reduction of Ag<sup>+</sup> to Ag was 1 mM in the Ag-TSC-TA for 1:7:2 mole ratio, and 0.25 mM for 1:7:0.2, 1:3:0.2 and 1:20:0.1 mole ratios.

### **Measurements and instrumentation**

For *UV-VIS absorption spectra* measurements the Jasco UV/Vis V650 spectrophotometer was used, in the domain from 800 to 190 nm wavelength. Too intense colored solutions (containing TA or its oxidation products) had to be diluted with water before measurements.

*STEM* is a combined scanning electron (*SE*) and scanning transmission electron (*TE*) microscope, Hitachi HD-2700, operating at a maximum accelerating voltage of 200 kV. STEM is equipped with energy-dispersive X-ray spectrometer (EDS), which has two EDX detectors, from Oxford Instruments. STEM-EDS equipment was also used for EDX elemental analysis.

Layers of AgNPs on glass support were prepared by vertical adsorption from the colloidal solutions for 10 s and dried in air. They were investigated by *atomic force microscopy*, AFM JEOL 4210 equipment, operated in tapping mode [42-48] using standard cantilevers with silicon nitride tips (resonant frequency in the range of 200-300 kHz and spring constant 17.5 N/m). On each film, areas from 10 μm x 10 μm up to 0.05 μm x 0.05 μm were scanned, and 2D- and 3D- topographies, phase and amplitude images and cross sections profiles in the adsorbed AgNPs layer were processed by the standard AFM procedures

*Antibacterial assays.* The bacterial strain tested was *Escherichia coli* ATCC 25922 (Gram negative bacteria), cultivated on glucose nutrient agar (Merck KGaA, Germany); suspensions in nutrient broth were prepared corresponding to a turbidity of 0.5 on the McFarland standards (1.5 x 10<sup>8</sup> CFU/mL).

The Kirby-Bauer technique [7] (disk diffusion assay) in nutrient agar with glucose, was applied to evaluate the antimicrobial effect of the AgNPs solutions. On nutrient agar plates (3 mm thick, diameter 90 mm), the bacterial suspension was introduced and uniformly dispersed. After drying the agar surface, on each plate 6 wells (diameter 6 mm) in radial disposition and a central one were cut and in each 20 μL liquid samples were introduced, namely the four AgNPs dispersions obtained by reduction with TSC and TA (Table 1), TSC solution, 1 mM AgNO<sub>3</sub> solution, and pure water as negative control in the

central well. The plates were incubated for 24 h at 37°C, and after that the diameters of the inhibition zones formed around the samples were measured and photographed. The plates were kept under surveillance for another 72 h, in order to reveal possible changes in time. The antimicrobial effect of AgNPs was sustained on *Escherichia coli* ATCC 25922.

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## REFERENCES

1. J.R. Morones, J.L. Elechiguerra, A. Camacho, K. Holt, J.B. Kouri, J.T. Ramirez, M. J. Yacaman, *Nanotechnology*, **2005**, 16(10), 2346.
2. M. Rai, A. Yadav, A. Gade, *Biotechnology Advances*, **2009**, 27(1), 76.
3. M.K. Rai, S.D. Deshmukh, A.P. Ingle, A.K. Gade, *Journal of Applied. Microbiology*, **2012**, 112(5), 841.
4. M.J. Hajipour, K.M. Fromm, A.A. Ashkarran, D.J. de Aberasturi, I.R. de Larramendi, T. Rojo, V. Serpooshan, W.J. Parak, M. Mahmoudi, *Trends in Biotechnology*, **2012**, 30(10), 499.
5. S. Chernousova, M. Epple, *Angewandte Chemie International Edition*, **2013**, 52(6), 1636.
6. A. Ravindran, P. Chandran, S. Sudheer Khan, *Colloids and Surfaces B: Biointerfaces*, **2013**, 105, 342.
7. A. Mocanu, G. Furtos, S. Rapuntean, O. Horovitz, C. Flore, C. Garbo, A. Danisteanu, G. Rapuntean, C. Prejmerean, M. Tomoaia-Cotisel, *Applied Surface Science*, **2014**, 298, 225.
8. G. Franci, A. Falanga, S. Galdiero, L. Palomba, M. Rai, G. Morelli, M. Galdiero, *Molecules*, **2015**, 20(5), 8856.
9. B. Le Ouay, F. Stellacci, *Nano Today*, **2015**, 10(3), 339.
10. N. Durán, G. Nakazato, A.B. Seabra, *Applied Microbiology and Biotechnology*, **2016**, 100(15), 6555.
11. J.S. Möhler, W. Sim, M.A.T. Blaskovich, M.A. Cooper, Z.M. Ziora, *Biotechnology Advances*, **2018**, 36(5), 1391.
12. V. Pareek, R. Gupta, J. Panwar, *Materials Science & Engineering C*, **2018**, 90, 739.

13. R. Ma, C. Levard, S.M. Marinakos, Y. Cheng, J. Liu, F.M. Michel, G.E. Brown Jr, G.V. Lowry, *Environmental Science & Technology*, **2012**, 46(2), 752.
14. J.J. Naddeo, M. Ratti, S.M. O'Malley, J.C. Griepenburg, D.M. Bubb, E.A. Klein, *Advanced Science, Engineering and Medicine*, **2015**, 7(12), 1044.
15. U.T. Khatoon, G.V.S. Nageswara Rao, K.M. Mohan, A. Ramanaviciene, A. Ramanavicius, *Vacuum*, **2017**, 146, 259.
16. N.L. Pacioni, C.D. Borsarelli, V. Rey, A.V. Veglia, In „Silver Nanoparticle Applications in the Fabrication and Design of Medical and Biosensing Devices”, Springer, Switzerland, **2015**, pp. 13-46.
17. K.S. Siddiqi, A. Husen, R.A.K. Rao, *Journal of Nanobiotechnology*, **2018**, 16:14. <https://doi.org/10.1186/s12951-018-0334-5>
18. V. Chahar, B. Sharma, G. Shukla, A. Srivastava, A. Bhatnagar, *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, **2018**, 554, 149.
19. A. Mocanu, O. Horovitz, C.P. Racz, M. Tomoia-Cotisel, *Revue Roumaine de Chimie*, **2015**, 60(7-8), 721.
20. A.R. Shet, P. Ghose, L. Patil, V. Hombalimath, *International Journal of Current Biotechnology*, **2015**, 3(2):1.
21. S. Aashriha, *International Research Journal of Pharmacy*, **2013**, 4, 111.
22. Y. Hao, N. Zhang, J. Luo, X. Liu, *NANO: Brief Reports and Reviews*, **2018**, 13 1850003, DOI: 10.1142/S1793292018500030.
23. A.L. Kubo, I. Capjak, I.V. Vrček, O.M. Bondarenko, I. Kurvet, H. Vija, A. Ivask, K. Kasemets, A. Kahru, *Colloids and Surfaces B: Biointerfaces*, **2018**, 170, 401.
24. T. Ahmad, *Journal of Nanotechnology*, **2014**, Article ID 954206, <http://dx.doi.org/10.1155/2014/954206>
25. M. Ocwieja, A. Popov, Z. Adamczyk, M. Morga, A. Ramanaviciene, A. Ramanavicius, *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, **2015**, 477, 70.
26. P. Orłowski, M. Krzyzowska, R. Zdanowski, A. Winnicka, J. Nowakowska, W. Stankiewicz, E. Tomaszewska, G. Celichowski, J. Grobelny, *Toxicology in Vitro*, **2013**, 27(6), 1798.
27. P. Orłowski, K. Soliwoda, E. Tomaszewska, K. Bien, A. Fruba, M. Gniadek, O. Labeledz, Z. Nowak, G. Celichowski, J. Grobelny, M. Krzyzowska, *Toxicology in Vitro*, **2016**, 35, 43.
28. P. Orłowski, M. Zmigrodzka, E. Tomaszewska, K. Ranoszek-Soliwoda, M. Czupryn, M. Antos-Bielska, J. Szemraj, G. Celichowski, J. Grobelny, M. Krzyzowska, *International Journal of Nanomedicine*, **2018**, 13, 991.
29. S.K. Sivaraman, I. Elango, S. Kumar, V. Santhanam, *Current Science*, **2009**, 97(7), 1055.
30. U.K. Parashar, V. Kumar, T. Bera, P.S. Saxena, G. Nath, S.K. Srivastava, R. Giri, A. Srivastava, *Nanotechnology*, **2011**, 22(41), 415104.
31. I. Călinescu, M. Pătrașcu, A.I. Gavrilă, A. Trifan, C. Boscornea, *U.P.B. Scientific Bulletin, Series B*, **2011**, 73(4), 1.
32. Y. Cao, R. Zheng, X Ji, H. Liu, R. Xie, W. Yang, *Langmuir*, **2014**, 30(13), 3876.
33. K. Watcharaporn, M. Opaprakasit, V. Pimpan, *Advanced Materials Research*, **2014**, 911, 110.

34. T.Y. Kim, S.H. Cha, S. Cho, Y. Park, *Archives of Pharmacal Research*, **2016**, *39*, 465.
35. N.G. Bastus, F. Merkoci, J. Piella, V. Puntès, *Chemistry of Materials*, **2014**, *26*, 2836.
36. K. Ranošzek-Soliwoda, E. Tomaszewska, E. Socha, P. Krzyczmonik, A. Ignaczak, P. Orłowski, M. Krzyżowska, G. Celichowski, J. Grobelny, *Journal of Nanoparticle Research*, **2017**, *19*:273, DOI 10.1007/s11051-017-3973-9
37. M. Akter, M.T. Sikder, M.M. Rahman, A.K.M.A. Ullah, K.F.B. Hossain, S. Banik, T. Hosokawa, T. Saito, M. Kurasaki, *Journal of Advanced Research*, **2018**, *9*, 1.
38. S. Agnihotri, S. Mukherji, S. Mukherji, *RSC Advances*, **2014**, *4*, 3974.
39. Z. Xiu, Q. Zhang, H.L. Puppala, V.L. Colvin, P.J.J. Alvarez, *Nano Letters*, **2012**, *12*(8), 4271.
40. P.V. Kamat, M. Flumiani, G.V. Hartland, *Journal of Physical Chemistry B*, **1998**, *102*, 3123.
41. O. Horovitz, M. Tomoaia-Cotisel, C. Racz, G. Tomoaia, L.D. Bobos, A. Mocanu, *Studia Universitatis Babeş-Bolyai, Chemia*, **2009**, *54*(3), 89.
42. M. Tomoaia-Cotisel, A. Tomoaia-Cotisel, T. Yupsanis, G. Tomoaia, I. Balea, A. Mocanu, C. Racz, *Revue Roumaine de Chimie*, **2006**, *51*(12), 1181.
43. G. Tomoaia, M. Tomoaia-Cotisel, A. Mocanu, O. Horovitz, L.D. Bobos, M. Crisan, I. Petean, *Journal of Optoelectronics and Advanced Materials*, **2008**, *10*(4), 961.
44. G. Tomoaia, P.T. Frangopol, O. Horovitz, L.D. Bobos, A. Mocanu, M. Tomoaia-Cotisel, *Journal of Nanoscience and Nanotechnology*, **2011**, *11*(9), 7762.
45. A. Mocanu, R.D. Pasca, G. Tomoaia, C. Garbo, P.T. Frangopol, O. Horovitz, M. Tomoaia-Cotisel, *International Journal of Nanomedicine*, **2013**, *8*, 3867.
46. U.V. Zdrengea, G. Tomoaia, D.V. Pop-Toader, A. Mocanu, O. Horovitz, M. Tomoaia-Cotisel, *Combinatorial Chemistry & High Throughput Screening*, **2011**, *14*(4), 237.
47. R.D. Pasca, A. Mocanu, S.C. Cobzac, I. Petean, O. Horovitz, M. Tomoaia-Cotisel, *Particulate Science and Technology*, **2014**, *32*(2), 131.
48. I. Petean, G. Tomoaia, O. Horovitz, A. Mocanu, M. Tomoaia-Cotisel, *Journal of Optoelectronics and Advanced Materials*, **2008**, *10*(9), 2289.



## MICROWAVE IRRADIATION EFFECT ON POLYPHENOL CONTENT AND ANTIOXIDANT ACTIVITY OF BASIL

ILDIKO LUNG<sup>a</sup>, MARIA-LOREDANA SORAN<sup>a\*</sup>, OCSANA OPRİȘ<sup>a</sup>,  
MANUELA STAN<sup>a</sup>, CONSTANTIN BELE<sup>b</sup>

**ABSTRACT.** This study investigates the influence of microwave field derived from wireless router and mobile telephony sources on polyphenol content and antioxidant activity of basil. The total phenolic content of basil extracts was determined by Folin-Ciocalteu method. The amount of polyphenolic compounds in basil plants exposed to microwave irradiation was higher compared to control plants. Extracts of irradiated basil exhibit higher antioxidant activity than extracts of control plants, as evidenced by the three methods of determination used: 2,2-diphenylpicrylhydrazyl radical scavenging activity, oxygen radical absorbance capacity and Trolox equivalent antioxidant capacity assays. It was determined that microwave irradiation increased both antioxidant activity and polyphenol content of basil extracts. In addition, hydroalcoholic extracts obtained from basil plants exposed to GSM microwaves showed higher antioxidant activity than hydroalcoholic extracts of plants exposed to WLAN microwaves.

**Keywords:** *polyphenols, basil, microwave influence, antioxidant activity*

### INTRODUCTION

The consumption of the phytochemical products/food supplements, in special phenolic compounds is correlated with a series of beneficial effects in the body, such as reducing the risk of diabetes, obesity, coronary artery disease, cancer of the colon and gastrointestinal dysfunctions [1]. Because in plants phenolic compounds are found under different forms the extraction from plant material raises special problems [2, 3]. Currently the existing analyzes

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<sup>a</sup> National Institute for Research and Development of Isotopic and Molecular Technologies, 67-103 Donat str., RO-400293, Cluj-Napoca, Romania.

<sup>b</sup> University of Agricultural Sciences and Veterinary Medicine, 3-5 Calea Manastur, RO-400372, Cluj-Napoca, Romania.

\* Corresponding author [loredana.soran@itim-cj.ro](mailto:loredana.soran@itim-cj.ro)

protocols appeals to classical techniques of liquid - solid extraction. Among these, the most commonly used are maceration, refluxing and Soxhlet extraction. Due to disadvantages that show they tend to be replaced by techniques with lower reagent consumption, lower energy and time, having however similar efficiency. For this purpose, have been implemented techniques such as ultrasound-assisted solvent extraction [4-6], microwave assisted solvent extraction [7], accelerated solvent extraction/pressurized liquid extraction carried out in dynamic and static mode [8] etc. Usually, for extraction of polyphenolic compounds must be performed an optimization [9] regarding the extraction system [10] solvent and plant material ratio, the effective time and extraction conditions, and each technique presenting specific parameters.

The obtained polyphenolic extract can be analyzed both globally - nonspecific, determining the total polyphenols and specific. In order to determine the total polyphenols, it is used spectrophotometric method, using specific reagents. There are two such reagents: Folin-Ciocalteu reagent and Arnou's reagent [11-13]. The specific determination can be achieved through chromatographic methods, most used being high-performance liquid chromatography with diode array detector [10] or coupled with a mass spectrometer [14].

Basil is an important medicinal plant and a culinary herb from the Lamiaceae family, known for carminative, galactogogue, stomachic and antispasmodic properties used in folk medicine. It is also known for its anti-inflammatory, antiviral and antimicrobial activities [15].

Under various biotic and abiotic stresses, plants can respond to produce secondary metabolites such as phenolic compounds, terpenoids, and alkanoids [16, 17].

Currently, due to the technological development, there is an exponential growth in the use of mobile phones and wireless devices, thus leading to an increase of electromagnetic radiation level and the emergence of a new type of stress, the electromagnetic field, especially in the microwave range (1-100 GHz) [18, 19]. Numerous studies confirm that microwave radiation influences the living organisms, including plants, even at irradiation power under limits [20-22].

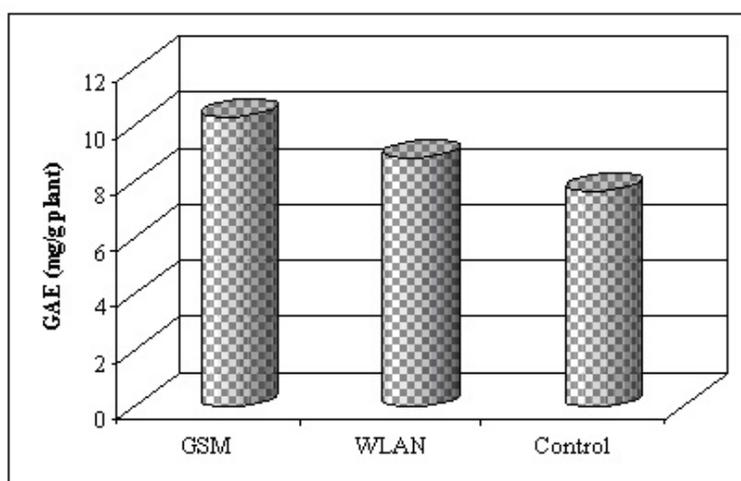
This manuscript reports the effect of the microwave field on polyphenolic content and antioxidant activity of basil. New results obtained by this work complete the study presented in our previous paper [23]. The basil was grown now in GSM and WLAN field. In this study, the grinding followed by centrifugation was used as extraction method instead of microwave assisted extraction. The antioxidant activity was determined and correlated with total polyphenol content.

## RESULTS AND DISCUSSION

The basil seedlings were exposed to microwave radiation of GSM and WLAN frequency bands. The effect on total phenolic content and antioxidant activity was studied for irradiated and control (non-irradiated) plant samples.

### *Determination of Total Phenolic Content*

In this work, the total polyphenol content of basil plants grown in the presence of microwave field and in normal conditions was analysed by the conventional spectrophotometric Folin-Ciocalteu method (Figure 1).



**Figure 1.** Total polyphenol content expressed as gallic acid equivalents in basil.

The amount of polyphenolic compounds was expressed as mg gallic acid/g fresh weight (FW), using the linear equation of the standard calibration curve:  $y = 0.6687x + 0.007$  ( $R^2=0.9987$ ). It was observed that in the case of microwave irradiated plants the polyphenolic amount was higher compared to the control plants. The highest polyphenol quantity was determined in basil irradiated with GSM frequency microwaves (10.23 mg GAE/g plant).

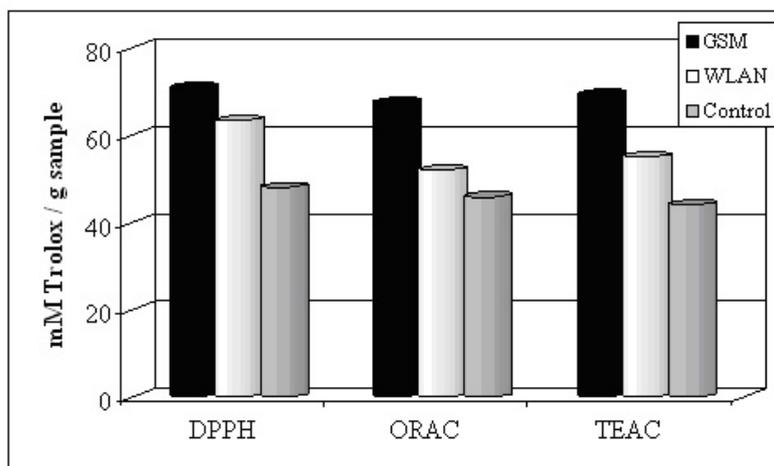
### *Total antioxidant activity*

Three antioxidant activity methods have been used to compare the antioxidant activity of irradiated and reference basil plants. DPPH method is often used due to stability, simplicity and its reproducibility [24].

The ORAC method is based on the use of fluorescein with progress of the reaction by decreasing of fluorescence emission. The decrease of fluorescence in the presence of radical species, including the peroxy species generated by AAPH is an index of antioxidant capacity against free radical species [25].

ABTS assay is considered as a method that measures the redox ability of the antioxidant mixture in relation to the radical cation ABTS+. This method is based on the absorption of plant extracts at 734 nm [26].

The antioxidant activity determined by DPPH, ORAC and TEAC methods (Figure 2) was expressed in mM Trolox equivalents (mM Trolox/g sample), using the linear equation of the standard calibration curve:  $y=0.0008x + 0.0025$  ( $R^2=0.9994$ ) for DPPH method,  $y=0.153x + 0.3451$  ( $R^2=0.9902$ ) for ORAC method and  $y=0.0008x + 0.0028$  ( $R^2=0.997$ ) for TEAC method.



**Figure 2.** The antioxidant activity of basil extracts.

Plants irradiated with microwaves have shown highest antioxidant activity, evidenced by the three methods of determination: DPPH, TEAC and ORAC. In addition, the extracts from basil plants exposed to GSM microwaves have shown highest antioxidant activity than those of the plants exposed to WLAN microwaves. The values of antioxidant activity determined by the three analysis methods were very close to each other.

The increased antioxidant activity of the extracts obtained from irradiated plants can be correlated with the increase in the amount of polyphenols.

## CONCLUSIONS

The objective of the present study was to investigate the effect of low-intensity microwave radiation on the total amount of polyphenols and antioxidant activity of basil. As result of our studies, microwave irradiation increased both antioxidant properties and polyphenol content of basil. Thus, the amount of total polyphenols in microwave irradiated plants increase with 33.73% (GSM) and 15.16% (WLAN) compared to control plants, respectively. In addition, the antioxidant activity was higher for the extracts obtained from GSM irradiated plants compared to WLAN.

## EXPERIMENTAL SECTION

### *Plant material*

The plant material used for the current experiment was basil, which was primarily grown from seeds (ARO, Romania). Three weeks after sowing, the plants were placed in identical anechoic chambers [27]. Test (irradiated) and control plants were subjected to the same environment (humidity and temperature). Stimulation was performed with microwaves modulated by a specific generator in GSM frequency domain (860 - 910 MHz range) and WLAN communications protocol, in the 2.412 - 2.48 GHz frequency band. The irradiation was performed over two weeks, afterwards the plants were taken from the chambers and the vegetal material was extracted.

### *Chemical reagents*

2,2'- Diphenyl – picrylhydrazyl (DPPH), 6 – hydroxy-2,5,7,8-tetramethylchroman – 2 carboxylic acid (Trolox), fluorescein sodium salt, 2,2'-azobis (2-amidinopropane) dihydrochloride (AAPH), 2,2'-azinobis(3-ethylbenzthiazoline-6-sulfonic acid) (ABTS), potassium persulfate, Folin-Ciocalteu reagent, gallic acid and anhydrous carbonate were employed from Sigma-Aldrich, Germany, while ethanol was purchased from Chimopar, Romania. All chemicals were of analytical grade.

### *Preparation of plant extracts*

One gram of fresh leaves was mortared with 1 g quartz sand after which 25 mL of 80% ethanol was added. The mixture was stirred for 1 h at 4°C and then centrifuged at 10.000 rpm for 15 minutes. The supernatant was removed and the pellet was resumed with 5 mL 80% ethanol, stirred for 15 minutes and centrifuged following the same procedure. Supernatants have been finally combined. Each sample was extracted independently in triplicate and the analyses were performed in the same day. Each extract was performed in three parallel samples.

### ***Determination of Total Phenolic Content***

The total polyphenol content was spectrophotometrically determined, using gallic acid as standard, according to the Folin-Ciocalteu method [28]. In a 10 mL volumetric flask to 1 mL of extract was added 5 mL of distilled water and 0.5 mL Folin-Ciocalteu reagent. After 3 min, 1.5 mL of sodium carbonate (5 g/L) was added and filled up to 10 mL with distilled water. Then, the samples were kept in a water bath at 50°C for 16 min. After cooling, their absorbances were read at 765 nm against distilled water as the blank, using a Shimadzu UV-160A spectrophotometer (Kyoto, Japan) with 1 cm optical path length quartz cuvette. All measurements were taken in triplicates and mean values were calculated.

The concentration of polyphenols in samples was derived from a standard curve of gallic acid ranging from 10 to 50 µg/mL (R<sup>2</sup>= 0.9996).

### ***Evaluation of total antioxidant activity***

Antioxidant activity was investigated by three different in vitro antioxidant methods: 2, 2-diphenylpicrylhydrazyl radical scavenging activity (DPPH), oxygen radical absorbance capacity (ORAC) and Trolox equivalent antioxidant capacity (TEAC) assays.

#### ***DPPH assay***

DPPH (80 µM) was dissolved in absolute ethanol (98%). The mixture was vigorously stirred and allowed to perfecting for 10 minutes at room temperature. 96 well plates were filled with 40 µL samples and 250 µL radical solutions [29, 30]. The decrease of absorption for resulting solutions was monitored at 515 nm for 30 minutes. The results were expressed as mM Trolox / g plant.

#### ***ORAC assay***

AAPH (0.414 g) was dissolved in 10 mL of 75 mM phosphates buffer (pH 7.4) to obtain a final concentration of 153 mM [31]. A stock solution of fluorescein (4 x 10<sup>-3</sup>M) was prepared in 75 mM phosphates buffer solution (pH = 7.4). For experimental measurements the exterior well plates were not used. These well plates were filled with 300 µL of water, while the interior well plates were used for the experimental measurements. Thus, were added 150 µL of the working solution of fluorescein sodium salt in each of the interior well plates. In the control well plates have been added 25 µL of phosphates buffer 75 mM. In the well plates for the standards were introduced 25 µL diluted Trolox and in the sample well plates 25 µL alcoholic extract of the analyzed samples. The samples were incubated for 30 min. for equilibration in the Synergy HT Multi-Detection Microplate TM reader device (Biotek Instruments, Winooski, VT) at 37°C. Reactions were initiated by adding 25 µL AAPH solution. The fluorescence was measured by kinetic monitorization with data taken at each minute at 485 nm excitation filter, 20 nm bandpass, emission filters at 585 nm, 20 nm bandpass.

**TEAC assay**

ABTS stock solution was obtained by the reaction of 7 mM/L aqueous solution and 2.45 mM/L potassium persulfate and a completion in the dark at room temperature for 12-16 h before use. The working solution of ABTS was obtained by ethanol dilution of the stock solution at absorption of  $0.70 \pm 0.02$  AU at 734 nm, verified by Biotek Synergy HT spectrophotometer, USA. 17  $\mu$ L of sample was added to 170  $\mu$ L ABTS solution. The results were expressed as mM Trolox/g plant [25].

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**REFERENCES**

1. K.A. Ross, T. Beta, S.D. Arntfield, *Food Chemistry*, **2009**, 113, 336.
2. S. Nyiredy, *Journal of Chromatography B*, **2004**, 812, 35.
3. P. Mattila, J.J. Kumpulainen, *Journal of Agricultural and Food Chemistry*, **2002**, 50, 3660.
4. M. Kivilompolo, T. Hyotylainen, *Journal of Chromatography A*, **2009**, 1216, 892.
5. R. Japon-Lujan, J.M. Luque-Rodriguez, M.D. Luque de Castro, *Journal of Chromatography A*, **2006**, 1108, 76.
6. R.M. Alonso-Salces, A. Barranco, E. Corta, L.A. Berrueta, B. Gallo, F. Vicente, *Talanta*, **2005**, 65, 654.
7. T.S. Ballard, P. Mallikarjunan, K. Zhou, S. O'Keefe, *Food Chemistry*, **2010**, 120, 1185.
8. X. Jun, S. Deji, Z. Shou, L. Bingbing, L. Ye, Z. Rui, *International Journal of Pharmaceutics*, **2009**, 382, 139.
9. E.M. Silva, H. Rogez, Y. Larondelle, *Separation and Purification Technology*, **2007**, 55, 381.
10. G.A. Akowuah, Z. Ismail, I. Norhayati, A. Sadikun, *Food Chemistry*, **2005**, 93, 311.
11. S. Albu, E. Joyce, L. Paniwnyk, J.P. Lorimer, T.J. Mason, *Ultrasonics Sonochemistry*, **2004**, 11, 261.
12. R.J. Grubestic, J. Vukovic, D. Kremer, S. Vladimir-Knezevic, *Journal of Pharmaceutical and Biomedical Analysis*, **2005**, 39, 837.

13. O. Yesil-Celiktas, P. Nartop, A. Gurel, E. Bedir, F. Vardar-Sukan, *Journal of Plant Physiology*, **2007**, 164, 1536.
14. A. Figueirinha, A. Paranhos, J.J. Perez-Alonso, C. Santos-Buelga, M.T. Batista *Food Chemistry*, **2008**, 110, 718.
15. M.T. Baratta, H.J.D. Dorman, S.G. Deans, A.C. Figueiredo, J.G. Barroso, G. Ruberto, *Flavour and Fragrance Journal*, **1998**, 13, 234.
16. R.N. Bennett, R.M. Wallsgrove, *New Phytologist*, **1994**, 127, 617.
17. D.K. Kliebenstein, *Plant Cell & Environment*, **2004**, 27, 675.
18. A. Vashisth, S. Nagarajan, *Bioelectromagnetics*, **2008**, 29, 571.
19. D. Roux, A. Vian, S. Girard, P. Bonnet, F. Paladian, E. Davies, G. Ledoigt *Planta*, **2008**, 227, 883.
20. A. Vian, D. Roux, S. Girard, P. Bonnet, F. Paladian, E. Davies, G..Ledoigt, *Plant Signaling & Behavior*, **2006**, 1, 67.
21. W. Stankiewicz, M.P. Dabrowski, R. Kubacki, E. Sobiczewska, S. Szmigielski *Electromagnetic Biology and Medicine*, **2006**, 25, 45.
22. I. Lung, M.L. Soran, C. Tudoran, C. Marutoiu, *Central European Journal of Chemistry*, **2013**, 11, 535.
23. I. Lung, M.L. Soran, M. Stan, D. Podar, *Advances in Research*, **2013**, 1, 1-10.
24. T. Katsube, H. Tabata, Y. Ohta, Y. Yamasaki, E. Anuurad, K. Shiwaku, Y. Yamane, *Journal of Agricultural and Food Chemistry*, **2004**, 52, 2391.
25. N. Pellegrini, M. Serafini, B. Colombi, D. Del Rio, S. Salvatore, M. Bianchi, F. Brighenti, *Journal of Nutrition*, **2003**, 133, 2812.
26. H.B. Li, C.C. Wong, K.W. Cheng, F. Chen, *LWT - Food Science and Technology*, **2008**, 41, 385.
27. E. Surducan, V. Surducan, A. Halmagyi, "Process and installation for stimulating plant development in microwaves field" Romanian Patent. RO 125068B1/2012.
28. K. Slinkard, V.L. Singleton, *American Journal of Enology and Viticulture*, **1977**, 28, 49.
29. W. Brand-Williams, M.E. Cuvelier, C. Berset, *LWT - Food Science and Technology*, **1995**, 28, 25.
30. M.B. Arnao, A. Cano, J.F. Alcolea, M. Acosta, *Phytochemical Analysis*, **2001**, 12, 138.
31. D. Huang, B. Ou, M. Hampsch-Woodill, J.A. Flanagan, R.L. Prior, *Journal of Agricultural and Food Chemistry*, **2002**, 50, 4437.

## GUM ARABIC: AN OPTIMIZATION OF ULTRASONIC-ASSISTED EXTRACTION OF ANTIOXIDANT ACTIVITY

AHMED. A. M. ELNOUR<sup>a b\*</sup>, MOHAMED E. S. MIRGHANI<sup>a,c,\*</sup>,  
NASSERELDIN A. KABBASHI<sup>a</sup>, MD Z. ALAM<sup>a</sup>  
AND KHALID HAMID MUSA<sup>d</sup>

**ABSTRACT.** Gum Arabic (GA), also known as *Acacia seyal* gum (ASG), is a dried exudate from trees of *Acacia senegal* and *Acacia seyal*. It provides a rich source of non-viscous soluble fiber with significant health benefits and high antioxidant properties. Tonnes of raw GA are exported annually at a high cost with limited utilization in extraction form. Techniques for the extraction of the bioactive components of GA are available but the high extraction time and the capacity and quality of extraction hinders these procedures. Ultrasonic-assisted extraction is one of the most effective techniques for the recovery of antioxidant and phenolic compounds from ASG. A comparatively low extraction time has been reported for ultrasonication, but the influence of several extraction conditions such as temperature, time and ultrasonic power on the yield of extraction has not been thoroughly studied. This study investigates the optimal ultrasonic extraction conditions for maximum recovery of antioxidant and phenolic compounds from ASG using Response Surface Methodology (RSM) under the Central Composite Design (CCD). Three ultrasonic parameters, namely time in the range of (1-3 hours), power in the range of (1-3 level or 12 to 40 kHz) and temperature from (25-60 °C) were tested for their impact on antioxidant activity. The capacity of the extracts was determined by the scavenging activity of 1, 1-diphenyl-2-picrylhydrazyl (DPPH) radical, ferric reducing antioxidant power (FRAP) assay, and total phenolic compounds (TPC). The results indicated that ultrasonic time, power and temperature had a positive impact on antioxidant capacity and phenolic compounds. The optimum ultrasonic conditions were

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<sup>a</sup> *Bioenvironmental Engineering Research Centre (BERC), Biotechnology Engineering, Faculty of Engineering, International Islamic University, Malaysia (IIUM), P. O. Box 10, Gombak. 50728 Kuala Lumpur, Malaysia.*

<sup>b</sup> *Department of Biochemistry & Gum Processing, Gum Arabic Research Centre, University of Kordofan, Box: 160. Elobied, Sudan*

<sup>c</sup> *International Institute for Halal Research and Training (INHART), IIUM, P. O. Box 10, Gombak. 50728 Kuala Lumpur, Malaysia.*

<sup>d</sup> *Department of Food Science and Human Nutrition College of Agriculture and Veterinary Medicine Qassim university, the kingdom of Saudi Arabia.*

\* *Corresponding author: elwathig@iium.edu.my*

found to be a time of 3 hours, a power of 40 kHz, and a temperature of 42.50°C, under which, forty-eight bioactive compounds from the ASG extract were separated by Gas Chromatography coupled to Tandem Mass Spectrometry (GC-MS/MS).

**Keywords:** *Acacia seyal gum, Bioactive compounds, Gum Arabic, Response Surface Methodology, Ultrasonication.*

## INTRODUCTION

Gum Arabic (GA) is an edible, dried, gummy biopolymer exudates obtained from trees and branches of *Acacia senegal* and *Acacia seyal*. It grows predominantly in the African region of Sahel in Sudan. GA is one of the world's major natural commodities of commerce and constitutes an important export commodity in countries such as Niger, Chad, Senegal, Nigeria, and especially Sudan which controls about 85% of the world's export of GA [1]. Tones of GA are exported annually and has been estimated to the tune of US\$ 40 million yearly over the last 20 years [1, 2]. Despite the fairly high prices, the supply of and demand for GA have declined steadily over the past few decades due to a number of reasons which bothers (aside from the uncontrollable natural phenomenon) on the regulatory policy to ensure food quality and safety of the raw gum[1, 3]. Strict international specification based on the microbiological and chemical characteristics of GA and similar products have been instituted to guarantee the identity, quality, and safety of the raw exudates. This poses several trade challenges.

Gum Arabic (GA) produced by the traditional method exhibit high variability in microbiological quality. In its raw form, it is highly susceptible to bacteriological spoilage especially when stored in warehouses [1]. Traditional solutions to ensure that Gum Arabic reaches the end-user in good quality are expensive.

Spray-drying, for instance, which is a form of pasteurization of the gum through controlled exposure to high temperature, is an energy-intensive and costly procedure estimated at the US \$1000 per ton [1]. Marketing GA to its end users is therefore costly, often requires a long chain of mediators, and mostly require further processing by its end users [4].

Chemically, GA is a complex mixture of macromolecules of different sizes and composition - mainly carbohydrates and proteins. It is rich in non-viscous soluble fibers with high dietary value, and also contains minerals like potassium, magnesium and calcium [5-8].GA antioxidant properties, nephroprotectant, and other effects have been highlighted in recent studies [9-11]. Its role in the metabolism of lipids [12, 13], and its positive effect in the

treatment of several degenerative diseases such as kidney failure [14-16], cardiovascular [17] and gastrointestinal diseases [18, 19] have also been reported. GA, therefore, promises a lot of benefits in medical, food, and pharmaceutical industries.

The major concern about GA, however, is the need to maintain the supply and demand chain of GA in a cost-effective way, and to ensure the purity, quality, and safety of GA that reached the end-users. Extraction of the bioactive components from GA suggests a viable approach to resolve these concerns. A powdered bioactive component of GA extract can prove cost-effective by minimizing tons of exported raw GA, and can also allow standardization of the quality and purity of GA. A big challenge, however, is that sufficient data for an efficient extract procedure of the antioxidant or bioactive component of GA are sparse.

Various techniques exist for extraction of antioxidants from plant materials and other foodstuffs. The techniques commonly adopted are shaking, homogenization at high speed, maceration, and stirring. A comprehensive experimental comparison of this techniques has not been reported, however, some studies have highlighted the drawbacks in these techniques to include low product quality, safety hazards, and prolonged extraction time [20].

Recently, novel extraction techniques such as microwave-assisted, ultrasonic-assisted, and enzyme-assisted super-critical fluid extraction technique have also been developed for extraction of antioxidants from plant materials [21-23]. The novel techniques have shown remarkable improvement in the extraction process but have also come with some drawbacks. For instance, the enzyme-assisted supercritical extraction is reported to be highly eco-friendly but requires a low range of temperature at a high cost [24]. The incurred high cost appears the major challenge in these techniques.

The ultrasonic-assisted extraction (UAE) is overall considered the most economical and efficient extraction methods of the newly introduced techniques [25-29]. It has been successfully applied for the extraction of antioxidant from plant materials; however, its implementation depends significantly on the condition of extraction. An appropriate choice of UAE extraction parameters such as ultrasonic power, temperature, and time should reasonably influence the yield of extraction.

This study aims to design an optimized ultrasonic-assisted extraction for the preparation of antioxidant powder from raw gum arabic using response surface method (RSM). Response surface method (RSM) is a mathematical and statistical tool, which has been widely used to optimize various parameters in process industries [30]. It has been used recently to determine the most influential parameters for simultaneous production of lactic acid, xanthan and

ethanol from molasses [31-34]. RSM can evaluate the impact of different multiple parameters, and simultaneously optimize experimental conditions [35-37]. Different RSM methods such as box–Behnken design (BBD), central composite design (CCD) and three-level full factorial designs (TFFD) have been used widely in various fields such as food, biology and chemistry. Although the CCD, which has been proven for its accurate fitness in different models and experiments, is more commonly adopted for RSM [32].

In this experimental study, UAE parameters namely power, time and temperature of extraction were optimized by RSM, adopting the central composite design (CCD), to obtain optimal extraction of total phenolic contents and antioxidant activity from ASG. To the best of the Authors' knowledge, this is the first time the RSM has been deployed to optimize UAE parameters for optimal extraction of total phenolic compounds and antioxidant activity from GA. We also identified the composition of the extracts by GC-MS/MS and assessed the influence of ultrasound on the extraction efficiency and chemical compositions of the extracts.

## RESULTS AND DISCUSSION

### FITTING THE MODEL

Twenty experiments were conducted under different conditions of ultrasonic extraction. Overall, the antioxidant activities or responses for the independent variables of ultrasonic-assisted extraction were recorded as shown in Table 1. The experiment labelled '8' with a total ultrasonic extraction time of 3h, extraction power of 40kHz, and temperature of 60 °C was found to produce the highest DPPH inhibition of 89.2%, FRAP of 26646 mg TE/100g DW, and total phenolic content (TPC) of 25145mg GAE/100g DW. In contrast, the experiment labelled '1' with an extraction time of 1h, power of 12kHz, and temperature of 25 °C produced the least responses with DPPH inhibition of 35.5%, FRAP of 6430 mg TE/100g DW and TPC of 4694 mg GAE/100g DW.

### ANALYSIS OF VARIANCE

In the prediction of optimized ultrasonic assisted extraction conditions for the DPPH, FRAP and TPC assays, it is important to check the fitting of the RSM mathematical model to ensure its reliability. Different techniques for analysis of variance, such as lack of fit;  $R^2$ ; Predicted Residual Sum of Square (PRESS) for models; F-ratio; and Prob > F methods were analyzed to identify the fitting of the RSM mathematical models. "Lack of fit" assesses the model for appropriate effects when the test is conducted.  $R^2$  estimates the proportion of

variation in the response that can be attributed to the model rather than to random errors. An  $R^2$  value near 1 indicates that the model is a good predictor of the response. PRESS shows how well the predictive model fits each point in the design. The F-ratio indicates whether the model differs significantly from a model where all predicted values are the response mean. Prob > F measures the probability of obtaining an F-ratio as large as what is observed, given that all parameters except the intercept are zero. Small values of Prob > F indicate that the observed F-ratio is unlikely. Such values are considered evidence that there is at least one significant effect in the model [30].

**Table 1.** Antioxidant activities, of the extract of *A. seyal*/gum under different conditions of ultrasonic-assisted extraction based on a central composite design (CCD) for response surface analysis.

Run	Extraction Condition			Analytical results		
	X <sub>2</sub> Extraction time (hours)	X <sub>2</sub> Extraction power (kHz)	X <sub>2</sub> Extraction Temperature (°C)	DPPH (Inhibition %)	Antioxidant Activity FRAP (mg TE/100g DW)	Total phenolic Content (mg GAE/100g DW)
1	1 (-1)	12 (-1)	25 (-1)	35.5	6430	4694
2	3 (1)	12 (-1)	25 (-1)	43.2	7607	5876
3	1 (-1)	40 (1)	25 (-1)	46.8	8254	5862
4	3 (1)	40 (1)	25 (-1)	57.2	17183	11798
5	1 (-1)	12 (-1)	60 (1)	58.4	10298	6770
6	3 (1)	12 (-1)	60 (1)	83.3	26646	18307
7	1 (-1)	40 (1)	60 (1)	56.5	11369	8963
8	3 (1)	40 (1)	60 (1)	89.2	26646	25145
9	1 (-1.68179)	26 (0)	42.5 (0)	55.4	9860	7406
10	3 (1.68179)	26 (0)	42.5 (0)	70.7	17451	12768
11	2 (0)	12 (-1.68179)	42.5 (0)	78.2	15740	12201
12	2 (0)	40 (1.68179)	42.5 (0)	80.9	14592	12194
13	2 (0)	26 (0)	25 (-1.68179)	81.5	13452	10814
14	2 (0)	26 (0)	60 (1.68179)	86.2	24995	17413
15	2 (0)	26 (0)	42.5 (0)	56.6	23141	15575
16	2 (0)	26 (0)	42.5 (0)	79.9	18560	13622
17	2 (0)	26 (0)	42.5 (0)	83.4	19887	13704
18	2 (0)	26 (0)	42.5 (0)	83.9	17380	13342
19	2 (0)	26 (0)	42.5 (0)	79.7	19158	13745
20	2 (0)	26 (0)	42.5 (0)	55.3	16503	12133

\* All results are the means  $\pm$  SD (n = 3).

Tables 2 - 4 present the analysis of variance for fitting the mathematical models used in the prediction of optimized ultrasonic extraction conditions. The results showed that the coefficient of determination ( $R^2$ ) of the models for all three responses (DPPH, FRAP and TPC) was close to 1 (ranging between 0.8117 and 0.9583), which imply that the mathematical models are reliable predictors for DPPH, FRAP and TPC assays. The  $R^2$  values indicate that at least 81% of the actual values were matched with the predicted values proposed by the mathematical models. The results also outlined that “lack of fit” of the models for DPPH, FRAP and TPC were all significantly higher than 0.05 indicating that the models had the appropriate effects when the experiments were conducted. In addition, the “Prob > F” values were found to be in the range of 0.0001 and 0.9988, and the F-ratio of the models were found to be low which further confirm that the RSM models adopted in the study were reliable in the prediction of optimal ultrasonic extraction conditions for DPPH, FRAP and TPC.

**Table 2.** ANOVA for DPPH fitted quadratic polynomial model of extraction parameter

Source	Sum of Squares	DF	Mean Squares	F – (Ratio) Value	p-value	Significant
<b>Model</b>	4184.98	6	697.5	9.34	0.0004**	Significant
$x_1$ -time	825.07	1	825.07	11.05	0.0055*	
$x_2$ -power	102.3	1	102.3	1.37	0.2628	
$x_3$ -temperature	1566.66	1	1566.66	20.98	0.0005**	
$x_1x_2$	195.26	1	195.26	2.62	0.1298	
$x_2x_2$	57.03	1	57.03	0.76	0.398	
$x_1^2$	1641.69	1	1641.69	21.99	0.0004**	
<b>Residual</b>	970.55	13	74.66			
<b>Lack of Fit</b>	108.4	8	13.55	0.079	0.9988	Not significant
<b>Pure Error</b>	862.15	5	172.43			
<b>Cor Total</b>	5155.53	19				
<b>R-squared</b>				0.8117		
<b>Adj R-squared</b>				0.7249		

\*(C. V= 12%), PRESS=1532.95.

\* $P \leq 0.05$  indicates the model terms are significant.

\*\* $P \leq 0.01$  indicates the model terms are highly significant.

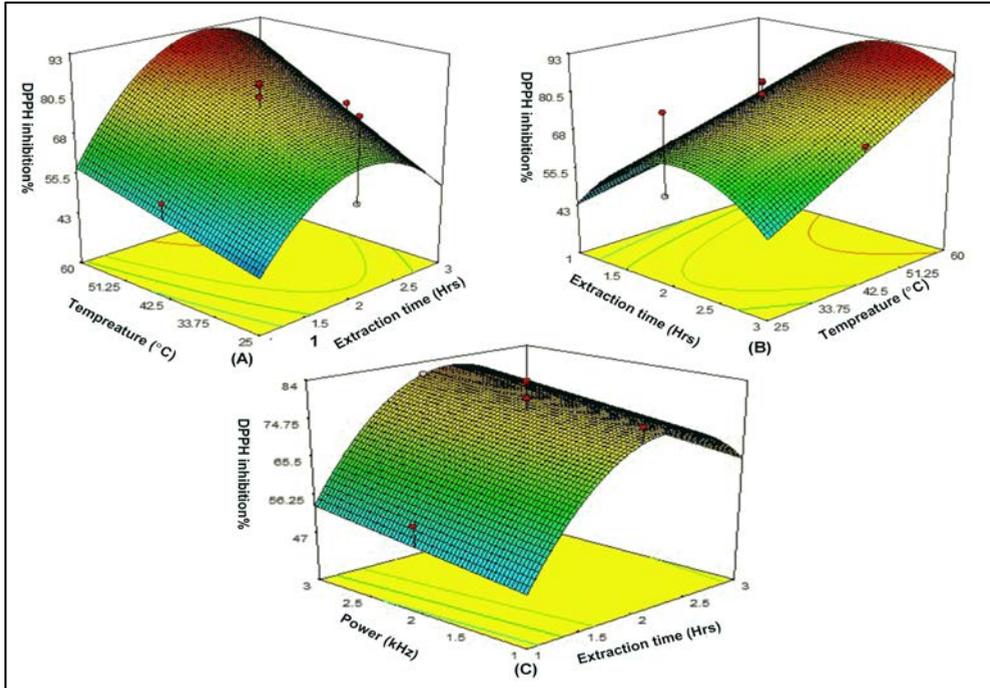
## THE DPPH RESPONSE SURFACE ANALYSIS OF ANTIOXIDANT ACTIVITY

The effects of factors such as extraction time, power, and temperature, denoted  $x_1$ ,  $x_2$  and  $x_3$  respectively in equation (3) were carefully studied. The significance of each coefficient was determined by F-values and p-values. The regression equation in coded level, neglecting insignificant terms, was generated as:

$$DPPH\% = +77.78 + 9.08x_1 + 3.20x_2 + 11.99x_3 + 4.94x_1x_3 - 2.67x_2x_3 - 18.16x_1^2 \quad (3)$$

Results indicated that the quadratic relationship between the DPPH inhibition and extraction factors have a good regression coefficient ( $R^2=0.8117$ ). Higher F-value with lower p-value always led to more significant correspondence amongst the independent variables. The terms  $x_1$ ,  $x_2$ ,  $x_3$ ,  $x_1x_3$  and  $x_1^2$  were significant with a p-value less than 0.05. However,  $x_1$ ,  $x_2$  and  $x_2^2x_3^2$ , were not significant due to a higher p-value (i.e. higher than 0.05).

Figure 1 showed the complex interaction between time, temperature, and extraction power. The highest anti-scavenging activity (DPPH) was observed at both higher time of extraction and lower temperature (see Figure 1A). However, the increase in temperature at a fixed extraction time led to an increase in the extraction of antioxidant activity using DPPH assay. The extraction reached a maximum but at the lowest temperature tested. Figure 1B showed the interaction between the temperature and time of extraction on the antioxidant scavenging (DPPH %). An increase in the antioxidant activity was noticed as the extraction temperature increased up to 60 °C, but decreased thereafter. A significant increase in the antioxidant capacity was also observed at high ultrasonic power. However, the trend was reversed as the ultrasonic power reached the third level at 40 kHz (see Fig 1C). High temperature might decrease the antioxidants activity due to the increase of thermal effect or due to degradation of bioactive compounds. High temperatures degrade phenolic compounds, leading to a reduction in the antioxidants [32, 38, 39].



**Figure 1.** 3D Response surface plots showing the operating parameter effect on antioxidant capacity. (A) The DPPH % vs. extraction time of 3 h and ultrasonic power of 40kHz at fixed temperature of 42.50 °C; (B) The DPPH vs. temperatures °C and extraction time of 3 h, at ultrasonic power of 40kHz; (C) The DPPH% vs extraction time of 3h and ultrasonic power of 40kHz at fixed temperature of 42.50 °C.

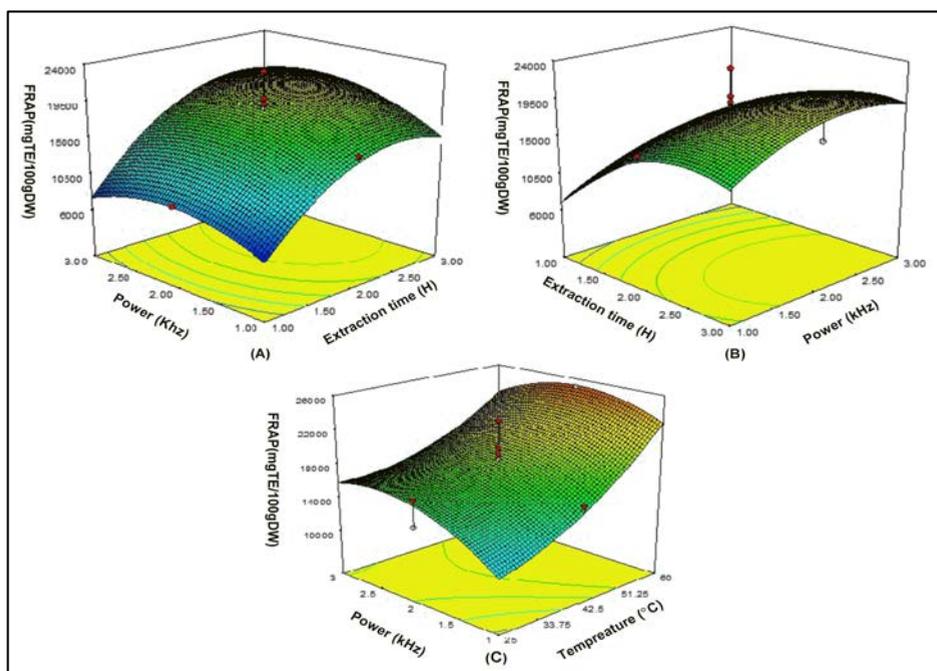
## THE FRAP RESPONSE SURFACE ANALYSIS OF ANTIOXIDANT ACTIVITY

Results for FRAP response surface analysis of the antioxidant activity of *Acacia seyal* gum (ASG) is presented in Table 3. The regression value ( $R^2 = 0.9234$ ) indicates a good relationship between the antioxidant activity of *Acacia seyal* gum (ASG) and the extraction parameters (time, power and temperature). The fitting quadratic polynomial equation of the data with only significant terms is given in equation (4).

$$\begin{aligned}
 FRAP \text{ (mgTE/100gDW)} = & \\
 & 18763.56 + 6932.07x_1 + 1132.07x_2 + 4646.15x_3 + 834.98x_1x_2 \\
 & + 2689.90x_1x_3 - 1291.14x_2x_3 - 4029.61x_1^2 - 2519.61x_2^2 \\
 & + 1820.71x_3^2
 \end{aligned} \tag{4}$$

where the variables are as defined as in table (3)

Figure 2 shows the relationship between the antioxidant activity and the extraction parameters. The effect of varying extraction time, ultrasonic power and their mutual interaction on the antioxidant activity are shown in Figure 2A. Antioxidant activity increased with increasing ultrasonication power but at high extraction time. The highest antioxidant activity was however observed at extraction time of 3 hours and ultrasonic power of 40 kHz (levels 3). The effects of extraction time, power and their interaction on antioxidants activity were shown in Figure 2B.



**Figure 2.** 3D Response surface plots showing the operating parameter effect on antioxidant capacity. (A) The FRAP vs. extraction time of 3 h and ultrasonic power of 40 kHz at fixed temperature of 42.50 °C; (B) The FRAP vs. ultrasonic power of 40 kHz and Extraction time of 3 h at fixed temperature of 42.50 °C; (C) The FRAP vs extraction temperature of 42.50 °C at fixed extraction time of 3 h.

Figure 2C shows the effect of varying the extraction power and temperature on the antioxidants activity. Notice that, at low temperature, less than 42.50 °C, increasing the extraction power resulted in the high amount of antioxidant extraction from ASG. Whereas, at an overheated temperature (greater than 42.50 °C), the reverse trend is noticeable. Moreover, higher extraction temperature can produce increasingly repulsive diffusion-solvent interaction, leading to a reduction of antioxidants in the extract. The highest antioxidant activity of ASG extracts was therefore determined at 42.50 °C for 3 hours.

**Table 3.** ANOVA for FRAP fitted quadratic polynomial model of Extraction parameter

Source	Sum of Squares	DF	Mean Squares	F -Value	p-value	Significant
<b>Model</b>	6.62E+08	9	7.35E+07	13.39	0.0002**	Significant
$x_1$ -time	2.43E+08	1	2.43E+08	44.3	< 0.0001**	
$x_2$ -power	1.28E+07	1	1.28E+07	2.34	0.1575	
$x_3$ -temperature	2.31E+08	1	2.31E+08	42.12	< 0.0001**	
$x_1x_2$	5.58E+06	1	5.58E+06	1.02	0.3373	
$x_1x_3$	5.79E+07	1	5.79E+07	10.54	0.0088**	
$x_2x_3$	1.33E+07	1	1.33E+07	2.43	0.1502	
$X_1^2$	4.73E+07	1	4.73E+07	8.62	0.0149*	
$x_2^2$	1.85E+07	1	1.85E+07	3.37	0.0962	
$x_3^2$	1.11E+07	1	1.11E+07	2.01	0.1864	
<b>Residual</b>	5.49E+07	10	5.49E+06			
<b>Lack of Fit</b>	3.14E+07	5	6.29E+06	1.34	0.3783	Not significant
<b>Pure Error</b>	2.35E+07	5	4.70E+06			
<b>Cor Total</b>	7.17E+08	19				
<b>R-squared</b>				0.9234		
<b>Adj R-squared</b>				0.8544		

\* CV=14.41%. PRESS=374100000.

\*P≤0.05 indicates the model terms are significant.

\*\*P≤0.01 indicates the model terms are highly significant.

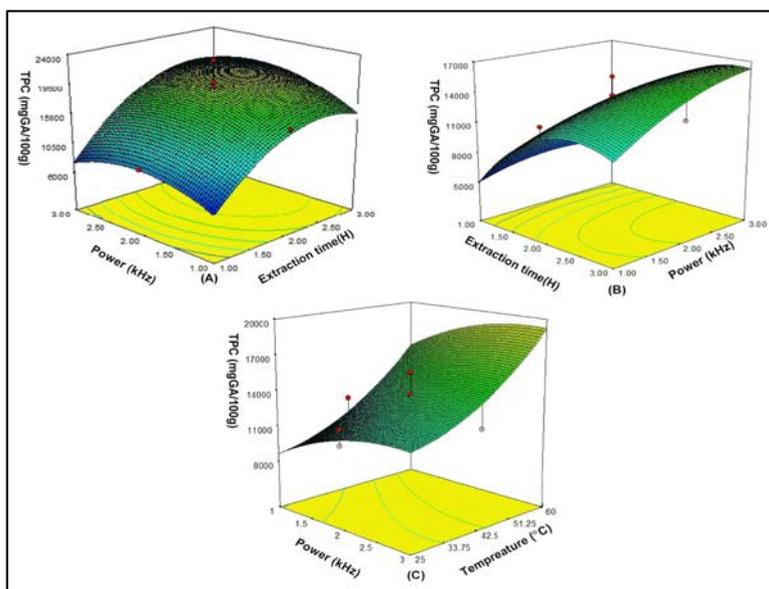
## THE TPC RESPONSE SURFACE ANALYSIS OF ANTIOXIDANT ACTIVITY

For Total Phenolic Content (TPC), the fitting regression equation in coded level, neglecting insignificant terms was generated as given in equation (5).

$$\begin{aligned}
 \text{TPC (mgGAE/100gDW)} = & \\
 & +13650.06 + 4019.18x_1 + 1611.34x_2 + 3671.68x_3 + 1174.86x_2x_3 + 2575.14x_1x_3 \\
 & -3129.30x_1 - 1018.64x_2^2 + 1316.29 + 1316.29x_3^2
 \end{aligned}
 \tag{5}$$

Where; the variables are as defined as in Table (4).

Results from experimentation (see, Table 4) indicate that the quadratic relationship between TPC and the extraction factors have a good regression coefficient ( $R^2 = 0.8117$ ). Terms with  $x_1x_2$  and  $x_1^2$  were not significant due to a higher p-value (i.e. higher than 0.05) and were eliminated. Figure 3 shows the plot of the complex interaction between extraction time, ultrasonication power, and extraction temperature.



**Figure 3.** 3D response surface plots showing the operating parameter effect on antioxidant capacity. (A) The TPC (mg GAE/100g DW) vs. extraction time of 3 h and Ultrasonic power of 40kHz at fixed temperature of 42.50 °C; (B) The TPC (mg GAE/100g DW) vs. ultrasonic power of 40kHz and extraction time of 3 h at fixed temperature of 42.50 °C; (C) The TPC (mg GAE/100g DW) vs extraction temperature 42.50 °C and ultrasonic power of 48kHz at fixed extraction time of 3h.

The highest total phenolic contents were observed at higher extraction time and ultrasonication power as shown in Figure 3A. Increase in both ultrasonication power and extraction time under a fixed temperature resulted in an increase in the total phenolic contents. However, a maximum is reached at the lowest extraction temperature in the range of temperature studied.

Figure 3B shows the interaction between ultrasonication power and extraction time on total phenolic extraction. Low total phenolic contents were observed for extraction time less than 3 hours but increased for higher time duration. A slight increase in the total phenolic contents at the low ultrasonic power and at higher extraction temperature was also noticed, however, the trend was reversed as the extraction temperature approached 42.50 °C (see Figure 3C). High temperature can reduce the polyphenolic contents as reported in the work of MA, Chen [40] which attributes the reduction to thermal degradation or polymerization reaction of phenols. Also, low ultrasonic power can result in less antioxidant activity due to low cavitations bubble size which has a negative impact on external and internal resistance to mass transfer of gum extraction [41].

**Table 4.** ANOVA for TPC fitted quadratic polynomial model of Extraction parameter.

Source	Sum of Squares	DF	Mean Squares	F – (Ratio) Value	p-value	Significant
<b>Model</b>	4184.98	6	697.5	9.34	0.0004**	Significant
$x_1$ -time	825.07	1	825.07	11.05	0.0055*	
$x_2$ -power	102.3	1	102.3	1.37	0.2628	
$x_3$ -temperature	1566.66	1	1566.66	20.98	0.0005**	
$x_1x_2$	195.26	1	195.26	2.62	0.1298	
$x_2x_2$	57.03	1	57.03	0.76	0.398	
$x_1^2$	1641.69	1	1641.69	21.99	0.0004**	
<b>Residual</b>	970.55	13	74.66			
<b>Lack of Fit</b>	108.4	8	13.55	0.079	0.9988	Not significant
<b>Pure Error</b>	862.15	5	172.43			
<b>Cor Total</b>	5155.53	19				
<b>R-squared</b>				0.8117		
<b>Adj R-squared</b>				0.7249		

\*C. V=10.81%; PRESS=453100000.

\*P≤0.05 indicates the model terms are significant.

\*\*P≤0.01 indicates the model terms are highly significant

## OPTIMIZATION AND VALIDATION OF ULTRASONIC EXTRACTION CONDITIONS

Based on the foregoing mathematical models, the predictions of optimal ultrasonic conditions for antioxidant activity of ASG (using DPPH, FRAP, and TPC assay) are a time of 3hours, a power of 40 kHz and a temperature of 42.50 °C. To validate these predictions, the actual experiment

was conducted keeping the ultrasonic conditions at these optimal. The experiment was repeated three times and the results averaged. Table 5 gives the comparison between the actual and predicted the antioxidant activity of ASG. The results indicate a close similarity between the experiment and the predicted values ( $p \geq 0.05$ ), with prediction errors approximately 6.2% for both DPPH and FRAP assays and 14% for TPC. This shows that for maximum recovery of polyphenolic compounds and antioxidant capacity from ASG, the applied ultrasonic conditions can be considered optimal. For both prediction and experiment, the antioxidant activities observed using FRAP was larger than that of TPC. The DPPH inhibition was less than that for FRAP and TPC, despite DPPH being a more stable antioxidant assay [42, 43].

**Table 5.** Validation of the predicted values for antioxidant properties

Antioxidant activity	Values	
	Predicted	Experimental (n=3)
DPPH (inhibition %)	74.9708±9.91 <sup>a</sup>	70.55±0.97 <sup>a</sup>
FRAP (mgTE/100g)	20499.4±3006.25 <sup>a</sup>	19303.54±231 <sup>a</sup>
TPC(GAE/100g)	17714±1677.75 <sup>a</sup>	15406.97±130.05 <sup>a</sup>

All the values are means  $\pm$  standard deviations and those in the same row not sharing the same superscript letter (a) are significantly different from each other ( $p \leq 0.05$ ).

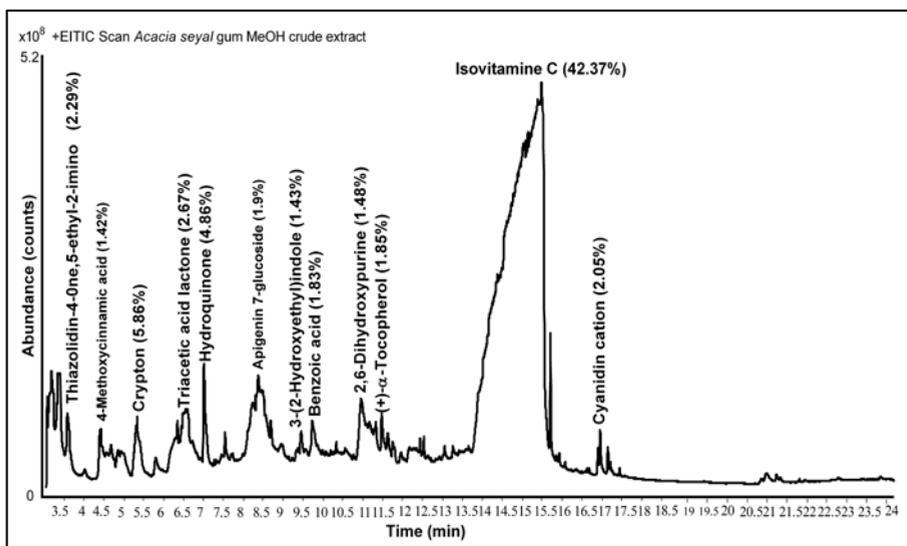
## CHEMICAL COMPOSITION OF OPTIMIZED ASG ANTIOXIDANT EXTRACT

Forty-eight components from the ASG extract were separated using the predicted optimum conditions as shown in Figure 4. The major components identified by GC-MS/MS were Isovitamin C (42.37%), Crypton (5.86%), Hydroquinone (4.8%), and Thiazolidin-4-one,5-ethyl-2-imino (2.49%). Table 6 presents the complete list of bioactive components from the ASG extract.

Similar studies were reported which consider the total phenolic contents and antioxidant values [32, 44, 45], however, unlike the current study, there was the detectable amount of hydroxybenzaldehyde, luteolin/ kaempferol, feruloyl-arabinose-arabinose and caffeoyltartaric acid which are not present in this experiment. The amount of isovitamin C determined in the current study was higher than the amount reported by Fiorito et al [45]. Overall, some bioactive components namely Cyanidin cation, gallic acid, chromone, 5-hydroxy-6,7,8-trimethoxy-2,3-dimethyl-, 2,3,5,5,8a-Pentamethyl-6,7,8a-tetrahydro-5H-chromen-8-ol identified in this study have not been reported in previous literature.

**Table 6.** The chemical composition of the *Acacia seyal* gum extracts using optimum conditions to detect the bioactive compounds achieved by GC-MS/MS.

NO	RT (min)	Name of Bioactive compounds	Area sum %	NO	RT (min)	Name of Bioactive compounds	Area sum %
1	3.106	4-Methylcatechol	1.42	25	12.437	Coniferyl aldehyde	1.3
2	3.161	2,5-Diamino-4,6-dihydropyrimidine	1.65	26	12.531	2,4-Di-tert-butylphenol	1.1
3	3.564	Thiazolidin-4-One,5-ethyl-2-imino	2.49	27	13.036	o-Cresol,6-tert-butyl	0.9
4	4.032	Albuterol	0.47	28	15.024	2,6-Dimethylol-p-cresol	2.16
5	4.545	4-Methoxycinnamic acid	1.11	29	15.528	Isovitamine C	42.37
6	4.694	Acetophenone,4'-ethyl	0.91	30	15.708	Cyanidin cation	2.05
7	4.765	Sinapyl alcohol	0.98	31	15.87	1,4-Naphthoquinone, 2-acetyl-3-hydroxy-5,6,8-trimethoxy-	0.64
8	5.302	Crypton	5.86	32	15.94	Fisetin	0.74
9	5.782	Isopinocampheol	0.98	33	16.087	Ferulic acid	0.63
10	6.292	4-Mercaptophenol	0.76	34	16.392	Resveratrol	0.7
11	6.539	Triacetic acid lactone	2.67	35	16.581	$\beta$ -Citronellol	0.71
12	7.003	Hydroquinone	4.86	36	16.908	Dihydrocarvone	0.54
13	7.519	ENDO, ENDO-2,3-BORNANEDIOL	0.61	37	16.972	Patchoulol	1.21
14	7.816	Isobornyl acetate	1.05	38	17.132	5,7,3',4'-Tetrahydroxyflavone	0.61
15	8.332	Apigenin 7-glucoside	1.9	39	17.211	Chromone,5-hydroxy-6,7,8-trimethoxy-2,3-dimethyl-	0.42
16	8.653	Dihydrouacil	1.15	40	17.43	$\alpha$ -Bisabolol	0.65
17	8.879	Phloroglucinol	0.63	41	17.312	Isolongifolol	0.54
18	9.343	Cumaldehyde	0.77	42	20.974	Genistin	0.67
19	9.416	3-(2-Hydroxyethyl) indole	1.34	43	21.063	Glycitein	0.67
20	9.682	Benzoic acid	1.83	44	21.389	Quercetin	0.44
21	10.332	4-Hydroxyphenylpyruvic acid	0.68	45	21.491	Vanylglycol	0.44
22	11.471	2,6-Dihydropyrimine	1.48	46	22.043	Quercetin 3-D-galactoside	0.54
23	11.682	(+)- $\alpha$ -Tocopherol	1.52	47	22.959	Propyl gallate	0.58
24	12.14	$\beta$ -Resorcylaldehyde	0.93	48	23.995	2,3,5,5,8a-Pentamethyl-6,7,8a-tetrahydro-5H-chromen-8-ol	0.62



**Figure 4.** Chromatogram of *Acacia seyal* gum extract (Rt, 7, 8.5, 11, and 15.5 min) respectively, using optimum conditions to detect the bioactive compounds achieved by GC- MS/MS.

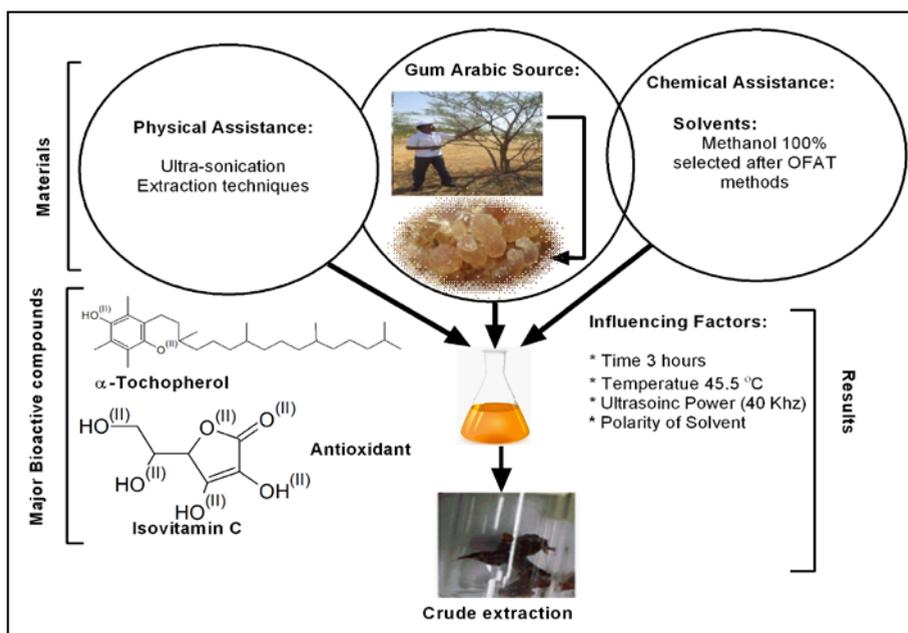
## CONCLUSIONS

In this study, the response surface methodology (RSM) using composite central design (CCD) technique was effectively deployed for the optimization of the ultrasonic extraction parameters in the extraction of antioxidants and functional components from *Acacia seyal* gum. Ultrasonic temperature, power, and time of extraction were found to have different degrees of impact on the extraction process. The parameters significantly affected the response of DPPH, FRAP, and TPC assays. However, temperature increase was found to be insignificant but rather impacted negatively on the antioxidant activity. The optimum ultrasonic extraction conditions for the maximum recovery of antioxidant activity from ASG extract were found to be a time of 3 hours, a power of 40 kHz (3-level), and temperature of 42.50 °C. As a result, forty-eight bioactive compounds were separated from the ASG extract which demonstrates a significant improvement in previous works. Overall, the study has also demonstrated the importance of ultrasonic-assisted extraction (UAE) which is a green approach to the preparation of rich natural antioxidants and functional components from ASG. It is hoped that this study will provide a sufficient foundation for the replacement of synthetic antioxidants. There could be enormous potential for direct utilization of the bioactive component of *Acacia seyal* gum in the food and pharmaceutical industries.

## EXPERIMENTAL SECTION

### MATERIAL PREPARATION

Raw gum exudates used in this study were collected from the Blue Nile State of Sudan and validated for identity and purity by experts from the Sudanese Ministry of Forestry and Agriculture. To ensure the homogeneity of the samples, each nodule was randomly selected and prepared in two pieces, and each piece was rendered into powder form by mechanical grinding and sieving using the U.S.A standard testing sieve (Fisher Company) with 1.40 mm (0.0555 inches) mesh size. By using Glossaries (DHAUS sensitive balance with 20 ml capacity in each vial), 1gram of gum powder was measured from the powder samples and a 10 mL of absolute methanol added to it in a vial. All vials containing samples and solvent were then placed inside the Ultrasonic Water Bath (JAC, SN: RD02AB109x) and centrifuged for 10 minutes at a stirring speed of 1000 rpm. At the end of the procedure, a filtering process is applied to the clarified suspension using Sartorius PTEF 0.45  $\mu\text{m}$  filter and the supernatants were stored in a freezer below  $-25\text{ }^{\circ}\text{C}$  until further analysis.



**Figure 5.** Gum Arabic (*Acacia seyal* gum), techniques of crude extraction and characterization.

## CHEMICALS AND REAGENTS

The chemicals and reagents used in the study include folin–ciocalteu phenol reagent, ferric chloride ( $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ ), and HCl obtained from Merck (Darmstadt, Germany). Others include 2, 2-diphenyl-1-picrylhydrazyl (DPPH), 2, 4, 6-tris (2- pyridyl)-s-triazine (TPTZ), Gallic acid, Trolox, and sodium acetate trihydrate from Sigma (USA). Sodium carbonate from RDH (Germany) and glacial acetic acid from Mallinckrodt Baker (USA) was also used. All chemicals and reagents were of analytical grade. Spectrophotometric measurements were done using Spectro Star Nano-spectrophotometer with microplate which contained 96 micro-cuvettes.

## ULTRASONIC-ASSISTED EXTRACTION (UAE)

For the extraction of the bioactive component using UAE, methanol is chosen as the extraction solvent based on cues from preliminary studies which indicated that methanol gives the highest extraction of polyphenols and antioxidant activities. A sample-to-solvent ratio of 1 gram in 10 mL is adopted to minimize the energy required for heating up the solvent, and also to minimize the energy utilized during solvent removal in the drying process to obtain the bioactive gum powder. All the ultrasonic extraction processes were conducted using an ultrasonic bath (JAC 1505, AC220 V, 60 Hz, 280w, SN: RD 02AB109x, Korea) at the pre-determined conditions designed by the Response Surface Methodology for time, power and temperature. After the extraction process is completed, the extracts are transferred immediately to an ice bath to cool to room temperature. Consequently, a filtering procedure is applied to the clarified suspension by means of a Sartorius PTEF 0.45  $\mu\text{m}$  filter, and the supernatants are stored in a freezer below  $-25\text{ }^\circ\text{C}$  until the analysis was conducted.

## EXPERIMENTAL DESIGN

The experimental design in this study is based on the response surface methodology (RSM) technique. RSM explores, by careful experimental design, the influence of several independent variables (input variables) on one or more response variables (output variables) with the objective of obtaining optimized output responses. Under the RSM, the popular Box-Wilson procedure, commonly called Central Composite Design (CCD) is adopted to evaluate the relevance of the three controllable factors (namely ultrasonic power, temperature and ultra-sonication duration) in the extraction process, and to identify eventual interactions between the variables.

CCD consists of a set of factorial or fractional factorial designed with center points for estimation of curvatures. A two-level full factorial design (coded  $\pm 1$ ), superimposed by center points (coded 0) and “star points” (coded  $\pm a$ ) can be used which permits shortening of the number of experiments.

“Star points” are axial experiments located on variable axes at a distance from the reference center. They establish new extremes for the parameters of the factors involved which provide the basis for the estimation of curvatures for the model. The value of star points is a function of the desired properties of the experimental design and also of the number of experiments involved in the model. A virtual cube can be imagined where each axis of the cube corresponds to a variable under study. In the experimental analysis, the central composite design points describe a sphere around the factorial cube. Preliminary experiments allow pointing the variables at five different coded levels:  $-\alpha$  ( $=1.68$ ),  $1$ ,  $0$ ,  $+1$ ,  $+\alpha$  ( $=+1.68$ ).

A total of 20 experiments were conducted including six replications at the center point to evaluate experimental error. The independent variables are the extraction temperature, 25–60 °C, extraction time, 1-3 hours, and ultrasonic power was 12–40 kHz (or Low to high level coded 1-3). The coded values of the independent variables for CCD are shown in Table 7. Total phenolic (TPC), DPPH and FRAP were selected as the response of the design experiments (Y). Table 7 shows the coded and actual values for CCD of the independent variables.

The model equations, 3D graph plots, and the 2D contour plots were developed using the design of experiment (DOE) software version ® 7.0.0. The software also allows predicting the optimum conditions of the independent variables. A second order equation given in (1) was used to express the level of antioxidant activity and the content of phenolic compounds as a function of the independent variables.

$$y = \beta_0 + \sum_{i=1}^2 \beta_{ix_i} + \sum_{i=1}^3 \beta_{iix_i^2} + \sum_{i=1}^2 \sum_{i=i+1}^3 \beta_{ijx_ix_j} \quad (1)$$

Y in the equation is the predicted response which represents the level of antioxidant activity (measured in mg TE or GAE/100g);  $\beta_0$  represent the regression coefficients or intercept;  $\beta_i$  models the linear effect of the variables,  $\beta_{ii}$  represents the squared effect;  $\beta_{ij}$  denotes the interaction effect; while  $x_i$  and  $x_j$  represent the independent variables affecting the output responses. The experimental data were analysed by the Design-Expert software ® [7.0.0] and the coefficients estimated by means of their F-value. Consequently, to obtain the optimum experimental conditions of total phenolic contents and the level of antioxidant activity, statistical analyses were performed namely analysis of variance (ANOVA), regression analysis, and plotting of response surface figures.

**Table 7.** The range of coded and actual values for central composite design

Independent variables	Code units	Coded levels		
		-1	0	1
Extraction Time (Hours)	$x_1$	1	2	3
Power (kHz)	$x_2, x_2$	12	26	40
Temperature (°C)	$x_3$	25	42	60

## ANTIOXIDANT ACTIVITY DETERMINATION

### DETERMINATION OF FOLIN-CIOCALTEU INDEX FOR TPC

For determination of TPC, the folin-Ciocalteu index (FCI) assay was used. The procedure adopted follows the method described by [46, 47]. Approximately 0.5 mL diluted Folin-Ciocalteu reagent was added to 100  $\mu$ L sample extracts and allowed to set for 5 minutes before addition of 1 mL (7.5%) of sodium carbonate (w/v). The absorbance was taken at 765 nm wavelength using the spectrophotometer after 2 h, and the result recorded in terms of mg of Gallic acid equivalent (GAE).

### RADICAL SCAVENGING ACTIVITY (DPPH)

A 2, 2-diphenyl-1-picrylhydrazyl (DPPH) was used to evaluate the antioxidant activity. This is based on the method presented by Musa, Abdullah, Kuswandi, and Hidayat [48]. The DPPH was freshly produced by liquefying 40 mg DPPH in 1000 ml of methanol to get a  $1.00 \pm 0.01$  unit of absorbance in a spectrophotometer (Spectro Nanostar, Germany) at 517 nm wavelength. Before keeping it in the dark for 2 hours, approximately 100  $\mu$ L of sample were mixed up with 1 ml of the DPPH solution. Equation (2) presents the determination rules of the DPPH scavenging activity.

$$\% DPPH_{sc} = (A_{con} - A_{sample}) \times 100 / A_{con} \quad (2)$$

Where;  $A_{con}$  and  $A_{sample}$  represent the absorbance of the control and sample respectively.

### DETERMINATION OF FERRIC REDUCING ANTIOXIDANT POWER (FRAP)

A working fresh FRAP reagent was prepared as described by [48] with some slight modification. The working reagent is formulated by mixing 300 mm acetate buffer having pH3.6 (constituted by 3.1 g sodium acetate trihydrate plus 16 ml glacial acid) with distilled water in a ratio 1:1; plus 10 mm TPTZ (2,4,6-tris (2-pyridyl)-s-triazine) in 40 mm HCL; and with 20 mm  $FeCl_3 \cdot 6H_2O$  in the

ratio of 10:1:1. After 30 minutes, about 1 ml of FRAP reagent was added to 100  $\mu$ L of samples. By using a spectrophotometer, the absorbance was placed at 595 nm wavelength, and the experimental result presented in milligram (mg) of Trolox equivalent (TE) per 100 g of fresh sample (mg TE/100 g of FW).

### GC-MS/MS ANALYSIS

GC-MS/MS analysis was used to identify compounds in the extracted *Acacia seyal* gum (ASG), in accordance with the method reported by Stankov-Jovanović et al. [49]. The analysis was carried out using a GC (from Agilent Technologies 7890A) interfaced with a mass-selective detector (MSD) (Agilent 7000 Triple Quad). The MSD is equipped with Agilent HP-5ms (5%-phenyl methyl polysiloxane) capillary column with dimension 30 m  $\times$  0.25 mm i. d. and 0.25  $\mu$ m film thickness. The carrier gas used is helium which has a linear velocity of 1 ml/min. Injector and detector temperatures were at 200  $^{\circ}$ C and 250  $^{\circ}$ C, respectively, and the volume of the injected sample was 1  $\mu$ L. The MS operating parameters were given as: ionization potential, 70 eV, interface temperature, 250  $^{\circ}$ C, and acquisition mass range, 50-600. Consequently, the identification of components was done by comparison of their mass spectra and retention time with those of the authentic/standard compounds; by computer matching with NIST and WILEY library; and by comparison of the fragmentation pattern of the mass spectral data with those reported in the literature.

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### REFERENCES

1. D. Anderson, *Forest Ecology and Management*, **1993**, 58.
2. B. Elmqvist, L. Olsson, E.M. Elamin, A. Warren, *Agroforestry systems*, **2005**, 64.
3. A. Islam, G. Phillips, A. Slijivo, M. Snowden, P. Williams, *Food Hydrocolloids*, **1997**, 11.

4. G.A. Yasseen, A.A. Salih, M.E. Ahmed, *Procedia-Social and Behavioral Sciences*, **2014**, 120.
5. S. Al Assaf, G.O. Phillips, P.A. Williams, *Food Hydrocolloids*, **2005**, 19.
6. B.H. Ali, S. Beegam, I. Al-Lawati, M. Waly, M. Al Za'abi, A. Nemmar, *Physiological Research*, **2013**, 62.
7. J.W. Bainbridge, A.J. Smith, S.S. Barker, S. Robbie, R. Henderson, K. Balaggan, A. Viswanathan, G.E. Holder, A. Stockman, N. Tyler, *New England Journal of Medicine*, **2008**, 358.
8. A.A.H.M. Elnour, Fractionation; Physicochemical and Functional Properties of Acacia Polyacantha Gum, in, UOFK, 2015.
9. B.H. Ali, M.S. Al Moundhri, *Food and Chemical Toxicology*, **2006**, 44.
10. J.A. Hinson, A.B. Reid, S.S. McCullough, L.P. James, *Drug Metabolism Reviews*, **2004**, 36.
11. H. Trommer, R.H. Neubert, *International Journal of Pharmaceutics*, **2005**, 298.
12. C.J. Evans, D. Keith, H. Morrison, K. Magendzo, R.H. Edwards, SCIENCE-NEW YORK THEN WASHINGTON-, **1992**, 258.
13. A. Tiss, F. Carrière, R. Verger, *Analytical Biochemistry*, **2001**, 294.
14. B.H. Ali, G. Blunden, M.O. Tanira, A. Nemmar, *Food and chemical Toxicology*, **2008**, 46.
15. D.Z. Bliss, T.P. Stein, C.R. Schleifer, R.G. Settle, *The American journal of clinical nutrition*, **1996**, 63.
16. N. Matsumoto, S. Riley, D. Fraser, S. Al-Assaf, E. Ishimura, T. Wolever, G. Phillips, A.O. Phillips, *Kidney international*, **2006**, 69.
17. D.A. Glover, K. Ushida, A.O. Phillips, S.G. Riley, *Food Hydrocolloids*, **2009**, 23.
18. K.U. Rehman, M.A. Wingertzahn, S. Teichberg, R.G. Harper, R.A. Wapnir, *Digestive diseases and sciences*, **2003**, 48.
19. R.A. Wapnir, B. Sherry, C.N. Codipilly, L.O. Goodwin, I. Vancurova, *Digestive diseases and sciences*, **2008**, 53.
20. F. Chemat, M.A. Vian, G. Cravotto, *International journal of molecular sciences*, **2012**, 13.
21. F. Fathordoobady, H. Mirhosseini, J. Selamat, M.Y.A. Manap, *Food Chemistry*, **2016**, 202.
22. H.K. Kala, R. Mehta, K.K. Sen, R. Tandey, V. Mandal, *Trends in Analytical Chemistry*, **2016**, 85, Part C.
23. D.-P. Xu, J. Zheng, Y. Zhou, Y. Li, S. Li, H.-B. Li, *Food Chemistry*, **2017**, 217.
24. M. Mushtaq, B. Sultana, F. Anwar, A. Adnan, S.S.H. Rizvi, *The Journal of Supercritical Fluids*, **2015**, 104.
25. F. Chemat, G. Cravotto, "Enhancing Extraction Processes in the Food Industry", CRC Press, Taylor and Francis Group, Boca Raton, London, New York, 2011, chapter 6.
26. F. Chemat, N. Rombaut, A.-G. Sicaire, A. Meullemiestre, A.-S. Fabiano-Tixier, M. Abert-Vian, *Ultrasonics Sonochemistry*, **2017**, 34.
27. E. Roselló-Soto, C.M. Galanakis, M. Brnčić, V. Orlien, F.J. Trujillo, R. Mawson, K. Knoerzer, B.K. Tiwari, F.J. Barba, *Trends in Food Science & Technology*, **2015**, 42.

28. G. Ruiz-Montañez, J. Ragazzo-Sánchez, M. Calderón-Santoyo, G. Velazquez-De La Cruz, J.R. de León, A. Navarro-Ocaña, *Food chemistry*, **2014**, 159.
29. L. Wang, C.L. Weller, *Trends in Food Science & Technology*, **2006**, 17.
30. R.H. Myers, D.C. Montgomery, C.M. Anderson-Cook, *Response surface methodology: process and product optimization using designed experiments*, John Wiley & Sons, 2016.
31. K. Chaisu, A.L. Charles, Y.-K. Guu, T.-B. Yen, C.-H. Chiu, *APCBEE procedia*, **2014**, 8.
32. M. Chen, Y. Zhao, S. Yu, *Food chemistry*, **2015**, 172.
33. C. Kotzamanidis, T. Roukas, G. Skaracis, *World Journal of Microbiology and Biotechnology*, **2002**, 18.
34. R.S. Makkar, S.S. Cameotra, I.M. Banat, *AMB express*, **2011**, 1.
35. L.V. Candiotti, M.M. De Zan, M.S. Cámara, H.C. Goicoechea, *Talanta*, **2014**, 124.
36. Z. Šumić, A. Vakula, A. Tepić, J. Čakarević, J. Vitas, B. Pavlić, *Food Chemistry*, **2016**, 203.
37. Y. Yuan, Y. Gao, L. Mao, J. Zhao, *Food Chemistry*, **2008**, 107.
38. E. Dorta, M.G. Lobo, M. González, *LWT-Food Science and Technology*, **2012**, 45.
39. F.B. Abdelaal, R.K. Rowe, *Geotextiles and Geomembranes*, **2014**, 42.
40. Y.-Q. Ma, J.-C. Chen, D.-H. Liu, X.-Q. Ye, *Ultrasonics Sonochemistry*, **2009**, 16.
41. M. Esclapez, J. García-Pérez, A. Mulet, J. Cárcel, *Food Engineering Reviews*, **2011**, 3.
42. M.N. Alam, N.J. Bristi, M. Rafiquzzaman, *Saudi Pharmaceutical Journal*, **2013**, 21.
43. J. Kubola, S. Siriamornpun, *Food Chemistry*, **2008**, 110.
44. W. Greenaway, F.R. Whatley, *Phytochemistry*, **1991**, 30.
45. S. Fiorito, V.A. Taddeo, S. Genovese, F. Epifano, *Tetrahedron Letters*, **2016**, 57.
46. K.H. Musa, A. Abdullah, K. Jusoh, V. Subramaniam, *Food Analytical Methods*, **2011**, 4.
47. Q.V. Vuong, C.D. Goldsmith, T.T. Dang, V.T. Nguyen, D.J. Bhuyan, E. Sadeqzadeh, C.J. Scarlett, M.C. Bowyer, *Antioxidants*, **2014**, 3.
48. K.H. Musa, A. Abdullah, B. Kuswandi, M.A. Hidayat, *Food chemistry*, **2013**, 141.
49. V. Stankov-Jovanović, M. Ilić, V. Mitić, T. Mihajilov-Krstev, S. Simonović, S.N. Mandić, J. Tabet, R. Cole, *Journal of pharmaceutical and biomedical analysis*, **2015**, 111.

## OXIDATIVE STRESS MARKERS IN ACUTE MYOCARDIAL INFARCTION TREATED BY PRIMARY PERCUTANEOUS CORONARY INTERVENTION

DAN-ALEXANDRU TĂTARU<sup>a,b</sup>, DAN-MIRCEA OLINIC<sup>a,b</sup>\*,  
ANDRADA URDĂ<sup>a</sup>, MARIA OLINIC<sup>a,b</sup>, REMUS ORĂȘAN<sup>a</sup>

**ABSTRACT.** The current study analysed the dynamics of oxidative stress markers in patients with acute ST-elevation myocardial infarction treated by primary percutaneous intervention. Excessive reactive oxygen species production is known to induce myocardial reperfusion injury. There are few studies that evaluated oxidative stress markers in the interventional era, the historical papers were all based on thrombolysis as a mean of reperfusion. Thirty-seven patients were included. Peripheral venous blood samples were obtained prior to coronary angioplasty, at 1 hour and 24 hours after that. Plasma malondialdehyde, reduced glutathione / oxidised glutathione ratio and total antioxidant capacity were determined. Malondialdehyde was significantly lower at 1 hour ( $3.1 \pm 0.96$  nmol/ml vs  $2.68 \pm 0.81$  nmol/ml,  $p < 0.01$ ) and 24 hours ( $3.1 \pm 0.96$  nmol/ml vs  $2.15 \pm 0.95$  nmol/ml,  $p < 0.01$ ). Also, reduced glutathione / oxidised glutathione ratio dropped significantly at 1 hour (3.25 Q1-Q3 2.17-5.19 vs 2.33 Q1-Q3 1.53-2.82,  $p < 0.01$ ) and at 24 hours (3.25 Q1-Q3 2.17-5.19 vs 1.96 Q1-Q3 1.28-2.85,  $p < 0.01$ ). Total antioxidant capacity had non-significant variation. There was no correlation between these markers and time from symptom-onset or left ventricular ejection fraction. Reperfusion of the occluded coronary artery by percutaneous coronary intervention in acute myocardial infarction led to a rapid decrease of reduced glutathione / oxidised glutathione ratio, that may indicate a depletion of antioxidants as a consequence of overproduction of reactive oxygen species in the damaged area. However, the malondialdehyde level significantly decreased after vessel opening. This may suggest low reperfusion injury after angioplasty.

**Keywords:** *Oxidative stress, acute myocardial infarction, malondialdehyde, glutathione, total antioxidant capacity*

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<sup>a</sup> Iuliu Hațieganu University, Faculty of General Medicine, 8 Victor Babeș street, RO-400012, Cluj-Napoca, Romania

<sup>b</sup> Cluj County Emergency Hospital, Interventional Cardiology Department, 3-5 Clinicilor street, RO-400006, Cluj-Napoca, Romania

\* Corresponding author: [dolinic@yahoo.com](mailto:dolinic@yahoo.com)

## INTRODUCTION

Cardiovascular diseases (CVD) are the leading cause of death globally [1]. Myocardial infarction and stroke account for 80% of CVD deaths in males and 75% in females [2]. The principal objective of therapy for acute ST-elevation myocardial infarction (STEMI) is timely restoration of coronary blood flow, to preserve as much heart muscle as possible and to prevent further complications. This can be done by thrombolysis or percutaneous coronary intervention (PCI). However, a sudden restoration of oxygen supply to a previously ischemic myocardium can lead to myocardial reperfusion injury [3], that can paradoxically reduce the beneficial effect of vessel opening. This culminates with the death of cardiac myocytes that were viable immediately before myocardial reperfusion, thus leading to an increase in infarct size.

The mechanism of myocardial reperfusion injury is not fully understood and it is most probably multifactorial: oxidative stress, lipid peroxidation [3], inflammation, microvascular obstruction [4] and excessive catecholamine release [5].

Malondialdehyde (MDA), a stable lipid peroxidation end-product, is frequently used as a marker of oxidative stress and reactive oxygen species (ROS) production [6].

Reduced glutathione (GSH) is an important antioxidant serving as an electron donor and thereby preventing damage to important cellular components caused by ROS. Oxidised glutathione, also known as glutathione disulphide (GSSG), is a compound that gives information about the usage of GSH during oxidative stress reactions [7]. In a resting cell, the molar GSH:GSSG ratio exceeds 100:1, while in various models of oxidative stress this ratio has been demonstrated to decrease to values of 10:1 and even 1:1 [8].

In contrast to Folin-Ciocalteu method which is used only 'in vitro' [9], the scavenging of the 1,1-diphenyl-2-picrylhydrazyl (DPPH) free radical offers a simple approach for evaluating the 'in vivo' total antioxidant capacity (TAC) of biological sources [10].

The aim of the current study was to determine the dynamics of MDA, GSH/GSSG ratio and TAC (before reperfusion, 1 hour and 24 hours after angioplasty) in patients admitted with STEMI undergoing emergency successful primary PCI of the infarct-related artery and study the correlation with the time from symptom-onset and left ventricular ejection fraction (LVEF). The variation of oxidative stress markers was statistically evaluated using t-test for paired samples or Wilcoxon Signed Ranks Test, depending on the variable distribution and Pearson's coefficient to determine correlations.

According to the authors' knowledge, no comprehensive work was dedicated to oxidative stress markers in acute myocardial infarction treated with primary PCI. The historical papers were all based on thrombolysis as a mean of reperfusion. These studies have showed that, after successful thrombolysis, there is an increase in plasma MDA levels [11-14] and a decrease in antioxidant capacity [11,15]. In the invasive era, PCI is the preferred reperfusion strategy in infarct-related arteries, since it reduces mortality, as compared to thrombolysis, provided it can be performed expeditiously by an experienced team [16].

## RESULTS AND DISCUSSION

Thirty-seven patients were enrolled. The measured variables of the study group are depicted in Table 1.

The mean MDA concentration before PCI ( $P_0$ ) was  $3.1 \pm 0.96$  nmol/ml. After reperfusion, a significant decrease of MDA levels occurred at 1 hour ( $P_1$ ,  $2.68 \pm 0.81$  nmol/ml,  $p < 0.01$ ), and 24 hours ( $P_{24}$ ,  $2.15 \pm 0.95$  nmol/ml,  $p < 0.01$ ). Also, levels at 24 hours were significantly lower than MDA concentrations noted at 1 hour,  $p < 0.01$  (Figure 1).

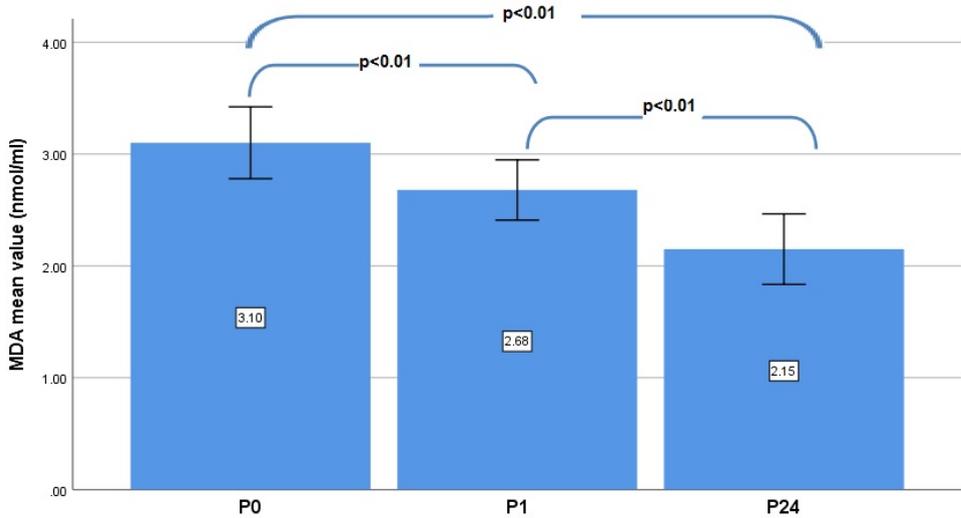
The median GSH/GSSG ratio before reperfusion ( $P_0$ ) was 3.25 (2.17-5.19). A significant decrease was observed after 1 hour following PCI ( $P_1$ , 2.33 (1.53-2.82),  $p < 0.01$ ) and at 24 hours ( $P_{24}$ , 1.96 (1.28-2.85),  $p < 0.01$ ). Also, the difference between  $P_1$  and  $P_{24}$  was significant, 2.33 (1.53-2.82) vs 1.96 (1.28-2.85),  $p = 0.036$  (Figure 2).

TAC did not have significant variations ( $p = \text{non-significant}$ ): before reperfusion ( $P_0$ ,  $32.7 \pm 4.86$  inhibition %); at 1 hour after vessel opening ( $P_1$ ,  $32.08 \pm 6.42$  inhibition %); at 24 hours ( $P_{24}$ ,  $33.04 \pm 8.34$  inhibition %) (Figure 3).

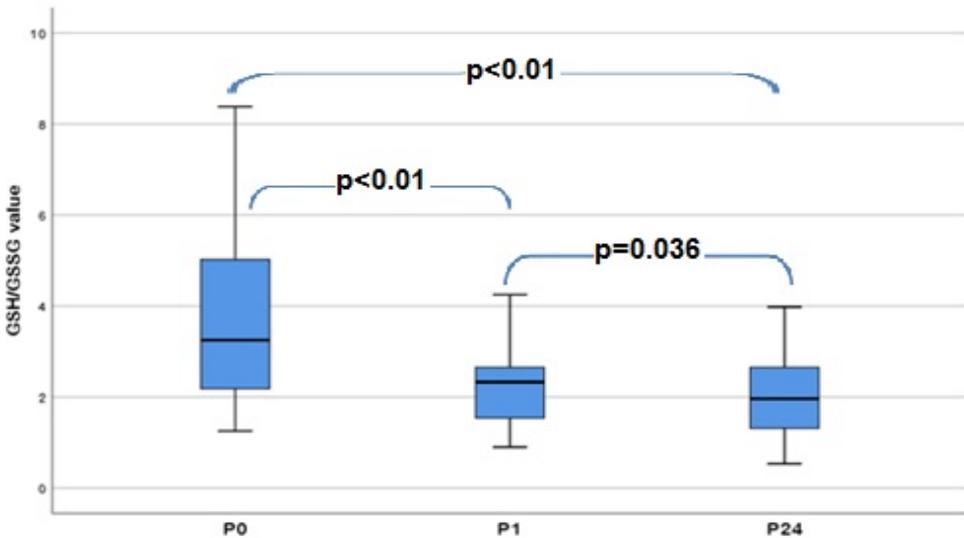
There was no correlation between MDA, GSH/GSSG ratio, TAC levels and the duration of coronary occlusion, as measured by the time from symptom-onset to the opening of the infarct-related artery. Also, no correlation was found between these markers and LVEF at admission.

**Table 1.** Variables of the study group. (TSR = time from symptom-onset to reperfusion (hours), LVEF = left ventricular ejection fraction (%), MDA = malondialdehyde (nmol/ml), G/G = reduced glutathione / oxidised glutathione ratio, TAC = total antioxidant capacity (inhibition %))

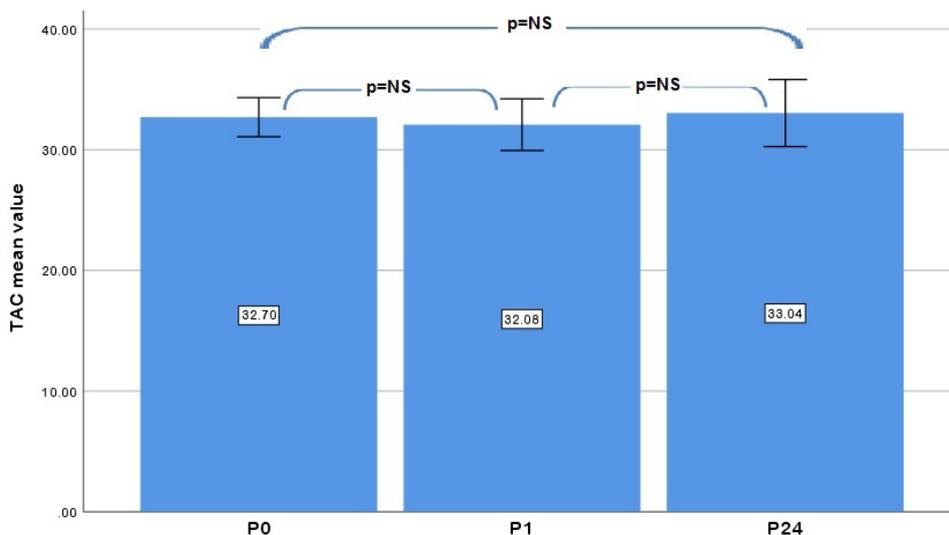
			P0			P1			P24		
	TSR	LVEF	MDA	G/G	TAC	MDA	G/G	TAC	MDA	G/G	TAC
Pt 1	8	35	1.94	7.96	37.61	2.09	2.33	26.03	1.66	1.56	26.66
Pt 2	6	50	1.59	8.38	37.63	1.64	4.88	40.25	2.22	4.95	39.71
Pt 3	3	45	2.82	5.44	24.33	3.10	1.73	24.20	2.49	2.38	24.07
Pt 4	12	40	4.71	3.10	27.06	3.87	1.21	26.32	4.23	1.43	28.41
Pt 5	6	45	3.02	2.69	41.86	3.67	1.04	49.15	3.94	1.15	58.22
Pt 6	2	50	4.02	5.02	29.09	3.01	2.98	32.90	3.77	0.79	35.61
Pt 7	3	50	3.48	2.31	39.13	3.20	1.88	37.49	1.54	1.79	41.74
Pt 8	6	30	2.19	3.81	26.19	1.17	4.06	27.85	0.75	3.05	28.98
Pt 9	2	50	3.45	3.58	28.84	4.35	2.53	18.48	2.83	2.65	12.80
Pt 10	12	50	3.30	6.35	31.26	2.17	2.34	31.68	0.53	1.25	41.93
Pt 11	2	30	2.03	8.35	38.48	1.65	4.25	41.28	0.76	3.98	42.25
Pt 12	5	50	2.46	1.35	36.07	1.88	1.54	37.61	1.69	1.12	33.41
Pt 13	3	35	3.16	1.25	28.38	2.28	1.35	28.02	1.32	1.05	25.72
Pt 14	4	50	2.86	2.32	24.15	2.46	2.62	22.09	2.22	1.96	21.81
Pt 15	5	45	3.56	5.63	25.44	3.35	2.35	25.24	1.66	3.25	20.39
Pt 16	10	40	1.96	1.65	32.38	1.72	1.36	31.45	1.06	1.98	36.40
Pt 17	12	45	4.98	3.25	25.96	3.86	2.04	25.50	3.15	1.96	27.68
Pt 18	4	45	4.71	1.35	33.35	3.57	1.05	29.01	2.43	0.53	32.25
Pt 19	4	50	4.06	1.98	29.57	2.57	1.99	34.13	2.03	2.02	31.53
Pt 20	4	50	1.96	2.63	32.88	1.92	2.02	31.48	1.21	1.25	32.64
Pt 21	5	30	2.86	2.15	35.65	2.71	1.88	37.70	1.62	1.00	35.70
Pt 22	4	50	4.13	9.60	37.22	3.02	2.44	36.00	2.66	2.62	33.10
Pt 23	6	50	2.80	1.48	34.30	2.08	3.99	33.45	1.93	3.26	30.52
Pt 24	6	50	3.30	1.46	39.88	3.27	1.15	30.18	3.03	1.31	29.32
Pt 25	5	45	3.96	3.57	32.21	3.66	3.02	36.13	2.43	3.12	41.38
Pt 26	2	50	4.53	2.25	29.64	3.80	2.39	31.05	3.34	1.36	31.60
Pt 27	3	50	2.91	4.41	36.40	2.61	0.95	33.44	2.08	1.59	32.21
Pt 28	4	50	1.28	3.25	32.64	1.71	2.50	31.08	1.09	2.03	37.64
Pt 29	6	45	1.48	1.65	37.06	1.29	2.02	37.38	1.52	1.02	43.41
Pt 30	12	50	3.44	2.97	27.90	2.44	8.10	20.05	2.81	3.34	22.40
Pt 31	3	50	2.01	6.56	32.38	1.84	5.34	33.95	1.66	2.56	37.60
Pt 32	12	50	2.69	5.35	39.81	2.40	4.35	37.85	1.37	3.25	37.42
Pt 33	12	40	4.16	4.25	36.45	3.19	2.53	31.54	2.33	1.53	41.67
Pt 34	6	50	4.00	3.36	32.46	3.43	2.65	37.00	3.97	1.98	36.56
Pt 35	12	40	3.28	2.18	32.97	3.06	1.98	31.57	2.70	5.66	31.10
Pt 36	12	50	2.69	4.55	27.03	2.49	0.90	26.82	2.20	1.82	21.26
Pt 37	12	50	2.95	2.94	36.14	2.58	1.52	41.62	1.30	2.43	37.45



**Figure 1.** Comparison of MDA mean values: P<sub>0</sub> - before reperfusion; P<sub>1</sub> - 1 hour after PCI; P<sub>24</sub> - 24 hours after PCI (t-test for paired samples, error bars: 95% confidence interval)



**Figure 2.** Distribution of GSH/GSSG values; P<sub>0</sub> - before reperfusion; P<sub>1</sub> - 1 hour after PCI; P<sub>24</sub> - 24 hours after PCI (Wilcoxon Signed Ranks Test; bars = range, box = first quartile to third quartile, horizontal black line = median)



**Figure 3.** Comparison of TAC mean values; P<sub>0</sub> - before reperfusion; P<sub>1</sub> - 1 hour after PCI; P<sub>24</sub> - 24 hours after PCI (t-test for paired samples, error bars: 95% confidence interval; NS = non-significant)

Several studies have evaluated MDA levels in patients with STEMI treated with thrombolytic therapy [10-14]. In all studies, there was an increase of MDA concentrations after thrombolysis, although in some studies, the increase did not reach the level of statistical significance. Two studies [10,12] reported that, after an initial increase, there is a decrease in MDA levels, 2 to 6 hours after successful thrombolysis.

Although a rise in MDA levels was expected, MDA concentrations actually decreased significantly after PCI. Similar findings were reported in other PCI-based studies [6,17]. This can be explained by the faster and complete coronary vessel opening after PCI, as compared to thrombolysis. The decrease in MDA supports low lipid peroxidation in myocardial infarction patients treated with primary PCI.

Lipid peroxidation was evaluated by the thiobarbituric acid (TBA) fluorescence method. This approach has potential limitations with respect to specificity. Another way of determining MDA is high-performance liquid chromatography, which is more specific [18]. Considering that the TBA method can detect even non-lipid related, TBA-reactive substances [18], the results suggest that oxidant-mediated injury may actually be modest in PCI treated patients. The finding of consistently and progressively decreasing MDA levels after reperfusion supports this conclusion.

GSH/GSSG ratio had a significant decrease after PCI, showing increased oxidative stress and a possible sign of myocardial reperfusion injury, which is in contrast with reduced lipid peroxidation. Similarly, another study reported that human hearts subjected to cardioplegia and subsequent reperfusion show oxidative stress, without an increase in lipid peroxidation [19].

TAC levels had non-significant variations before and after vessel opening. Other authors reported a significant drop in the first hours after PCI reperfusion, but the levels returned to baseline values on the third day [6]. In the mentioned study [6], TAC levels were assessed by a different method, using the 2,2'-azino-bis(3-ethylbenzthiazoline-6-sulphonic acid) reaction [20].

Although an experimental study on rat hearts reported increased MDA levels during prolongation of ischemia [21], there was no correlation in the present study between oxidative stress markers and the time from symptom-onset to reperfusion, which is similar to other PCI studies [6].

In animal models with acute myocardial infarction, a significant positive correlation between GSH/GSSG ratio and LVEF was observed [22]. In contrast, there was no correlation between LVEF and oxidative stress markers in this study group.

The present study showed that in STEMI patients, PCI reduces some of the oxidative stress burden, such as lipid peroxidation, while enhancing ROS production by other pathways. This may suggest different mechanisms leading to oxidative stress. Further studies are needed to properly differentiate the pathophysiology of oxidative stress production after reperfusion.

## CONCLUSION

In STEMI patients, the decrease in GSH/GSSG ratio after PCI suggests increased oxidative stress and is a possible marker of myocardial reperfusion injury.

However, the decrease in MDA levels immediately after vessel opening suggests low oxidative damage after PCI.

There were no significant variations in TAC before and after PCI.

No correlations between MDA, GSH/GSSG ratio, TAC and time from symptom-onset to reperfusion or LVEF were observed.

## EXPERIMENTAL SECTION

The study complied with the Declaration of Helsinki [23]. Written consent was obtained from each patient before the procedure. Patients diagnosed with STEMI and admitted to the Cluj County Emergency Hospital,

Interventional Cardiology Department were enrolled between April 2017 and December 2017. Inclusion criteria were as follows: electrocardiographic evidence of ST elevation of  $\geq 1$  mV in two or more standard limb leads or  $\geq 2$  mV in two or more precordial leads; typical chest pain lasting more than 20 min; time of presentation under 12 hours since symptom onset. PCI was confined to the infarct-related artery and all patients received drug eluting stents. The procedure was considered successful, if the thrombolysis in myocardial infarction (TIMI) flow was 3 and a residual stenosis was under 30%. Only patients with successful reperfusion were included in the study. Patients with Killip class  $> 3$  (cardiogenic shock) were excluded because of the very severe condition and need of intensive care, which could influence the results. All patients were pretreated with double anti-platelet therapy before PCI and received 100 UI/kg of unfractionated heparin during the procedure. Baseline echocardiography before the procedure was performed to assess LVEF and possible complications. All images were stored in the Department's Digital Imaging and Communications in Medicine database [24], for future analysis. The baseline demographic, clinical and non-clinical characteristics are presented in Table 2.

**Table 2.** Baseline characteristics of the study group. (SD = standard deviation, TSR = time from symptom-onset to reperfusion, BMI = body-mass index, PCI = percutaneous coronary intervention, LAD = left anterior descending artery, CX = circumflex artery, RCA = right coronary artery, LVEF = left ventricular ejection fraction)

<b>Presentation</b>		<b>Medical history</b>	
Age (years), mean $\pm$ SD	63 $\pm$ 11	Previous PCI (yes), no (%)	2 (5.4)
Sex (male), no (%)	24 (64.9)	Prior infarction (yes), no (%)	5 (13.5)
TSR (hours), mean $\pm$ SD	6.35 $\pm$ 3.63	<b>Infarct-related artery</b>	
Killip I (yes), no (%)	30 (81.1)	LAD (yes), no (%)	18 (48.6)
Killip II (yes), no (%)	5 (13.5)	CX (yes), no (%)	10 (27)
Killip III (yes), no (%)	2 (5.4)	RCA (yes), no (%)	9 (24.4)
<b>Risk factors</b>		<b>Echocardiography</b>	
Hypertension (yes), no (%)	26 (70.3)	LVEF (%), mean $\pm$ SD	45.54 $\pm$ 6.43
Dyslipidemia (yes), no (%)	23 (62.1)		
Smokers (yes), no (%)	14 (37.8)		
Diabetes (yes), no (%)	7 (18.9)		
BMI (kg/m <sup>2</sup> ), mean $\pm$ SD	29.1 $\pm$ 4.3		

Peripheral venous blood samples were obtained from each patient immediately before coronarography ( $P_0$ ) and then after 1 hour ( $P_1$ ) and 24 hours ( $P_{24}$ ) after successful PCI. Samples were drawn into plastic tubes with

EDTA. Blood was centrifuged within 1 hour at 1500 rpm for 15 minutes, and the collected plasma was stored at  $-30^{\circ}\text{C}$  until analysis, within 28 days. All the reagents were supplied by Sigma (Deisenhofen, Germany).

MDA was determined by the TBA reaction [25]. First, 50  $\mu\text{l}$  of plasma was boiled with 1 ml of 10 mM 2-TBA and 1 ml of 75 mM  $\text{K}_2\text{HPO}_4$ , pH=3. After sudden cooling, the product was extracted with n-butanol. Using fluorescence spectrometry (Lambda 35, Perkin Elmer, USA), the sample was measured at an emission wavelength of 534 nm with a synchronous fluorescence technique at a difference of 14 nm between the excitation and emission wavelength. The MDA levels were calculated according to a calibration curve realised with known concentrations of MDA.

GSH was determined by fluorescence spectrometry [26]. A 500  $\mu\text{l}$  aliquot of plasma is added to 500  $\mu\text{l}$  of cold 10% trichloroacetic acid. After 10 minutes in ice, the mixture is centrifuged (3000 rpm for 15 minutes) and 200  $\mu\text{l}$  of the supernatant is mixed with 1.7 ml of phosphate buffer and 0.1 ml of o-phthalaldehyde. After 15 minutes, the fluorescence at 350 nm excitation and 420 nm emission is read against a blank (Lambda 35, Perkin Elmer, USA).

For GSSG estimation [27], 250  $\mu\text{l}$  of the plasma sample was incubated with 0,1 ml 40 nM N-ethylmaleimide for 30 minutes followed by addition of 0,65 ml of 0.1 M NaOH. Thereafter, the same procedure was followed for fluorescence development as used in GSH measurement, except in place of the buffer, 0.1 M NaOH was used.

The concentrations of GSH and GSSG were calculated from calibration curves and are expressed in  $\mu\text{mol/ml}$ .

TAC was determined by measurement of DPPH method [28]. The reduction assay was performed by adding 20  $\mu\text{l}$  of plasma to 400  $\mu\text{l}$  of 0.1 mM methanol solution of DPPH and phosphate buffer, pH=7,4. After 30-minute incubation at ambient temperature, absorbance of the samples at 520 nm was measured (Lambda 35, Perkin Elmer, USA) and compared with that of a reference sample containing only DPPH solution and phosphate buffer. TAC was measured in inhibition % as  $[(\text{reference extinction} - \text{serum extinction}) / \text{reference extinction}] \times 100$ .

The statistical analysis was performed using SPSS software v25 for Windows (IBM, USA). The Shapiro-Wilk test was used to assess for a normal distribution. The t-test for paired samples was used for normally distributed data and Wilcoxon Signed Ranks Test for paired samples for not normally distributed. The significance threshold was set at  $p=0.05$ . The data are presented as mean  $\pm$  standard deviation for normally distributed samples; otherwise as median and interquartile range (Q1–Q3), where Q1 = first quartile and Q3 = third quartile. Correlation analysis was performed using Pearson's correlation coefficient.

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## REFERENCES

1. D. Olinic, M. Spinu, M. Olinic, C. Homorodean, D. Tataru, A. Liew, G. Schernthaner, A. Stanek, G. Fowkes, M. Catalano, *International Angiology*, **2018**, 37, 327.
2. A.S. Go, D. Mozaffarian, V.L. Roger, *Circulation*, **2013**, 127, e6.
3. D. Yellon, D. Hausenloy, *New England Journal of Medicine*, **2007**, 357, 1121.
4. M. Marc, A.C. Iancu, C. Ober, C. Homorodean, S. Balanescu, A.V. Sitar, S. Bolboaca, I. M. Dregoes, *Scientific Reports*, **2018**, 8, 1897.
5. C. Homorodean, M. Ober, M. Olinic, R. Homorodean, A. Hassoune, D. Tătaru, M. Spînu, D. Olinic, *Medical Ultrasonography*, **2016**, 18, 475.
6. E. Sedláková, O. Rácz, E. Lovásová, R. Beňačka, M. Kurpas, A. Chmelárová, J. Sedlák, M. Studenčan, *Central European Journal of Medicine*, **2009**, 4, 26.
7. D.M. Mitrea, S. Clichici, A. Filip, D. Olteanu, I. Baldea, R. Moldovan, N. Decea, O.A. Hoteiuc, *Studia UBB Chemia*, **2017**, 62, 89.
8. Y. Chai, S. Ashraf, K. Rokutan, R. Johnston, J. Thomas, *Archives of Biochemistry and Biophysics*, **1994**, 310, 273.
9. I. Simon, D. Simedru, L. Dordai, E. Luca, V. Fuss, A. Becze, *Studia UBB Chemia*, **2016**, 61, 505.
10. S.B. Kedare, R.P. Singh, *Journal of Food Science and Technology*, **2011**, 48, 412.
11. T. Beard, D. Carrie, M. Boyer, B. Boudjemaa, J. Ferrières, M. Delay, *Archives des maladies du coeur et des vaisseaux*, **1994**, 87, 1289.
12. K. Iqbal, M.A. Rauoof, M. Mir, N. Trambo, J. Malik, B. Naikoo, *American Journal of Cardiology*, **2002**, 89, 334.
13. I. Young, J. Purvis, J. Lightbody, A. Adgey, *European Heart Journal*, **1993**, 14, 1027.
14. S. Pucheu, C.H. Coudray, G. Vanzetto, A. Favier, J. Machecourt, J. de Leiris, *Free Radical Biology and Medicine*, **1995**, 19, 873.
15. K. Berg, P. Jynge, K. Bjerve, S. Skarra, S. Basu, R. Wiseth, *Free Radical Research*, **2005**, 6, 629.
16. B. Ibanez and the ESC Taskforce, *European Heart Journal*, **2017**, 00, 1.
17. K. Olsson, J. Harnek, A. Ohlin A, N. Pavlidis, B. Thorvinger, H. Ohlin, *Scandinavian Cardiovascular Journal*, **2002**, 36, 237.

18. C. Ceconi, A. Cargoni, E. Pasini, et al, *American Journal of Physiology-Heart and Circulatory Physiology*, **1991**, 260, H1057.
19. M. Janssen, J.F. Koster, E. Bos, J. De Jong, *Circulation Research*, **1993**, 73, 681.
20. C. Rice Evans, N. Miller, *Methods in Enzymology*, **1994**, 234, 279.
21. B. Tavazzi, D. Di Pierro, M. Bartolini, M. Marino, S. Distefano, M. Galvano, *Free Radical Research*, **1998**, 1, 25.
22. A. Tavares, A. da Rosa Araujo, S. Llesuy, et al, *Experimental and Clinical Cardiology*, **2012**, 17, 263.
23. World Medical Association, *Journal of the American Medical Association*, **2013**, 310, 2191.
24. C. Homorodean, M. Olinic, D.M. Olinic, *Medical Ultrasonography*, **2012**, 14, 29.
25. M. Conti, P.C. Morand, P. Levillain, A. Lemonnier, *Clinical Chemistry*, **1991**, 37, 1273.
26. M.L. Hu, *Methods in Enzymology*, **1994**, 233, 380.
27. P. Vats, V.K. Singh, S.S. Sing, *Aviation, Space and Environmental Medicine*, **2008**, 79, 1106.
28. A. Janaszewska, G. Bartosz, *Scandinavian Journal of Clinical and Laboratory Investigation*, **2002**, 62, 231.



## SYNTHESIS AND CHARACTERIZATION OF MUC-1 FUNCTIONALIZED GOLD NANOPARTICLES

CRISTIAN T. MATEA<sup>a</sup>, TEODORA MOCAN<sup>a,b\*</sup>, FLAVIU TABARAN<sup>a,c</sup>,  
TEODORA POP<sup>d</sup>, OFELIA MOSTEANU<sup>d</sup>, LUCIAN MOCAN<sup>a,e</sup>,  
CLAUDIU ZDREHUS<sup>e</sup>

**ABSTRACT.** Gold nanoparticles have been functionalized with the biologically active MUC-1 peptide conjugated to KLH and the obtained bio-nanostructure was characterized using an array of techniques such as: UV-Vis and ATR-FT-IR spectroscopy, dynamic light scattering (DLS) and atomic force microscopy (AFM). A 54 nm monodisperse bio-nanostructure with high aqueous stability was obtained.

**Keywords:** *gold nanoparticles, functionalization, MUC-1 peptide*

### INTRODUCTION

Metal nanoparticles and especially gold nanoparticles (GNPs) have attracted a lot of interest from the scientific community due to their unique properties such as: large surface-to-volume ration, optoelectronic properties, rich functionality, low toxicity and high biocompatibility [1, 2]. Recent advances in the fields of nanoparticles synthesis and functionalization resulted in novel medical applications [3, 4]. One area of biomedical research in which GNPs have center stage is the development of carriers for cellular delivery of drugs, proteins and non-cell-penetrating peptides [5-8].

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<sup>a</sup> Department of Nanomedicine "Octavian Fodor" Gastroenterology Institute, Cluj-Napoca, Romania

<sup>b</sup> Department of Physiology, "Iuliu Hatieganu" University of Medicine and Pharmacy, Cluj-Napoca, Romania

<sup>c</sup> Department of Pathology, Faculty of Veterinary Medicine, University of Agricultural Sciences and Veterinary Medicine

<sup>d</sup> 3rd Gastroenterology Department "Iuliu Hatieganu" University of Medicine and Pharmacy, Cluj-Napoca, Romania

<sup>e</sup> 3rd Surgery Clinic "Iuliu Hatieganu" University of Medicine and Pharmacy Cluj-Napoca Romania

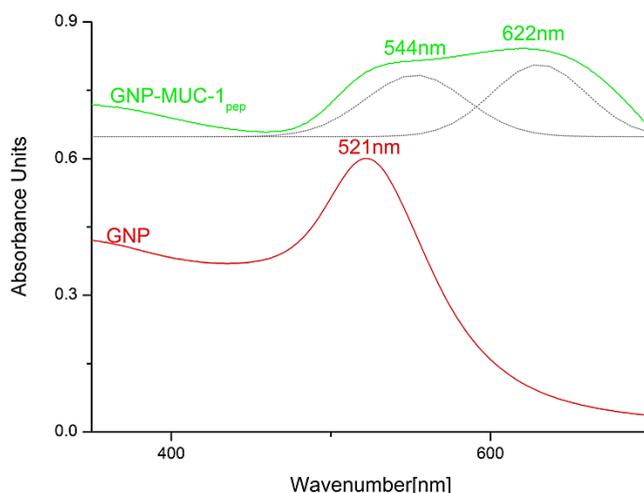
\* Corresponding author: teodora.mocan@umfcluj.ro

Keyhole limpet hemocyanin (KLH) is a high molecular weight copper-containing protein which can act as a nonspecific immune stimulant [9]. A study conducted by Kim et al. showed that there is a strong correlation between the antibodies induced against peptide antigen MUC1 and carbohydrate antigen GD3 with different immunological adjuvants and the strength of Interferon- $\gamma$  release against KLH [10].

We have previously showed that a MUC-1 protein fragment attached on the surface of gold nanoparticles can elicit the activation of peritoneal macrophages as a possible anticancer vaccine [11]. In our present study we have functionalized gold nanoparticles with the biologically active MUC-1 peptide which is in turn conjugated to KLH and characterized the obtained bio-nanostructure using an array of techniques such as: UV-Vis and ATR-FT-IR spectroscopy, dynamic light scattering (DLS) and atomic force microscopy (AFM).

## RESULTS AND DISCUSSION

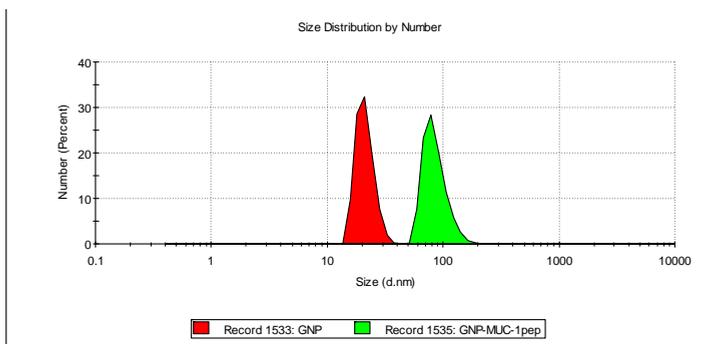
When developing a novel bio-nanostructure a key aspect is its thorough characterization. An incomplete characterization of nanomaterials limits scientific understanding and slows the development and transfer of new technologies [12]. Citrate capped GNPs and MUC-1<sub>pep</sub> functionalized GNPs were characterized by means of UV-Vis, ATR-FT-IR, DLS and AFM techniques.



**Figure 1.** UV-Vis spectra of GNPs (red line) and GNP-MUC-1<sub>pep</sub> (green line); the grey dotted lines represent the deconvoluted sub bands assigned to the Au<sup>0</sup> surface plasmon resonance band(544nm) and KLH protein (622nm).

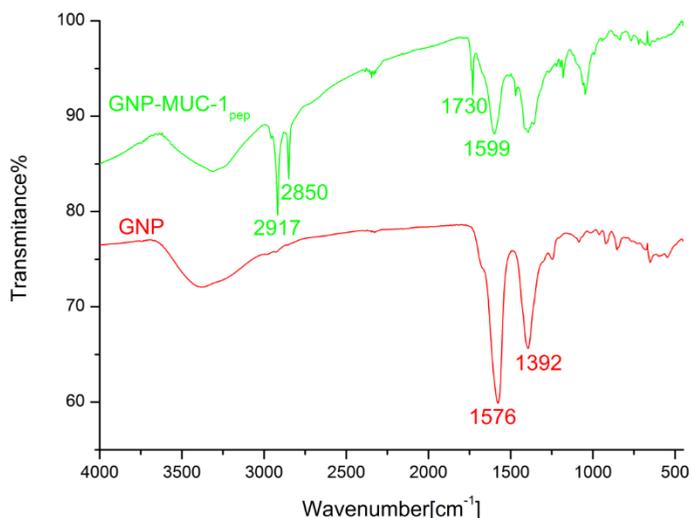
Figure 1 presents the UV-Vis spectra for the citrate capped gold nanoparticles (GNPs –red line) and for the MUC-1<sub>pep</sub> functionalized gold nanoparticles (GNP-MUC-1<sub>pep</sub> - green line). In the case of GNPs the absorption peak registered at 521nm is attributed to the surface plasmon resonance (SPR) band of gold nanoparticles [13, 14]. The GNP-MUC-1<sub>pep</sub> sample registered a broad peak in the region 500-650nm, after spectral deconvolution the resulting two sub bands (grey dotted lines) at 544nm and 622nm were assigned to the gold nanoparticle SPR and the oxidized form of the KLH protein, respectively.

The size of both GNP and GNP-MUC-1<sub>pep</sub> were measured by dynamic light scattering. DLS size distribution curves are presented in figure 2, both samples were monodispersed and presented good stability at room temperature and at 37°C. The increase in size suggested by the bathochromic shift of the SPR peak of colloidal gold after functionalization is also confirmed by the DLS measurements. The citrate capped gold nanoparticles registered a mean hydrodynamic size of 17nm, while de MUC-1<sub>pep</sub> functionalized GNP recorded a mean diameter of 62nm and a polydispersity index of 0.288.



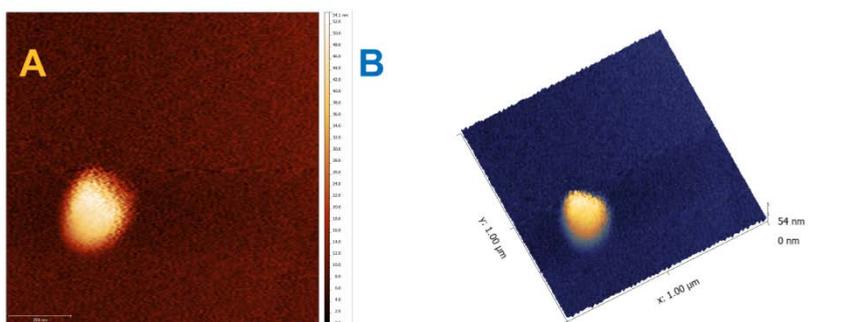
**Figure 2.** DLS size distribution curves for GNP (red) and GNP-MUC-1<sub>pep</sub> (green) samples.

The results from the ATR-FT-IR spectroscopy measurements conducted on GNP and GNP-MUC-1<sub>pep</sub> samples are presented in figure 3. The citrate capped GNPs presented IR adsorption bands centered at 1392 and 1576  $\text{cm}^{-1}$ , which are attributed to the symmetric and antisymmetric stretching of  $\text{COO}^-$  of citrate ions [15-17]. GNP-MUC-1<sub>pep</sub> presented IR adsorption bands at 1730 and 1599  $\text{cm}^{-1}$  which were attributed to the amide I and amide II vibrations from the KLH-MUC-1<sub>pep</sub> conjugate layer present on the surface of the GNPs [15]. Thus, the successful functionalization of the citrate gold nanoparticles with the MUC-1 peptide is confirmed by comparing the IR spectra of the GNP and GNP-MUC-1<sub>pep</sub> samples.



**Figure 3.** ATR-FT-IR spectra for GNP (red) and GNP-MUC-1<sub>1pep</sub> (green)samples

In order to further investigate the size and the shape of the MUC-1<sub>1pep</sub> functionalized gold nanoparticles AFM measurements were conducted. Figure 4.A. depicts a 2D representation, while figure 4.B. shows a 3D representation of a single GNP-MUC-1<sub>1pep</sub> nanoparticle, in both cases the spherical shape of the bio-nanostructure can be observed. GNP-MUC-1<sub>1pep</sub> presented an average diameter of ~54nm, as measured by AFM, while DLS measurements on the same sample provided a mean diameter of ~62nm. This can be explained by the fact that the DLS technique provides a mean hydrodynamic diameter of GNPs surrounded by the KLH protein-MUC-1<sub>1pep</sub> and the solvation layers [15].



**Figure 5.** AFM image of GNP-MUC-1<sub>1pep</sub> : (A) 2D image; (B) 3D image.

## CONCLUSIONS

Gold nanoparticles were functionalized with a KLH-MUC-1 peptide conjugate yielding a 54nm monodisperse bio-nanostructure with high aqueous stability. The characterization of the obtained material involved UV-Vis and FT-IR spectroscopies in order to confirm the success of the functionalization step. The dynamic light scattering and atomic force microscopy techniques were employed to determine the size and shape of the GNPs prior and after functionalization. Investigations into the biological interactions of this novel bio-nanostructure are required in order to determine its potential for medical applications.

## EXPERIMENTAL SECTION

HAuCl<sub>4</sub> (≥99.9%), tri-sodium citrate (≥99%), DL-dithiotreitol (DTT) (≥99%) and NaH<sub>2</sub>PO<sub>4</sub> were purchased from Sigma-Aldrich™ (Darmstadt, Germany). MUC-1 peptide, aminoacid one letter code – TSAPDNRPALGSTA, attached to keyhole limpet hemocyanin (KLH) protein was purchased from Abcam (Austin, USA). All reagents were used as received, without further purification.

Citrate capped gold nanoparticles (GNPs) were obtained by employing a modified Turkevich method. Briefly, 9.8mg HAuCl<sub>4</sub> were dissolved in 20 mL distilled water and heated to 100°C, to this, 2mL sodium citrate (20mg/mL) were added. The reaction was allowed to continue for 2 hours under reflux and continuous stirring, during this stage the solution turned from pale-yellow to dark-red. For the functionalization of the GNPs with KLH-MUC-1 conjugate, a 100µL KLH-MUC-1pep solution (1mg/mL) was mixed with 300µL DTT 100mM (pH=8.5) and was added to 10mL GNP solution and kept under vigorous agitation for 5 minutes. The obtained GNP-MUC-1pep was subjected to a centrifugation step at 15000RPM/30min. and the resulting pellet was re-dispersed in H<sub>2</sub>O bidist. The purified GNP-MUC-1<sub>pep</sub> sample presented good stability over several weeks, at room temperature and at 4°C.

UV-Vis spectroscopy was performed on the samples with a Shimadzu UV-1800 spectrophotometer. The GNP and GNP-MUC-1<sub>pep</sub> spectras were recorded from 800nm to 200nm with a spectral resolution of 0.5nm. OriginLab® software 7.0 was used to normalize all recorded spectras.

Dynamic light scattering (DLS) measurements were performed with a Nano S90 instrument (Malvern Instruments, Westborough, UK) at 25°C at 90° scattering angles.

Universal attenuated total reflectance Fourier transform infrared spectroscopy (ATR-FT-IR) measurements were conducted on a Perkin-Elmer Spectrum Two<sup>®</sup> instrument equipped with an UATR single reflection diamond. Spectral data processing and baseline corrections were done using the Spectrum10<sup>™</sup> software.

Atomic force microscopy (AFM) data were recorded with the aid of a Workshop TT-AFM<sup>®</sup> (AFMWorkshop, CA, USA) operated in vibrating mode using super-sharp ACTA-SS cantilevers (AppNano, CA, USA). The recorded data was further processed with the Gwyddion<sup>®</sup> 2.36 software.

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## REFERENCES

1. N. Elahi; M. Kamali; M.H. Bachersad, *Talanta* **2018**, *184*, 537.
2. A. Gharatape; R. Salehi, *European Journal of Medicinal Chemistry* **2017**, *138*, 221.
3. L. Mocan; F.A. Tabaran; T. Mocan; T. Pop; O. Mosteanu; L. Agoston-Coldea; C. T. Matea; D. Gonciar; C. Zdrehus; C. Iancu, *International Journal of Nanomedicine* **2017**, *12*, 2255.
4. T. Mocan; C.T. Matea; T. Pop; O. Mosteanu; A.D. Buzoianu; S. Suci; C. Puia; C. Zdrehus; C. Iancu; L. Mocan, *Cellular and Molecular Life Sciences* **2017**, *74* (19), 3467.
5. S. Bathia, Nanoparticles Types, Classification, Characterization, Fabrication Methods and Drug Delivery Applications. In *Natural Polymer Drug Delivery Systems*, Springer, Cham: 2016.
6. B. Duncan; C. Kim; V.M. Rotello, *Journal of Controlled Release* **2010**, *148* (1), 122.
7. C.T. Matea; T. Mocan; F. Tabaran; C. Iancu; L. Mocan, *Journal of Nanobiotechnology* **2015**, *13* (41).
8. Z.P. Xu; Q.H. Zeng; G.Q. Lu; A.B. Yu, *Chemical Engineering Science* **2006**, *61* (3), 1027.
9. D.W. McFadden; D.R. Riggs; B.J. Jackson; A. Ng; C. Cunningham, *The American Journal of Surgery* **2007**, *193* (2), 284.

10. S.K. Kim; G. Ragupathi; C. Musselli; S.-J. Choi; Y.S. Park; P.O. Livingston, *Vaccine* **2000**, *18*, 597.
11. T. Mocan; C. Matea; F. Tabaran; C. Iancu; R. Orasan; L. Mocan, *Journal of Cancer* **2015**, *6* (6), 583.
12. D.R. Baer; M.H. Engellhard; G E. Johnson; J. Laskin; J. Lai; K. Mueller; P. Munusamy; S. Thevuthasan; H. Wang; N. Washton, *Journal of Vacuum Science and Technology A* **2013**, *31* (5).
13. L. Shang; Y. Wang; J. Jiang; S. Dong, *Langmuir* **2007**, *23*, 2714-2721.
14. Y. Wang; Y. Ni, *Talanta* **2014**, *119*, 320.
15. G. Mandal; M. Bardhan; T. Ganguly, *Colloids and Surfaces B: Biointerfaces* **2010**, *81* (178-184).
16. J. Park; J. Shumaker-Parry, *Journal of the American Chemical Society* **2014**, *136*, 1907-1921.
17. X. Zou; E. Ying; S. Dong, *Nanotechnology* **2006**, *17* (4758-4764).



## PREPARATION AND CHARACTERIZATION OF HYDROXYAPATITE BASED NANO-COMPOSITE BIOMORPHIC IMPLANTS

RÉKA BARABÁS<sup>a</sup>, MELINDA RIGÓ<sup>a</sup>, MARGIT ENISZNÉ-BÓDOGH<sup>b</sup>,  
CORINA MOISA<sup>c</sup>, OANA CADAR<sup>d\*</sup>

**ABSTRACT.** The aim of this study was the preparation and characterization of different biomorphic implants based on calcined cattle bones coated with hydroxyapatite (Hap) based nanocomposites in different ratios. For comparison, molded nanocomposites were also produced as biomorphic implants. The obtained nanocomposite/implants were characterized using X-ray diffraction, light microscopy and scanning electron microscopy, Brunauer-Emmett-Teller surface area and apparent porosity. The release of Ca and P in simulated body fluid was monitored by X-ray fluorescence. The adsorption capacity and extended-release dosage of implants were investigated with ibuprofen, an anti-inflammatory drug, by UV-VIS spectroscopy. The highest adsorption efficiency and stability were obtained for sintered (S) (Hap-) and 20% gelatin (G) nanocomposite (Hap-20G-S) and bone parts coated with Hap-S and Hap-20G-S nanocomposite, respectively. The best results (high adsorption efficiency and slow release - low desorption capacity) were obtained for molded Hap-20G-S composite, without bone. In summary, the cattle bones with hydroxyapatite coatings show great promise in production of inexpensive and patient-specific bone implants.

**Keywords:** *biomorphic implants, hydroxyapatite, gelatin, carbon nanotubes, ibuprofen*

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<sup>a</sup> Babeş-Bolyai University, Faculty of Chemistry and Chemical Engineering, Hungarian Line of Study, 11 Arany Janos Street, RO-400028 Cluj-Napoca Romania

<sup>b</sup> University of Pannonia, Institute of Materials Engineering, 10 Egyetem Street, H-8200 Veszprem, Hungary

<sup>c</sup> University of Oradea, Medicine and Pharmacy Faculty, Department of Pharmacy, 29 Nicolae Jiga Street, RO-410028, Oradea, Romania

<sup>d</sup> INCDO-INOE 2000, Research Institute for Analytical Instrumentation, 67 Donath Street, RO-400293 Cluj-Napoca, Romania

\* Corresponding author: oana.cadar@icia.ro

## INTRODUCTION

The biocompatibility of orthopedic or dental metallic implants can be significantly enhanced using bioceramics as coating materials [1]. The biomorphic implants minimize the bone resorption, as well as inflammations using coatings containing anti-inflammatory drugs such as ibuprofen (IBU) or ketoprofen [2]. Also, these biomorphic materials can improve bone regeneration [3]. Due to their similar behaviors to bones in terms of elasticity, lightness and strength, the natural woods were used for biomorphic implants [4]. Also, the animal bones such as cattle bones could be a better alternative in order to evaluate the bone-implant interactions [5]. Hydroxyapatite (Hap) is a naturally occurring mineral form of calcium apatite and has been identified as the major mineral constituent of bones and teeth [6, 7]. Hap is a non-toxic, non-allergenic, non-mutagenic, osteoconductive, bioactive and biodegradable material with applications like coatings on different types of implants, carriers for drug delivery systems [8, 9], water purification, chromatography, food industry, gene therapy and anti-cancer drugs [10]. The synthesized Hap (molar ratio of Ca/P=1.67) has similar chemical composition and crystal structure to Hap in the human skeletal system. Recently, great attention has been paid to the ions (Na, Mg, K, Sr, Zn, Ba, Cu, Al, Fe, F, Cl and Si) substituted Hap in order to create chemical compositions with more similarity to natural bone mineral [9, 11]. Furthermore, the mechanical properties of Hap can be improved by adding carbon nanotubes (CNTs), but their toxicity still represents a major challenge, numerous conflicting studies demonstrating both toxic and non-toxic behavior [12, 14]. The synthesized CNTs tend to agglomerate due to van der Waals forces and can be well-dispersed in water or organic solvents, without physical or chemical modification [15, 16]. CNTs were discovered by Jijima in 1991 and can be categorized into single-wall carbon nanotubes (SWCNTs), double-wall carbon nanotubes (DWCNTs) and multi-wall carbon nanotubes (MWCNTs) [13, 16]. Besides Hap, the human skeletal system also contains gelatin (G), a natural polypeptide with biocompatible, biodegradable and non-immunogenic characteristics. G can be obtained from animal bones collagen by hot water extraction, followed by sol-gel transformation at 35 °C [17]. Due to their high specific surface and biocompatibility, G-CNT-Hap nanohybrids can be used as coatings and for drug delivery systems [17].

IBU is a non-steroidal anti-inflammatory drug; racemic IBU and S-enantiomer are used to reduce inflammations, fever and mild to moderate pain by blocking enzymes and protein in the human body [18, 19]. IBU has been frequently employed as a model drug for sustained/controlled release due to its short biological half-life (2h), suitable molecule size (0.6-1.0 nm)

and good pharmacological activity [20]. The absorption of IBU on activated carbon surface is an endothermic process, but the adsorption mechanism is not completely elucidated. IBU can form aggregates before the adsorption [21]. The time of drug release depends on the strength of hydrogen bonding between drug and carrier molecules, as well as the molecular size of drug [8, 9].

Various studies reported Hap coating on metallic (orthopedic or dental) implants due to their chemical and mechanical properties closer to those of human bone, good biocompatibility; however, the release of ions in the blood stream could be harmful to the patient, inducing allergic, inflammatory or carcinogenic responses [22]. The internal structure and surface properties of systems facilitates the therapeutic activity by maintaining drug release over the minimum inhibitory concentration for a long period of time. Furthermore, the rough structure of the scaffolds facilitates the drug penetration through [23].

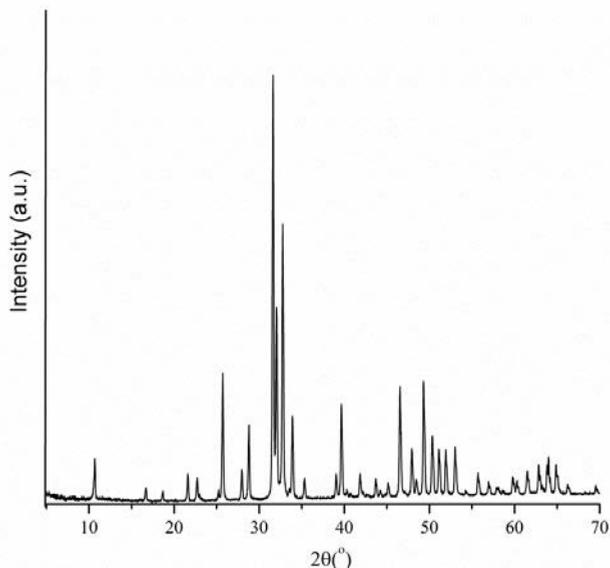
The synthesis reproducibility, the quality and properties of the reaction product could be improved by a continuous monitoring and a precise control using IoT (Internet of Things), a virtually connected network of physical devices through the infrastructure of Internet [24]. IoT is appropriate for automating the chemical procedures and to use the data collected from experiments, being an emerging, low cost and easy-to-use method.

The aim of this study was the production and characterization of biomorphic implants consisting in cattle bones coated with new Hap-based nanocomposites (NCs). For comparison, molded NCs were also produced and characterized. The proposed implants (Hap coatings on a material with very similar structure to those of human bone) have good potential as they can exploit the biocompatible and bonding properties of Hap, while using the mechanical properties of substrate (animal bone). Furthermore, these materials can be used to stimulate bone regeneration or to fill defects in bone [25]. The adsorption and desorption properties of these samples were also investigated by using IBU as an anti-inflammatory agent.

## **RESULTS AND DISCUSSION**

### **X-ray diffraction (XRD)**

The characteristic lines of pure Hap (JSPDS file No. 09-432) were observed in the XRD spectrum of the sintered cattle bone powders (Figure 1).



**Figure 1.** XRD patterns of Hap of the sintered bones.

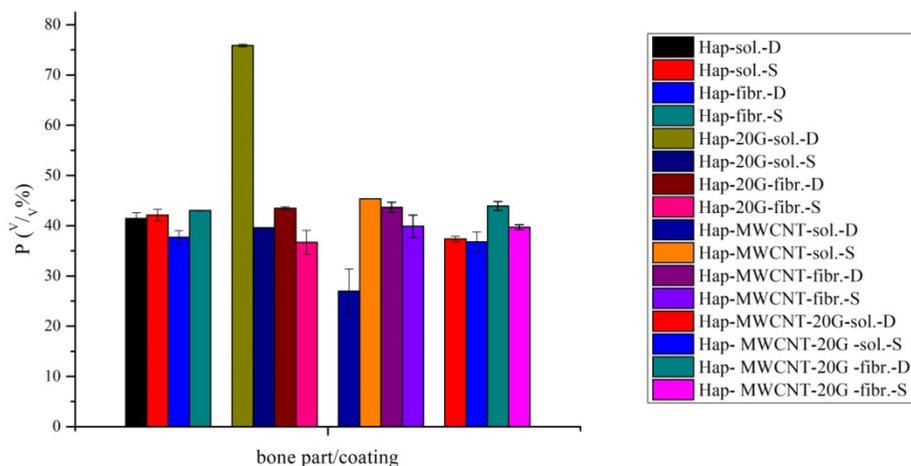
### **Thermal analysis**

The thermal analysis of the sintered samples at 900 °C revealed no significant weight loss or exothermic/endothermic effects for decomposition or other transformation processes, while for dried samples at 105 °C, the combustion of fMWCNT between 400 and 600 °C was observed [12].

### **Apparent porosity**

The apparent porosity (P) of samples can influence their mechanical strength, presence of structural defects and permeability [26].

Figure 2 shows that the solid bone part coated with Hap-20G-D exhibits the highest apparent porosity, but it has to be taken into consideration that the sample was dried and G could potentially form hydrogen bonds with surroundings water molecules. G is a hydrophylic substance and the three-dimensional structure is stabilized by both –NH and –COOH (-OH) groups which can form hydrogen bonds with other G chains and water molecules [27]. The dried bone-coated samples had the highest apparent porosity. Sintered samples obviously had lower apparent porosity because the sintering process decreases the porosity of materials; the number of OH-groups is also lower due to the thermal treatment of the samples.



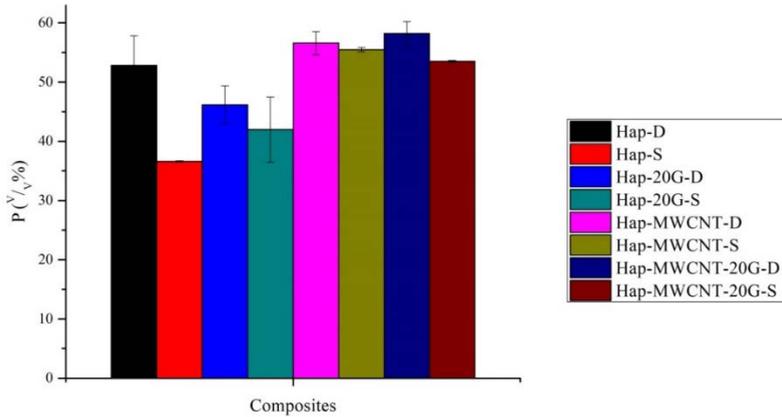
**Figure 2.** Apparent porosity of Hap based NCs - coated cattle bones.

In the case of molded NCs, dried samples exhibited the highest apparent porosity (Figure 3); this effect can be attributed not only to the opened pores, but to the hydrogen bonds between –OH groups and water used for measurement [28]. In the case of samples containing CNT, the apparent porosity increased. The highest porosity was achieved by the dried, molded NCs containing three components: Hap, CNT and G. Among the sintered samples, the Hap-MWCNT-S sample had the highest (and stable) porosity. The porosity measurements revealed higher porosity for NCs-coated cattle bones comparing to molded NCs. These results are in good agreements with SEM images.

### Specific surface area (BET)

According to BET measurements, the dried NCs (Hap-MWCNT-D – 206 m<sup>2</sup>/g) had larger specific surface area than the sintered NCs (Hap-MWCNT-S – 7.20 m<sup>2</sup>/g). A possible explanation for this difference is that MWCNTs were combusted from the structure during sintering at 900 °C resulting in lower specific surface area [12].

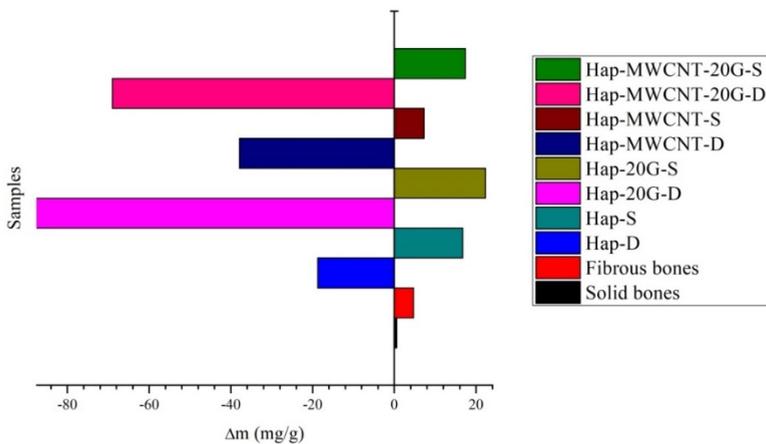
In the case of dried samples containing three components (Hap, MWCNT and G – 119 m<sup>2</sup>/g), the specific surface area was lower than in the case of samples containing two components, which can be explained by the different interactions between the functional groups of G and Hap.



**Figure 3.** Apparent porosity of molded NCs.

### X-ray Fluorescence (XRF)

XRF was used to monitor Ca and P content (mass change due to the ion transfer). The dried samples had a mass loss, but the sintered samples had a mass increase, because a new Hap layer was formed on their surface from the SBF (Simulated Body Fluid) [29]. The dried samples were not stable, their weight loss could be explained by the partial dissolution of the samples in the SBF. In the case of bone coated samples (black and red columns), the solid part did not really change, but the fibrous parts' weight increases due to the new Hap layer deposition in the pores (Figure 4).

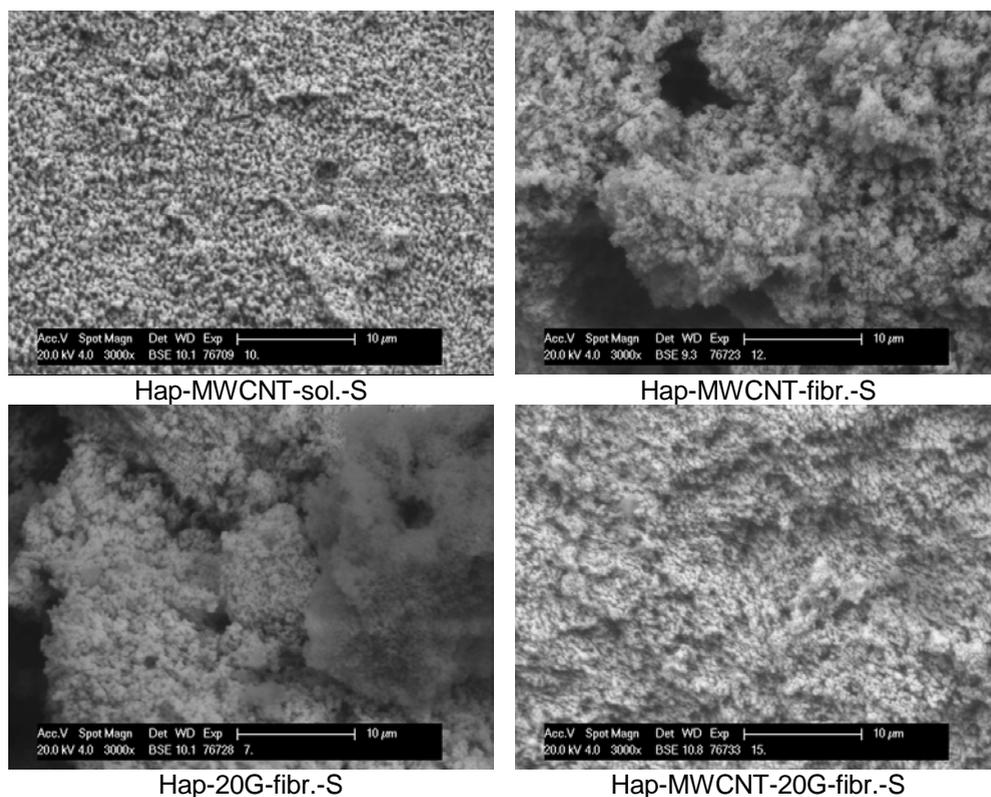


**Figure 4.** Mass change of NCs after 7 days of soaking in SBF.

Based on these results, it can be concluded that the behavior of Hap-based NC in SBF solution strongly depends on the heat treatment of the samples: for dried samples, occurred a partial dissolution, while on the surface of the sintered samples, a new Hap layer formed.

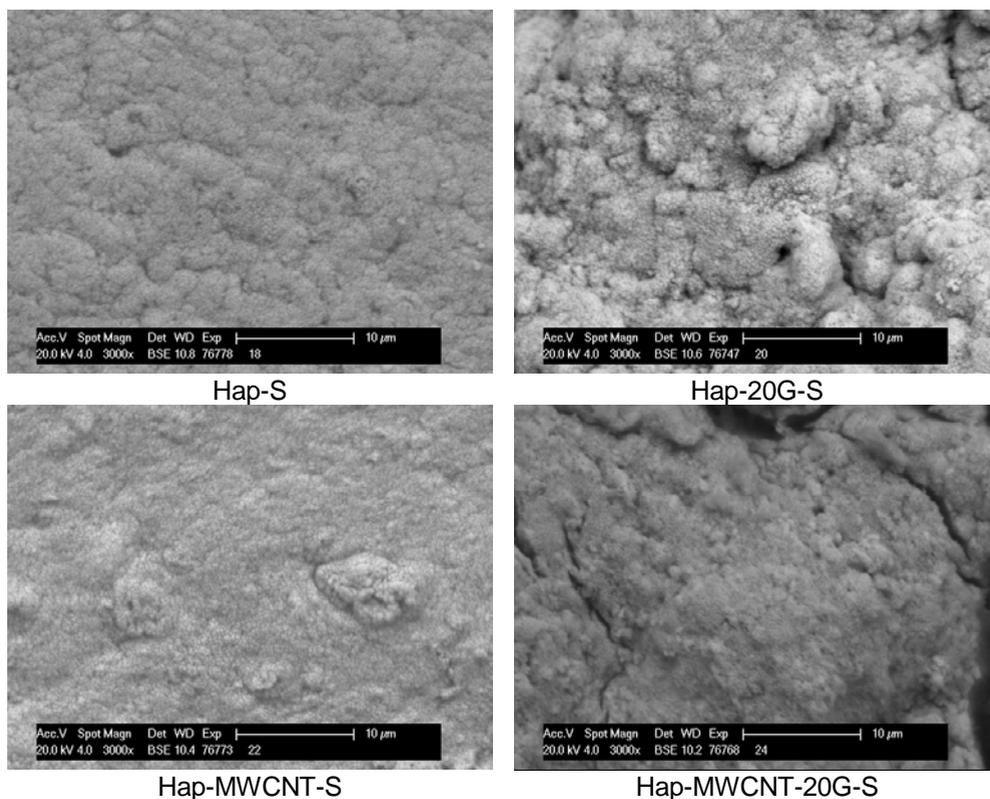
### Scanning electron microscopy (SEM)

SEM images of Hap based NCs - coated cattle bones and molded NCs are presented in Figures 5 and 6.



**Figure 5.** SEM images of Hap based NCs - coated cattle bones.

The SEM images revealed a difference in porosity: Hap based NCs used as coatings for cattle bones are more porous, while the molded NCs (no substrate) are compact with low porosity. Hap based NCs used as coatings for cattle bones mimic the natural bone surface structure favoring the growth of neighbouring tissues in the coating pores.



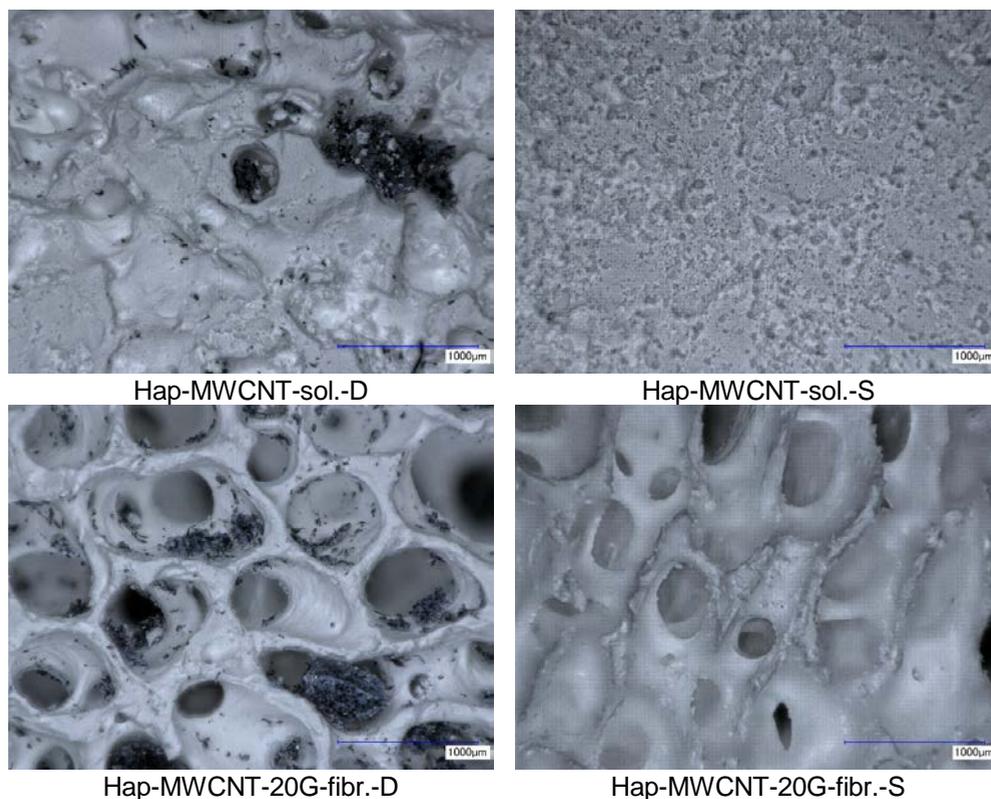
**Figure 6.** SEM images of molded NCs.

### **Light microscopy**

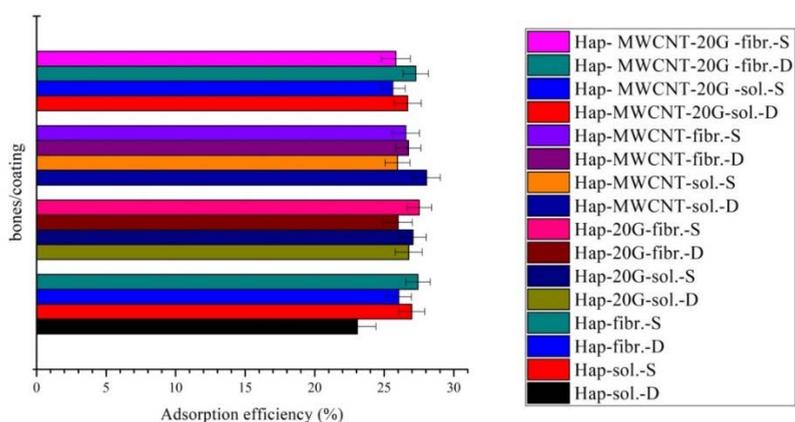
The light microscopy images of Hap based NCs - coated cattle bones are shown in Figure 7, revealing the heterogenous character of dried samples. In the case of sintered samples, the light microscopy images confirm that the MWCNTs were combusted between 400 and 600 °C [12].

### **Adsorption studies**

In this study, four types of coatings and two types of cattle bones (fibrous and solid) were obtained, resulting four groups of samples. In case of NCs - coated cattle bones, the adsorption capacity depended on the following parameters: bone part type, coating composition and post-heat treatment (Figure 8).



**Figure 7.** Light microscopy images of Hap based NCs - coated cattle bones.



**Figure 8.** Adsorption efficiency ( $\eta$ ) of Hap based NCs - coated cattle bones.

The highest adsorption efficiency of every group was selected (Table 1).

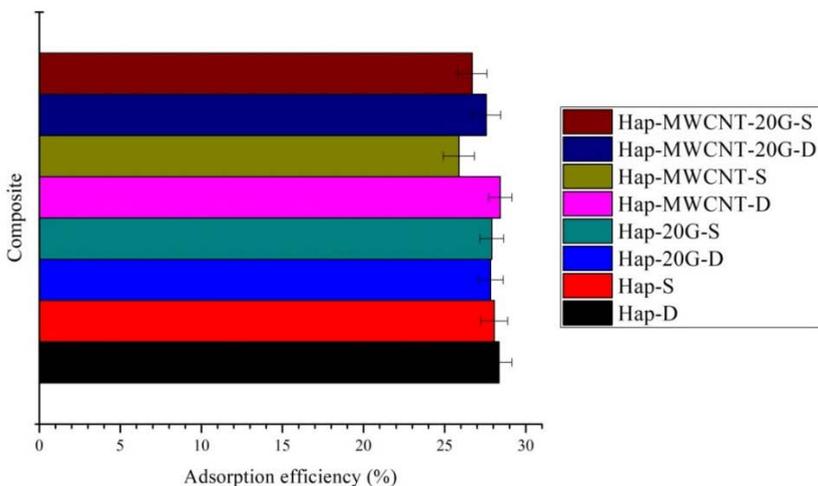
**Table 1.** The highest adsorption efficiency of Hap based NCs - coated cattle bones.

Type of composite used for coating (group of sample)	The highest adsorption efficiency (type of sample)	Adsorption efficiency (%)
Hap	Hap-fibr.-S	27.4
Hap-20G	Hap-20G-fibr.-S	27.5
Hap-MWCNT	Hap-MWCNT-sol.-D	28.0
Hap	Hap-MWCNT-20G-fibr.-D	27.3

BET analyses also confirmed that Hap-MWCNT had the highest specific surface area. XRF analyses indicated that the dried NCs were partially soluble in SBF, but the sintered samples were stable in SBF. As a result, although the Hap-MWNT-sol.-D sample had the highest adsorption efficiency, it could be not considered stable in SBF. After sintering (Hap-MWNT-sol.-S), the CNTs were combusted and the specific surface area of the samples decreased.

Both samples were stable and had high adsorption efficiency: Hap-fibr.-S (27.4%) and Hap-20G-fibr.-S (27.5%). So, the adsorption efficiency also depends on the cattle bone structure: as expected, the fibrous samples had higher adsorption efficiency than the solid ones.

In the case of molded NCs, Hap-MWCNT-D exhibited the highest adsorption efficiency (28.4%) (Figure 9). XRF analyses showed that NCs without sintering were not stable, the highest adsorption efficiency was achieved by the samples: Hap-S (28.1%) and Hap-20G-S (27.9%).



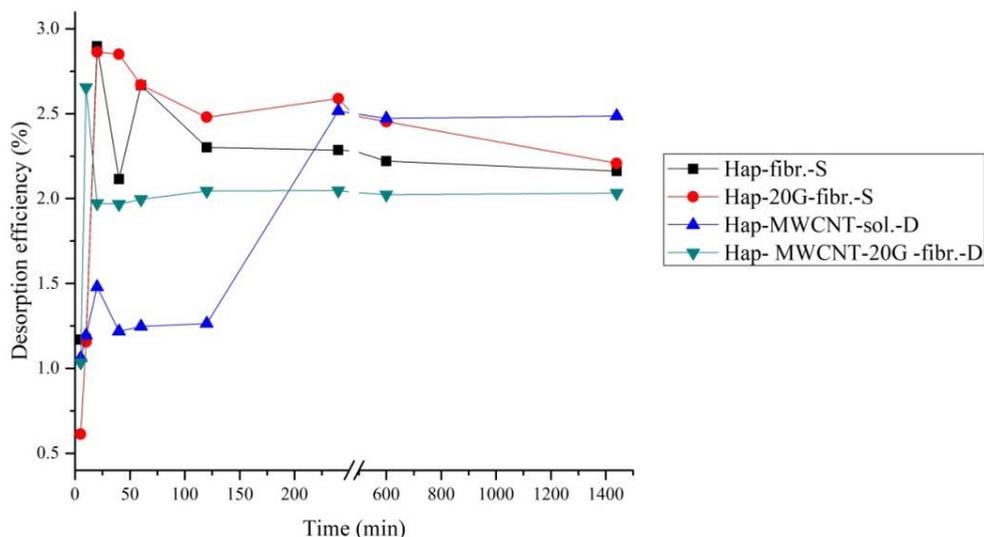
**Figure 9.** Adsorption efficiency ( $\eta$ ) of molded NCs.

The adsorption efficiency values in the case of dried and sintered samples were close to each other, so it can be concluded that the sintering process did not reduce essentially the adsorption capacity of the samples.

The highest adsorption efficiency and stability were obtained by the Hap-S and Hap-20G-S molded NCs and bone parts with Hap-S and Hap-20G-S NCs coatings. So, the same NC showed the most appropriate results in both cases.

### Desorption studies

In the case of cattle bones-coated NCs, the highest IBU release were observed for samples with the highest adsorption efficiency (Figure 10). A scale break was introduced, because the values between 240 and 600 min remained quasi-constant. Based on the adsorption capacity and stability in SBF, the IBU release was studied for Hap-fibr.-S and Hap-20G-fibr samples (Table 2). After 24 h, similar results were obtained for Hap-fibr.-S (2.16 %) and Hap-20G-fibr.-S (2.20 %).

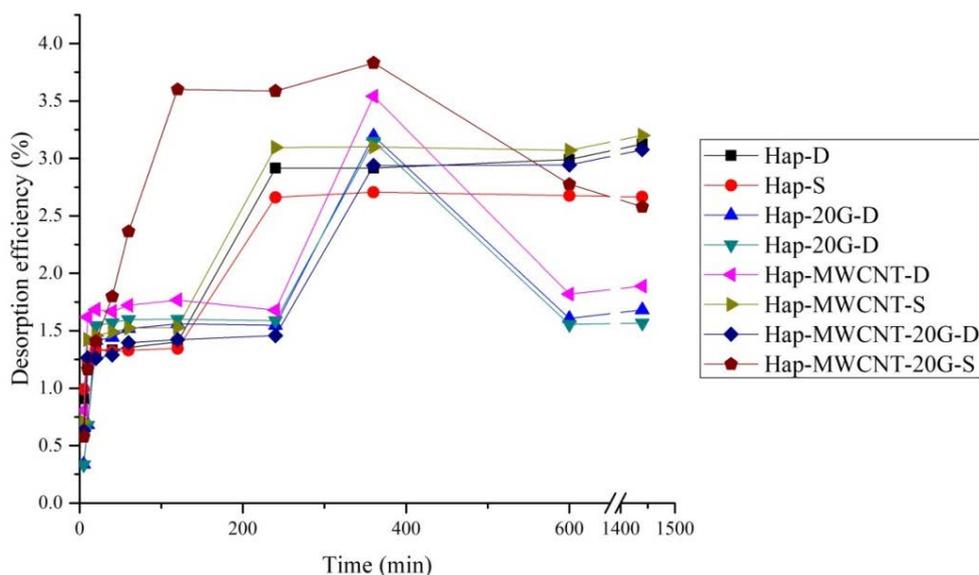


**Figure 10.** IBU release from Hap based NCs - coated cattle bones.

**Table 2.** IBU release of four cattle bone - coated samples.

Sample	Adsorption efficiency (%)	Desorption efficiency (%)
Hap-fibr.S	27.4	2.16
Hap-20G-fibr.-S	27.5	2.20
Hap-MWCNT-sol.-D	28.0	2.49
Hap-MWCNT-20G-fibr.-D	27.3	2.03

Based on XRF and adsorption (stability in SBF) results, Hap-S, Hap-20G-S, Hap-MWCNT-S, and Hap-MWCNT-20G-S were chosen as relevant to investigate the desorption efficiency (Figure 11). A scale break between 600 and 1440 min on x-axis was introduced, because the values at 120, 240 and 360 min remained quasi-constant. Hap-20G-S had the slowest IBU release (retard effect), after 24 h, only 1.57% of IBU was desorbed.



**Figure 11.** Desorption efficiency of molded NCs.

Regarding the desorption process, the difference in behavior of bone and boneless samples can be explained by the different surface morphology on which IBU was adsorbed. The different surface morphology was also confirmed by SEM images.

For the molded NCs, a rapid decrease of desorption after 5 to 10 h was observed, after which the process stabilized and increased steadily. This variation can be attributed to equilibrium processes in the solvent, desorption being an oscillating process until reaching equilibrium [35].

Table 3 shows a comparison between the adsorption and desorption efficiency of both types of samples (molded NCs and cattle bones with coating).

**Table 3.** The best results for IBU release – molded NCs.

Sample	Adsorption efficiency (%)	Desorption efficiency (%)
Hap-fibr.S	27.4	2.16
Hap-20G-fibr.-S	27.5	2.21
Hap-S	28.1	2.68
Hap-20G-S	27.9	1.58
Hap-MWCNT-S	25.9	3.20
Hap-MWCNT-20G-S	26.7	2.59

The best results (high adsorption efficiency and slow release - low desorption capacity) were obtained for molded and sintered Hap-20G-S NCs. In the case of Hap based NCs - coated fibrous cattle bones, a low desorption capacity due to the porous character of bones was observed.

## CONCLUSIONS

The purpose of this study was the preparation and characterization of NCs containing Hap, MWCNT and G in different ratios and their application as coatings for cattle bones (solid and fibrous) and for comparison molded NCs were also prepared. In the case of samples dried at 105 °C, the presence of CNTs increased the absorption capacity, while in the case of samples sintered at 900 °C, CNTs were combusted at 400-600 °C, resulting in a decrease of the absorption capacity. However, the sintered samples were more stable in SBF than dried ones. As a result, future studies to find the optimal sintering temperature to provide both the presence of CNTs and a good stability of samples in SBF are required. HAP-S and Hap-20G-S samples (NCs - coated cattle bones and molded NCs) presented high adsorption capacity. According to porosity measurements and SEM images, the NCs - coated cattle bones exhibited higher porosity than molded NCs. Comparing the obtained results for NCs - coated cattle bones and molded NCs, it can be concluded that the quality of coating depends not only on the coated material but also on the quality and structure of support (solid or fibrous). The studied NCs are materials with promising applications in engineering of bone implants due to their excellent chemical and structural properties.

## EXPERIMENTAL SECTION

### Materials

All reagents were of analytical grade (Merck, Germany) and used without further purification. Pepsin from porcine gastric mucosa (P7125) and trypsin from porcine pancreas (T7409) were purchased from Sigma Aldrich

(USA). The multiwalled carbon nanotubes (MWCNTs) were synthesized at 700 °C by chemical vapor deposition process (purity ~90 wt.%, average diameter 10-20 nm, length >50 μm) [31].

### Preparation of Hap NC

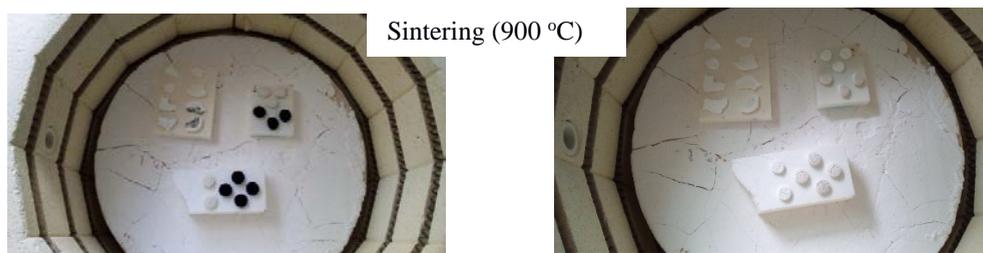
The cattle leg bones Alfa-Lox Bt., Hungary were calcined at 950 °C for 1 h to remove the organic part. The calcined bones were cut into pieces of approx. 1.0-1.5 cm length, boiled with trypsin and pepsin and washed with water. The protein-free bones were cleaned with phosphoric acid 85% and water.

The Hap-based NCs (Table 1) were prepared by precipitation in a similar way to the synthesis of pure Hap [12]. The solutions of 0.5 mol/L  $\text{Ca}(\text{NO}_3)_2$  and 0.3 mol/L  $(\text{NH}_4)_2\text{HPO}_4$  were prepared in order to meet the stoichiometric Ca/P = 1.67. G or/and MWCNTs were added to a 0.5 mol/L  $\text{Ca}(\text{NO}_3)_2$  solution and the pH was adjusted to 11 by adding a 25% ammonia solution, under constant stirring, at room temperature, resulting solution A. After the pH adjustment, a 0.3 mol/L  $(\text{NH}_4)_2\text{HPO}_4$  solution was added dropwise to solution A under magnetic stirring, at room temperature. The resulted solution was mixed for 22 h and the precipitate was filtered and washed with distilled water.

The process parameters were anytime/anywhere monitored using an in-house “IoT-smartbox” equipped with temperature sensor, heating element, pH-meter and peristaltic pump. The main advantages of “IoT-smartbox” are: (i) the safety-related control by automatically turned off if the critical temperature is reached or sending an alert message through SMS and email (in “safe zone”) and (ii) the possibility to track the history of the process.

### Preparation of Hap based NCs - coated cattle bones

The solid and fibrous parts of bones were placed under vacuum (680 mmHg) for 15 min to remove the air pollutants. Then, each bone part (approx. 1 cm) was soaked in a 1 g cm<sup>-3</sup> mixture (composite/ethanol), for 30 min. The resulted samples were: (i) only dried at 105 °C for 24 h (D) or (ii) dried at 105 °C for 24 h and sintered at 900 °C for 1 h (S) (Figure 12, Table 4).



**Figure 12.** Dried/sintered Hap based NCs - coated cattle bones and molded NCs.

### Preparation of molded Hap-based NCs

For comparison, the obtained Hap-based NCs were molded using round shape silicone forms with diameter of 2 cm, (i) only dried at 105 °C for 24 h (D) or (ii) dried at 105 °C for 24 h and sintered at 900 °C for 1 h (S) (Table 1).

### Adsorption of IBU

The samples (Hap based NCs - coated cattle bones and molded NCs) were soaked in IBU solution (17 g IBU /L ethanol, the concentration which grants the maximum adsorption capacity) [32]. After adsorption (24 h), the solid parts were separated by centrifugation, rinsed with ethanol and dried at 40-50 °C, for 24 h. The concentration of IBU in the liquid part was investigated using a UV-VIS spectrophotometer at 272 nm. The experiments were carried-out in triplicate. The adsorption efficiency,  $\eta_a$  (%), was calculated using Eqs. 1 and 2.

$$Q = \frac{m_a}{m} \quad (1)$$

$$\eta_a = Q \times \frac{m}{c_0 \times V} \times 100 \quad (2)$$

where: Q - adsorption capacity (mg/g),  $m_a$  - adsorbed mass of IBU (mg), m - mass of sample (g),  $c_0$  - initial IBU concentration (g/L) and V - volume of IBU solution used for adsorption (mL).

**Table 4.** Synthesized Hap-based NCs - coated cattle bones.

No	Hap	fMWCNT (% wt.)	G (% wt.)	Composite	Bone type	Operation	Implant type (abbreviations)
1	Hap	0	0	Hap	Solid	Drying	Hap-sol.-D
						Sintering	Hap-sol.-S
					Fibrous	Drying	Hap-fibr.-D
						Sintering	Hap-fibr.-S
2	Hap	0	20	Hap-20G	Solid	Drying	Hap-20G-sol.-D
						Sintering	Hap-20G-sol.-S
					Fibrous	Drying	Hap-20G-fibr.-D
						Sintering	Hap-20G-fibr.-S
3	Hap	10	0	Hap-MWCNT	Solid	Drying	Hap-MWCNT-sol.-D
						Sintering	Hap-MWCNT-sol.-S
					Fibrous	Drying	Hap-MWCNT-fibr.-D
						Sintering	Hap-MWCNT-fibr.-S
4	Hap	10	20	Hap-MWCNT-20G	Solid	Drying	Hap-MWCNT-20G-sol.-D
						Sintering	Hap-MWCNT-20G-sol.-S
					Fibrous	Drying	Hap-MWCNT-20G-fibr.-D
						Sintering	Hap-MWCNT-20G-fibr.-S

### Desorption of IBU

The release of IBU was investigated by soaking each sample in 20 mL SBF (pH=7.4) at room temperature. SBF was prepared according to Kokubo method [33, 34]. 2 mL sample was removed at different time intervals,  $t$  (5, 10, 20, 30, 60, 120, 240, 360, 600 and 1440 min) and immediately replaced with an equal volume of fresh SBF. The amount of IBU released was measured using a UV-VIS spectrophotometer at 224.5 nm. The experiments were carried-out in triplicate. The desorption efficiency,  $\eta_d$  (%), was calculated using the amount of IBU adsorbed and the amount of IBU released in SBF.

### Characterization

XRD analysis was performed at room temperature, using a PW3710 X'Pert X-ray diffractometer (Philips, Netherlands), operating at 40 kV, 30 mA with  $\text{CuK}\alpha$  radiation ( $\lambda=1.542 \text{ \AA}$ );

The thermal analysis was performed using a Q-1500 D Derivatograph (MOM, Hungary) in Pt crucibles using  $\alpha\text{-Al}_2\text{O}_3$  as inert material, by heating from room temperature to 1000 °C (5 °C/min);

The apparent porosity ( $P$ ), that represents pores accessible to the fluid used in the test, was determined using the Archimedes method based on the principle that a solid with certain bulk immersed in a liquid displaces an amount of liquid equal to the bulk of the liquid when immersed: The following equation was used to calculate the apparent porosity (open pores).

$$P = \left( \frac{m_{\text{sat}}}{m_{\text{dr}}} - \frac{m_{\text{sat}}}{m_{\text{sub}}} \right) \times 100 \quad (5)$$

where:  $m_{\text{dr}}$  - the mass of the dry sample (g),  $m_{\text{sat}}$  - the mass of the sample saturated with liquid (g) and  $m_{\text{sub}}$  - the mass of the sample submerged in liquid (g) [35];

The specific surface area was determined using Asap 2000 equipment (Micromeritics Instruments Inc., USA) from data on nitrogen adsorption; The morphological characteristics of NCs was examined by a XL30-ESEM microscope (Philips, Netherlands) at 20 kV using a Keyence-VHX 2000 Light microscope;

The mass change of NCs was investigated by XRF. The samples were soaked in SBF for 7 days, at 37 °C and removed after 1, 3 and 7 days. The Ca and P release was monitored using an energy dispersive X-ray Fluorescence (XRF) MiniPal4, PANalytical operating at 14 kV, 202  $\mu\text{A}$ . The Ca and P content (mass change due to the ion transfer) was calculated considering the Ca and P content of SBF;

The IBU adsorption-desorption studies were performed using a V-560 UV-VIS spectrophotometer (Jasco, Japan).

## ACKNOWLEDGMENT

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## REFERENCES

1. O. Gryshkov, N.I. Klyui, V.P. Temchenko, V.S. Kyselov, A. Chatterjee, A.E. Belyaev, L. Lauterboeck, D. Iarmolenko, B. Glasmacher, *Materials Science and Engineering C*, **2016**, *68*, 143.
2. A.I. Pearce, R.G. Richards, S. Milz, S.G. Pearce, *European Cells & Materials*, **2007**, *13*, 1.
3. Y. Li, S.K. Chen, L. Li, L. Qin, X.L. Wang, Y.X. Lai, *Journal of Orthopaedic Research*, **2015**, *3*, 95.
4. G. Filardo, E. Kon, A. Tampieri, R. Cabezas-Rodriguez, A. Di Martino, M. Fini, G. Giavaresi, M. Lelli, J. Martinez Fernandez, L. Martini, J. Ramirez-Rico, F. Salamanna, M. Sandri, S. Sprio, M. Marcacci, *Tissue Engineering Part A*, **2014**, *20*, 763.
5. N. Baldini, M. De Sanctis, M. Ferrari, *Dental Materials*, **2011**, *27*, 61.
6. D. McConnell, "Carbonate Apatites", pp. 39-47 in *Apatite: Its crystal chemistry, mineralogy, utilization, and geologic and biologic occurrences*, Springer-Verlag, New York, **1973**.
7. T. Sakae, H. Nakada, J.P. LeGeros, *Journal of Hard Tissue Biology*, **2015**, *24* 111.
8. K. Tomoda, H. Ariizumi, T. Nakaji, L. Makino, *Colloids and Surfaces B*, **2010**, *76* 226.
9. M. Cziko, E.S. Bogya, C. Paizs, G. Katona, Z. Konya, A. Kukovecz, R. Barabas, *Materials Chemistry and Physics*, **2016**, *180*, 314.
10. C.F. Dai, S.P. Li, X.D. Li, *Colloids and Surfaces B*, **2015**, *136*, 262.
11. R. Barabas, M. Cziko, I. Dekany, L. Bizo, E.S. Bogya, *Chemical Papers*, **2013**, *67*, 1414.
12. R. Barabas, G. Katona, E.S. Bogya, M.V. Diudea, A. Szentes, B. Zsirka, J. Kovacs, L. Kekedy-Nagy, M., Cziko, *Ceramics International*, **2015**, *41*, 12717.
13. S. Dadras, V.M. Farahani, *Physica B*, **2015**, *477*, 94.
14. D. Lahiri, S. Ghosh, A. Agarwal, *Materials Science and Engineering C*, **2012**, *32*, 1727.
15. A. Abrishamchian, T. Hooshmand, M. Mohammadi, F. Najafi, *Materials Science and Engineering C*, **2013**, *33*, 2002.
16. D. Gopi, E. Shinyjoy, M. Sekar, M. Surendiran, L. Kavitha, T.S. Sampath Kumar, *Corrosion Science*, **2013**, *73*, 321.

17. I.K. Yoon, J.Y. Hwang, J.W. Seo, W.C. Jang, H.W. Kim, U.S. Sing, *Carbon*, **2014**, *77*, 379.
18. S.D. Bergese, K. Candiotti, S.S. Ayad, S. Soghomonyan, T.J. Gan, *Clinical Therapeutics*, **2015**, *37*, 360.
19. T.J. Gan, K. Candiotti, A. Turan, A. Buvanendran, B.K. Philip, E.R. Viscusi, S. Soghomonyan, *Clinical Therapeutics*, **2015**, *37*, 368.
20. M. Oner, E. Yetiz, E. Ay, U. Uysal, *Ceramics International*, **2011**, *37*, 2117.
21. D. Musmarra, M. Prisciandaro, M. Capocelli, D. Karatza, P., Iovino, S. Canzano, A. Lancia, *Ultrasonics Sonochemistry*, **2016**, *29*, 76.
22. B.G.X. Zhang, D.E. Myers, G.G. Wallace, M. Brandt, P.F.M. Choong, *International Journal of Molecular Sciences*, **2014**, *15*, 11878.
23. P. Diaz-Rodriguez, A. Perez-Estevez, R. Seoane, P. Gonzalez, J. Serra, M. Landin, *ISRN Pharmaceuticals*, **2013**, *Article ID 104529*, 8 pages.
24. E. Brown, "Who Needs the Internet of Things?", Linux.com, Retrieved 23 October **2016**.
25. S.T. Ho, D.W. Hutmacher, *Biomaterials*, **2006**, *27*, 1362.
26. P. Spulveda, F.S. Ortega, M.D.M. Innocentini, V.C. Pandolfelli, *Journal of the American Ceramic Society*, **2000**, *83*, 3021.
27. A. Duconseille, T. Astruc, N. Quintana, F. Meersman, V. Sante-Lhoutellier, *Food Hydrocolloids*, **2015**, *43*, 360.
28. Z.C. Chen, X.L. Zhang, K. Zhou, H. Cai, C.Q. Liu, *Advances in Applied Ceramics*, **2015**, *114*, 183.
29. P. Sooksaen, N. Pengusawan, S. Karawatthanaworrakul, S. Pianpraditkul, *Advances in Condensed Matter Physics*, **2015**, *Article ID 158582*, 9 pages.
30. G. Neupane, R.J. Donahoe, Y. Arai, *Chemical Geology*, 2014, *368*, 31.
31. A. Szentés, G. Horvath, C., Varga, G. Harrach, *Hungarian Journal of Industry and Chemistry*, **2011**, *39*, 113.
32. E. Chevalier, M. Viana, S. Cazalbou, L. Makein, J. Dubois, D. Chulia, *Acta Biomaterialia*, **2010**, *6*, 266.
33. T. Kokubo, H. Kushitani, S. Sakka, T. Kitsugi, T. Yamamuro, *Journal of Biomedical Materials Research*, **1990**, *24*, 721.
34. C. Moisa, L.G. Vicas, M. Ganea, E.A. Levei, O. Cadar, C. Berce, *Farmacia*, **2018**, *66*, 176.
35. S.T. Ho, D.W. Hutmacher, *Biomaterials*, **2006**, *27*, 1362.

## IONIC EXCHANGE STUDIES FOR CORRECTING WATER QUALITY INDICATORS

SILVIA BURCĂ<sup>a</sup>, CERASELLA INDOLEAN<sup>a\*</sup>

**ABSTRACT.** For this study, a natural ionic exchanger (zeolitic volcanic tuff) and a synthetic anionic exchanger were considered, to remove iron and manganese ions, reduce COD, chlorides and conductivity values from water. It was worked with a zeolitic volcanic tuff cropped out from Macicas area, Romania, and a synthetic anionic exchanger Amberlite A-21. A groundwater and surface sources of drinking water from Ialomița-Muntenia (Romania) catchment were analysed, with iron and manganese contents (0.2527–0.3059 mg Fe<sub>tot</sub>/L, 0.0234 mg Mn/L), COD (14.530–17.064 mg KMnO<sub>4</sub>/L), chlorides contents (170.95 mg/L), pH=7.70, and with electrical conductivity of 1433-1451 μS/cm. The ionic exchange process has been achieved in batch and continuous-flow conditions. The process was realized in static regime, with good results, the highest efficiencies for iron and manganese ions removal being 99% and 79-95%, respectively.

**Keywords:** *municipal water source, food water, zeolitic volcanic tuff, ionic exchange*

### INTRODUCTION

The availability of adequate freshwater resources, is vital to food safety and production (primary such as irrigation, livestock watering, aquaculture and processing: ingredient, transport medium and hygiene aid) [1].

If the water is drinkable, then, it could be accepted for all food uses [2]. However, not all uses require that water to be of this standards and, where it is possible, it can use other sources of water or reuse water. Consequently, the water quality requirements will need to be appropriately tailored [3-6].

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<sup>a</sup> Babeș-Bolyai University, Faculty of Chemistry and Chemical Engineering, 11 Arany Janos str., RO-400028, Cluj-Napoca, Romania

\*Corresponding author: cella@chem.ubbcluj.ro

Water, like food, is a potential vehicle for the transmission of diseases, causing agents or chemicals toxic and/or dangerous for health. Water is capable of introducing contamination into food, if appropriate care is not taken [7]. Raw water can be contaminated with pathogens, usually as a consequence of human or animal faecal material or run-off contaminated with faecal material. It can also be contaminated with a wide range of chemicals, both natural and anthropogenic, which are of concern under some circumstances, depending on the concentrations present. Contamination can arise in raw water or as a consequence of improper storage or pick-up of contaminants from distribution systems. It can also occur as a consequence of leakage from a dirty water system into a clean water system. For example, drinking water was identified as the source of a significant and fatal outbreak of *Escherichia coli* O157:H7 in Canada [8].

In areas where groundwater flows through an organic rich soil, iron and manganese will also dissolve there. Natural sources of iron and manganese may include weathering of iron and manganese bearing minerals (olivine, magnetite, chromite, ilmenite, pyrrhotite, pyrite, hematite, hydrate, goethite, lepidokrokit, iron carbonate, vivianite, strengite) [9]. The soils can have iron content between 0.5% and 5%, depends on the rocks from where soil was derived, mechanism of transport and geochemical history. Iron and manganese release in water can have anthropogenic sources: industrial effluents, landfill leakages and acid mine drainage, but also well casing, pump parts, piping and storage tank [10].

The too high concentrations of  $Mn^{2+}$  will have as a consequence a metallic taste of water, staining of different products like clothes, paper and plastics [11].

Iron and manganese can cause different types of nuisance problems. The colour and flavour of food and water can be affected by iron and manganese, because they can react with tannins in coffee, tea and alcoholic beverages, which conduct to black sludge obtaining. Iron can cause reddish-brown staining of laundry, utensils, dishes and glassware [12]. Iron and manganese can also cause build up in pipelines, water heaters and pressure tanks. This build up is linked with a decrease in pressure and amount of available water and also increase in cost of water-using appliances [13]. The deposition of iron and manganese in the distribution systems can cause reduction of diameter of pipes and eventually clogging of those will take place [14].

The removal of dissolved iron and manganese from groundwater is generally accomplished by the oxidation and precipitation. The removal process is affected by the different chemical and physical characteristics of water: pH, temperature, total organic carbon (TOC) and concentration of dissolved oxygen [15]. It can use the ion exchange method, if the aim is to

remove small quantities of iron and manganese, because it will be the risk of rapid clogging. The process of iron and manganese removing by ion exchange is accomplished by using strong acid cations [16].

In the brewery industry, for example, water is an essential component for the final product quality, together with yeast, barley and hops, and breweries pay great attention on sourcing of this raw material. The production and processing of high quality water is a very important factor, due to its influence on both the production process (yeast requires potassium, sodium and calcium for optimum fermentation) and on the taste of the final product [17, 18]. Aside from these requirements, there is legislation governing water quality parameters in such industries, obliging brewers to use high drinking quality water for most production processes [19, 20].

In this study, zeolitic volcanic tuff and cationic exchanger were used to retain iron and manganese ions, and reduce the concentration of dissolved salts, in order to obtain adequate quality water for use in the brewery industry.

## RESULTS AND DISCUSSION

Samples of drinking water collected from Ialomița-Muntenia (Romania) catchment (groundwater and surface water sources) are characterized by a high content of dissolved salts, iron and manganese concentration, close to the maximum allowable value (MVA). The initial pH was around neutral. The presence of ammonium and nitrite ions in samples was not detected. The measured values of physico-chemical indicators are shown in Table 1.

**Table 1.** Physico-chemical indicators values for Ialomița-Muntenia (Romania) catchment

Water Sample	Fe <sub>tot</sub> mg/L	Mn <sup>2+</sup> mg/L	COD mg KMnO <sub>4</sub> /L	Cl <sup>-</sup> mg/L	Conductivity μS/cm	pH
Groundwater (GW)	0.305	0.023	14.536	177.90	1433	7.6
Surface water (SW)	0.252	0.023	17.064	170.95	1451	7.7

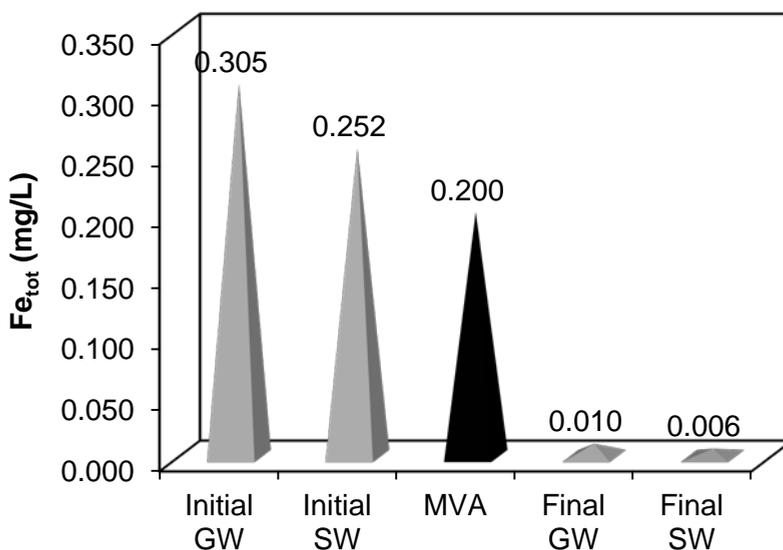
### *Batch mode analysis*

Experimental determinations, in static regime, showed that the equilibrium of the iron and manganese ions removal by ion exchange on ZVT-H was reached after 24 hours. Concentrations of the iron and manganese ions are very small in the samples collected, below the MVA, imposed by the Romanian legislation (50 μg Mn<sup>2+</sup>/L and 200 μg Fe<sub>total</sub>/L [21], Figure 1 and Figure 2).

High values of cationic exchange process efficiencies were found (95-97%, for the  $Fe_{tot}$  removal ions, respectively 79.5-95%, in the case of manganese ions).

Chloride ion concentrations, determined in water samples, collected at equilibrium, were found to be around 35.45-37.48 mg  $Cl^-/L$  and the efficiencies of anionic exchange process were between 77.77 and 79.25%.

Regarding at COD analysis of drinking water samples, this parameter has decreased from 14.53 mg  $KMnO_4/L$ , respectively 17.06 mg  $KMnO_4/L$  to 4.26 mg  $KMnO_4/L$  respectively 9.10 mg  $KMnO_4/L$  (values recorded at the end of the cationic/anionic exchange process), values found to be below the maximum allowed by Romanian law.

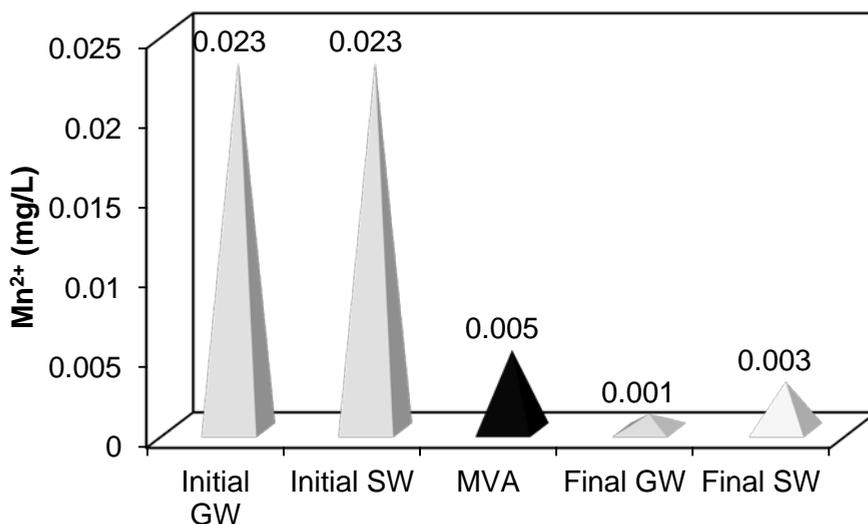


**Figure 1.** Initial, MVA and equilibrium iron ions concentration values, GW and SW samples, cationic exchange, static regime, 2 g ZVT-H, 100 mL sample, room temperature.

Electrical conductivity, measured at the end of the cationic/anionic exchange process, in static regime, presents a sharply decreased from 1433  $\mu S/cm$  (SW) and 1451  $\mu S/cm$  (GW), to 428  $\mu S/cm$  (SW) and 514  $\mu S/cm$  (GW), respectively. MAV value, according to Romanian legislation, is 2500  $\mu S/cm$  [21].

*Fixed bed column analysis*

The breakthrough curves for  $\text{Fe}_{\text{tot}}$  and  $\text{Mn}^{2+}$  sorption on ZVT-H, at an inlet concentration of  $0.252 \text{ mg Fe}_{\text{tot}}/\text{L}$  and  $0.023 \text{ mg Mn}^{2+}/\text{L}$  are shown in Figure 3. As it was expected,  $\text{Fe}_{\text{total}}$  and  $\text{Mn}^{2+}$  sorption was very high, in 370-400 minutes, and then the concentration at the column outlet increased slowly, because the zeolitic volcanic tuff was getting saturated.

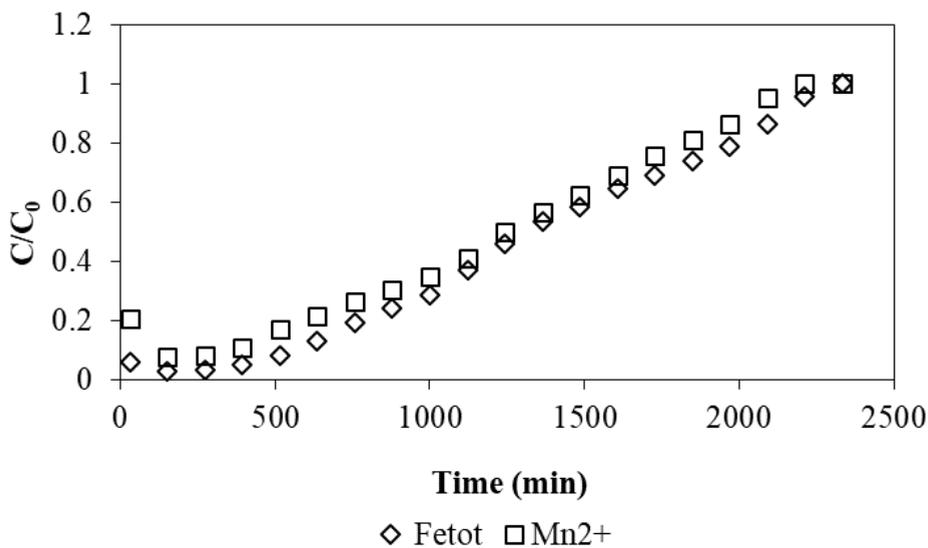


**Fig. 2.** Initial, MVA and equilibrium manganese ions concentrations, cationic exchange, static regime, 2 g ZVT-H, 100 mL sample, room temperature.

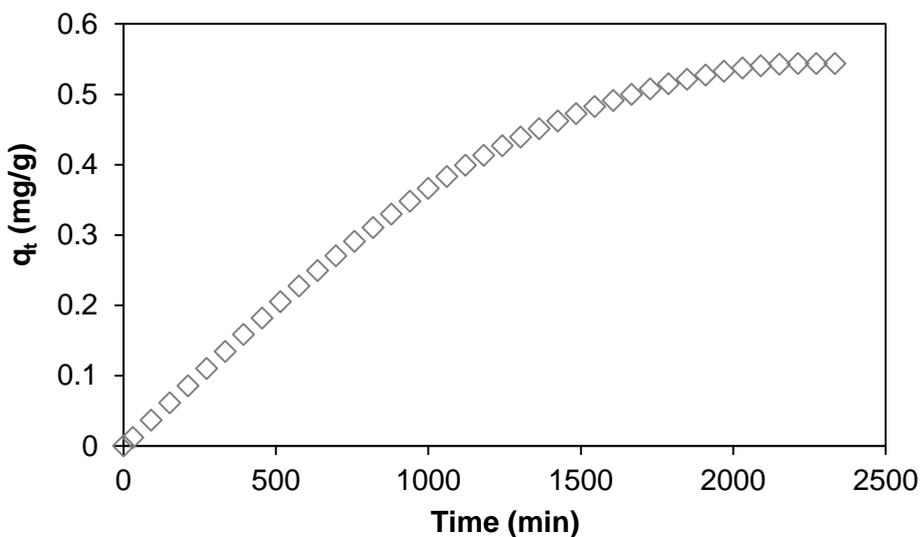
The values of the breakthrough time ( $t_b$ ) and exhaustion time ( $t_s$ ), of the breakthrough curves, obtained for iron and manganese ions were 10.6 h and 38.89 h respectively, for manganese ions sorption, 32.82 h and 36.86 h, respectively for iron ions sorption. It can be observed that the values of breakthrough time for manganese ions was almost three times smaller than breakthrough time for sorption of iron ions (10.6 h for manganese ions, compared to 32.82 h, for iron ions).

MVA for manganese ions has a lower value than the MVA for iron ions ( $0.005 \text{ mg Mn}^{2+}/\text{L}$ , compared to  $0.200 \text{ mg Fe}_{\text{tot}}/\text{L}$ ) [22].

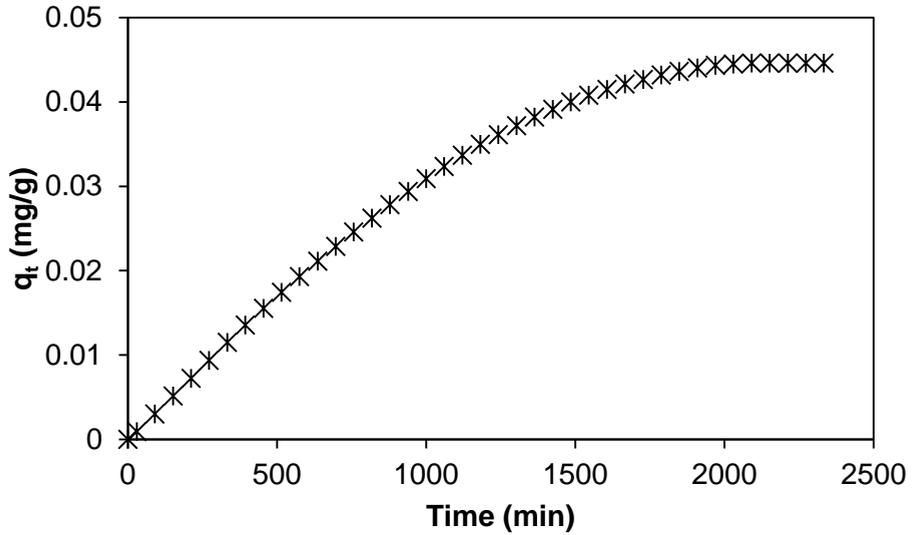
It was found that the maximum sorption capacity values of iron ions is higher than the maximum sorption capacity values for manganese ions ( $0.543 \text{ mg Fe}_{\text{tot}}/\text{g ZVT-H}$  compared to  $0.044 \text{ mg Mn}^{2+}/\text{g ZVT-H}$ , respectively), because the inlet concentration of iron ions is higher than inlet concentration of manganese ions (Figure 4 and Figure 5).



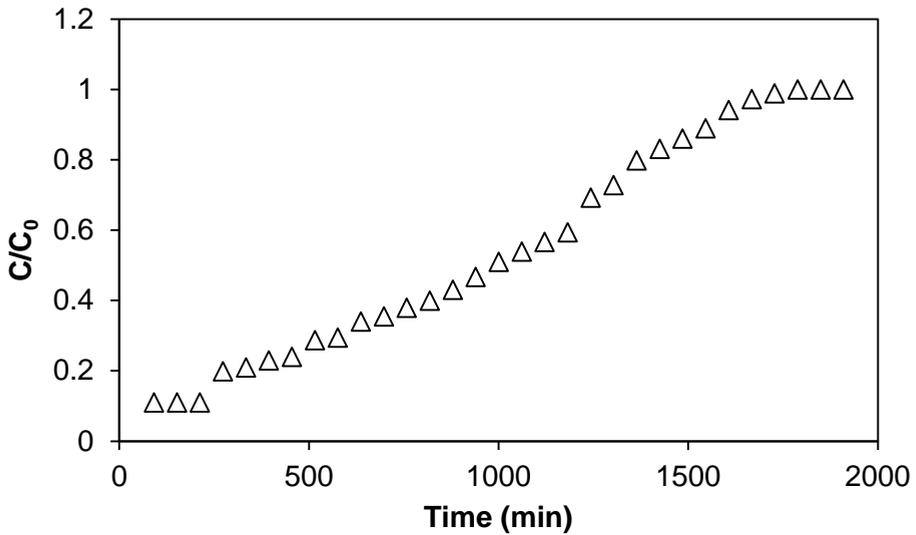
**Figure 3.** Breakthrough curves for iron and manganese ionic exchange process, on ZVT-H, dynamic regime.



**Figure 4.** Removal capacity for Fe<sub>tot</sub> sorption, 2 g ZVT-H, Q = 0.055 mL/s, C<sub>0</sub> = 0.252 mg/L.

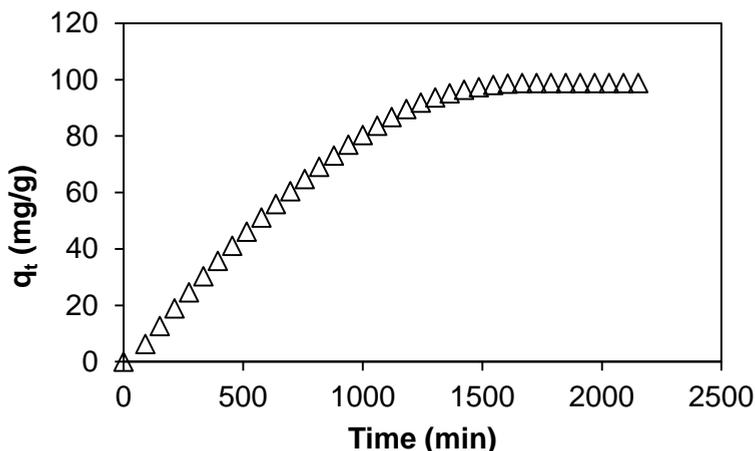


**Figure 5.** Removal capacity for  $Mn^{2+}$  sorption, 2 g ZVT-H,  $Q = 0.055$  mL/s,  $C_0 = 0.023$  mg/L



**Figure 6.** Breakthrough curve for  $Cl^-$  sorption, 2 g A-21 anionic exchanger,  $C_0 = 170.95$  mg  $Cl^-$ /L,  $Q = 0.055$  mL/s.

The breakthrough curve for  $\text{Cl}^-$  sorption, on synthetic resin Amberlite A-21, with an inlet concentration of  $170.96 \text{ mg Cl}^-/\text{L}$  is shown in Figure 6. The exit time value ( $t_s$ ) of the breakthrough curves obtained for chlorides ions was  $41.91 \text{ h}$ , MVA for chlorides ions in the drinking water being  $250 \text{ mg Cl}^-/\text{L}$  [22]. The maximum sorption capacity values of chlorides ions was  $98.85 \text{ mg Cl}^-/\text{g}$ , Figure 7.



**Figure 7.** Removal capacity for chlorides sorption,  $2 \text{ g A-21}$ ,  $Q = 0.055 \text{ mL/s}$ ,  $C_0 = 170.95 \text{ mg Cl}^-/\text{L}$ .

#### *Breakthrough curves modeling*

To describe the fixed bed column behaviour and to scale it up for industrial applications, three models – Thomas, Wolborska, and Yoon-Nelson – were used, in order to fit better the experimental data with the appropriate model.

#### *Thomas model*

This model is suitable for adsorption process where the external and internal diffusion limitations are absent, this is frequently applied to estimate the adsorptive capacity of adsorbent and predict breakthrough curves, assumes plug flow behavior in the bed, uses Langmuir isotherm for equilibrium and second order reversible reaction kinetics [23, 24].

Adsorption kinetics is described as the following equation:

$$\frac{\partial q}{\partial t} = k_1(q_0 - q)C - k_2q(C_0 - C) \quad (1)$$

where:  $k_1$  and  $k_2$  are velocity constants ( $s^{-1}$ )  
 $q_0$  is the initial exchange capacity (mg/g),  
 $C_0$  is the inlet concentration (mg/L).

By applying boundary conditions and linearization, the equation becomes:

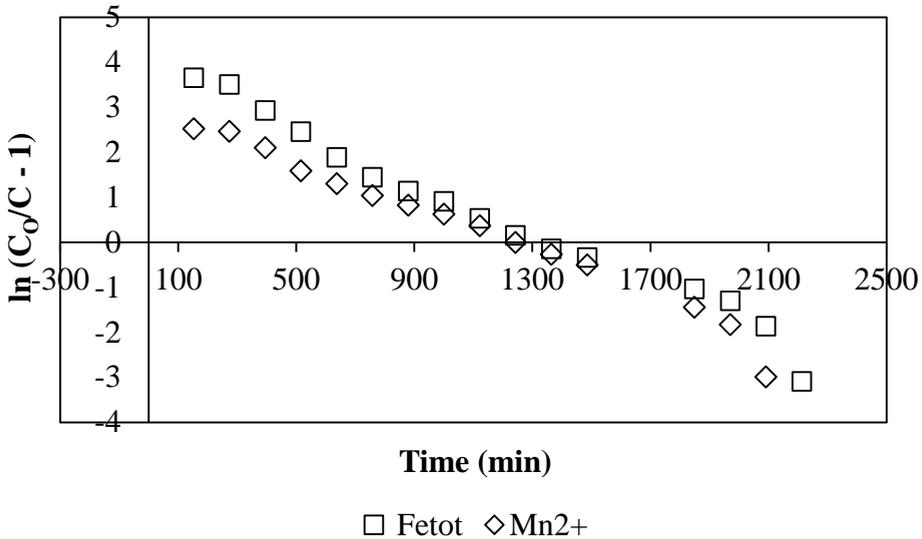
$$\ln\left(\frac{C_0}{C} - 1\right) = \frac{k_T q_0 m_c}{Q} - k_T C_0 t \tag{2}$$

where:  $k_T$  is the Thomas rate constant ( $dm^3/(s \cdot g)$ ),  
 $q_0$  is the adsorption capacity (mg/g),  
 $m_c$  is the ionic exchanger mass in the column (g),  
 $V_{eff}$  is the throughput volume (L),  
 $Q$  is the volumetric flow rate ( $dm^3/s$ ),  
 $t = V_{eff}/Q$ .

For the cationic/anionic exchange experimental data, a plot of  $\ln(C_0/C - 1)$  vs.  $t$  should give a linear relationship, if the above model is correct.

In Figure 8 is presented a linear regression of the cationic exchange experimental data, showing a  $R^2$  value of 0.9776. However, it is visible from the shape of the curve that the relationship is linear. For the anionic exchange process,  $R^2$  value was 0.9128.

The determined coefficients and relative constants, obtained for kinetics considered models are shown in Table 2.



**Figure 8.** Thomas model linear regression, cationic exchange.

*Wolborska Model*

Wolborska found that the breakthrough had two regions in which the migration rate of the concentration front is described in different ways [25]: low concentration region, in the range from  $10^{-5}$  to  $5 \times 10^{-2}$  of the normalized outlet concentration ( $C_{\text{outlet}}/C_{\text{inlet}}$ ) and high concentration region, containing the other range of the curve.

The model developed by Wolborska is exclusively for the low concentration region of the breakthrough curve and based on the following observations from Dubinin et al [26]: the initial concentration distribution is translocated along the column at a constant, velocity, the width of the breakthrough curve in the range of low concentration is constant, the low concentration area is characterized by constant kinetic coefficients, the process rate is controlled by the external mass transfer resistance [27].

Below is presented the continuity equation on the column:

$$\frac{\partial C}{\partial t} + u \frac{\partial C}{\partial h} + \frac{\partial q}{\partial t} = D \frac{\delta^2 C}{\delta h^2} \quad (3)$$

where: C is the adsorbate concentration (mg/L),

t is time (min),

u is the flow rate (L/min),

q is the adsorbate concentration in the solid phase (mg/g adsorbent),

D is axial diffusion coefficient,

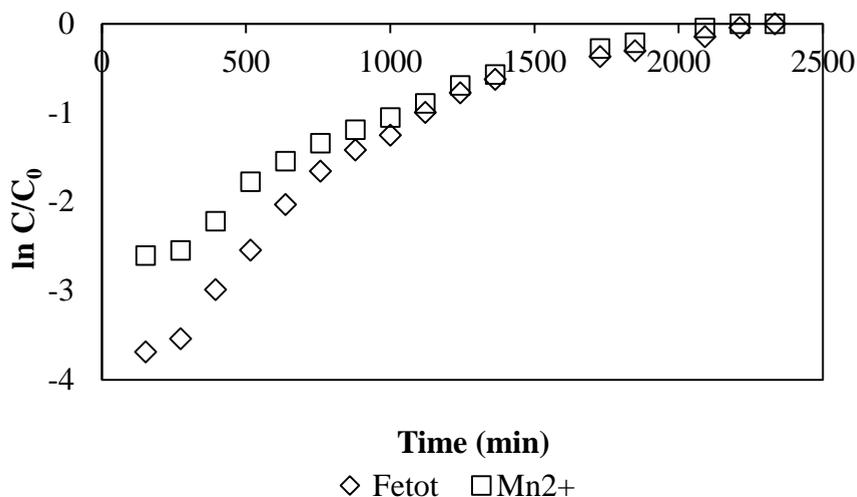
h is the distance from the column inlet (dm).

By applying boundary conditions and linearization, equation the equation becomes:

$$\ln \frac{C}{C_0} = \frac{\beta_a C_0}{q_0} t - \frac{\beta_a}{u} h \quad (4)$$

where  $\beta_a = \beta_0$  for process without axial diffusion and  $\beta_a = \frac{u^2}{2D} \left( \sqrt{1 + \frac{4\beta_0 D}{u^2}} - 1 \right)$  for column dynamics with axial dispersion.

By plotting  $\ln(C/C_0)$  versus t of the experimental data, the resulting curve would be linear, if the Wolborska model is the appropriate for the experimental data. From Figure 9, it can be seen that the relationship is linear for anionic exchange process and, in this case, the  $R^2$  value is 0.9854. For the cationic exchange process (iron and manganese sorption on ZVT-H) the Wolbraska model does not fit properly with the experimental data.



**Figure 9.** Wolborska model linear regression, iron and manganese ions, ZVT-H, cationic exchange.

*Yoon and Nelson Model*

Kinetic adsorption model developed by Yoon and Nelson [28] is based on the assumption that the rate of decrease in the probability of adsorption for each adsorbate molecule is proportional to the probability of adsorbate adsorption and the probability of adsorbate breakthrough, on the adsorbent.

The Yoon and Nelson model has the following differential form:

$$-\frac{dQ}{dt} = \frac{kCU}{m_a} QP \tag{5}$$

where Q is the probability for adsorption, (%),

P is the probability for breakthrough, (%),

t is time, (s).

Experimental evidence also shows that the rate of decrease in the probability of adsorption is directly proportional to the contaminant concentration, C (g/L), and the volumetric flow rate U (L/s), and inversely proportional to the weight of the adsorbent  $m_a$  (g) [28].

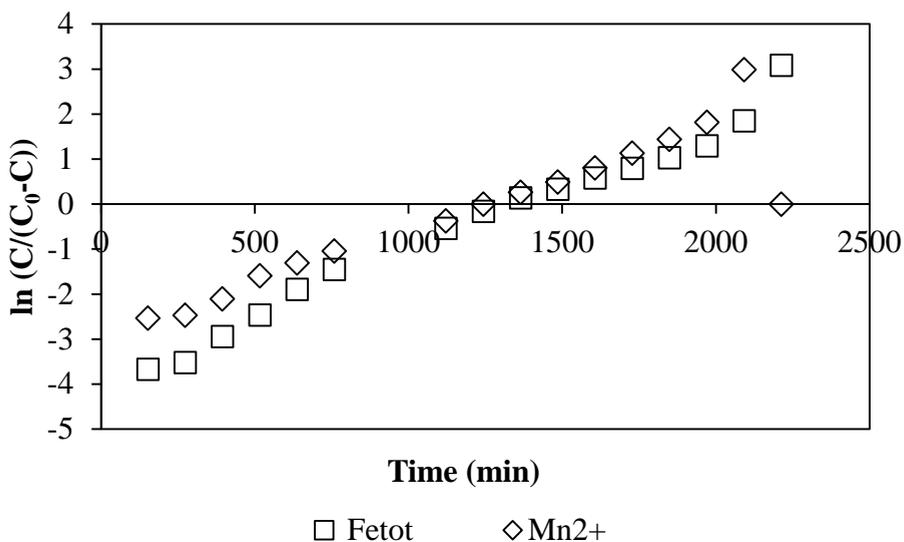
$$\frac{kCU}{m_a} = k_{YN} \tag{6}$$

Solving the above differential equation, using the boundary condition at 50% breakthrough,  $t = t_{0.5}$ ,  $Q = 1/2$  and  $P = 1/2$ , following solution is given:

$$\ln\left(\frac{C}{C_0 - C}\right) = k_{YN}t - t_{0.5}k_{YN} \quad (7)$$

where  $C_0$  is the inlet concentration, mg/L.

In case of Yoon-Nelson, Figure 10 and Table 2, the results obtained suggested that the considered models do not fit properly with the experimental data for iron manganese sorption on ZVT-H and chlorides ions on anionic resin A-21 (small correlation coefficients were obtained).



**Figure 10.** Yoon and Nelson model linear regression, iron and manganese, ZVT-H, cationic exchange.

**Table 2.** Thomas, Wolborska, Yoon and Nelson parameters for  $Fe_{tot}$ ,  $Mn^{2+}$  and  $Cl^-$  sorption at ZVT-H and Amberlite A-21 anionic resin.

Model/ions	Thomas			Wolborska			Yoon-Nelson		
	$q_0$ mg/g	$k_{TH}$ L/min·mg	$R^2$	$h \cdot 10^{-2}$ 1/min	$q_0$ mg/g	$R^2$	$k_{YN} \cdot 10^{-3}$ 1/min	$t_{0.5}$ min	$R^2$
$Fe_{tot}$	0.69	$1.151 \cdot 10^{-2}$	0.977	6.737	1.41	0.930	2.2	29.98	0.848
$Mn^{2+}$	0.074	$1.087 \cdot 10^{-1}$	0.977	9.025	0.129	0.886	2.9	22.60	0.977
$Cl^-$	116.81	$1.88 \cdot 10^{-5}$	0.912	5.567	144.25	0.985	3.5	14.15	0.912

## CONCLUSIONS

Two materials, a natural ionic exchanger (zeolitic volcanic tuff) and a synthetic anionic exchanger were utilized to remove iron and manganese ions, chlorides and to reduce conductivity and COD values from ground water (GW) and surface water (SW) samples, from Ialomița-Muntenia (Romania) catchment.

When the experimental determinations were conducted in static regime, it was found that the equilibrium of the iron and manganese ions removal was reached after 24 hours, utilizing an ion exchange process, on ZVT-H.

In dynamic regime, the values of breakthrough time for manganese ions was almost three times smaller than breakthrough time for sorption of iron ions (10.6 h for manganese ions, compared to 32.82 h, for iron ions).

The maximum sorption capacity values for  $Fe_{tot}$  is higher than the maximum sorption capacity values for  $Mn^{2+}$  (0.543 mg  $Fe_{tot}$ /g ZVT-H compared to 0.044 mg  $Mn^{2+}$ /g ZVT-H).

To describe the fixed bed column behaviour and to scale it up for industrial applications, three models – Thomas, Wolborska and Yoon-Nelson – were used, in order to fit the experimental data with the suitable model.

Good agreement between the predicted theoretical breakthrough curves and the experimental results were observed for Thomas model, in the case of  $Fe_{tot}$  and  $Mn^{2+}$  ions (correlation coefficients of 0.977), and for Wolborska model, in the case of  $Cl^-$  ions (correlation coefficients of 0.985).

## EXPERIMENTAL SECTION

### *Ionic exchanger*

In this study was used zeolitic volcanic tuff (ZVT) from Măcișaș (M), Romania. The sedimentary zeolitic rocks belong to the Dej Tuff Complex of a Lower Badenian age [21]. According to our previous studies of the ZVT from this outcrop, the main zeolite present in these rocks is clinoptilolite, representing 60–70% of the total mineral phases [22- 24]. ZVT samples were first subjected to a physical treatment (grinding, size separation, washing) followed by an acidic treatment [29-30].

Beside the ZVT in H form (ZVT-H) sample and anionic resin Amberlite 21 (A-21)(Sigma-Aldrich) were also used. The synthetic exchanger considered is dimethylamino-resin (weakly basic), with maximum temperature operating of 100°C, moisture content 57%, surface area 25 m<sup>2</sup>/g, porosity 48% and total exchange capacity 1.3 meq/ml, 4.8 meq/g respectively.

*Water sampling and analysis*

For the ionic exchange study it were used drinking water samples from Ialomița-Muntenia (Romania) catchment. Water samples were characterized using the physico-chemical methods according to Romanian legislation (Laws 458/2002, 311/2004). The following water quality parameters were determined: pH, chemical oxygen demand (COD) in mg KMnO<sub>4</sub>/mg/L, electrical conductivity (μS/cm), Mn<sup>2+</sup>, Fe<sub>total</sub>, Cl<sup>-</sup>, NO<sub>2</sub><sup>-</sup>, NH<sub>4</sub><sup>+</sup> (mg/L). pH and electrical conductivity were determined using a Consort C863 pH meter (SR ISO 10523/96, SR ISO 7887/97). Chemical oxygen demand and chlorides were determined by volumetric method (Romanian STAS 7313-82, STAS 3326-76, STAS 3002-85). Concentration of Mn<sup>2+</sup>, Fe<sub>total</sub>, NO<sub>2</sub><sup>-</sup> and NH<sub>4</sub><sup>+</sup> were determined using molecular absorption (SR 8662-1, 2/96, SR 13315/96, STAS 3048/2-96, STAS 6328/85) with a UV VIS Spectrometer Jenway 6305. Each sample was measured three times, the averaged values were presented.

*Ionic exchange experiments*

The cationic exchange process was realised in batch reactor, in static regime using 2 g of the 0.2-0.4 mm particle size of ZVT-H and 100 ml water (exchanger: water =2:100). Solution samples were collected every 24 h for iron, manganese and COD concentrations measurement, until the equilibrium was reached. Then anionic exchange process was conducted under the same conditions (static regime, 2 g of the anionic resin A-21 and 50 ml water), samples were collected for chloride concentration measurement, until the equilibrium was reached.

It also worked in dynamic regime, on a two fixed bed columns (d<sub>i</sub>= 14.5 mm), containing 2 g of the 0.2-0.4, mm grain size ZVT-H, respectively 2 g anionic resin A-21. It was worked with a flow rate of 0.055 mL/s. The effluent was collected at the outflow of the column every 100 mL, until the zeolitic volcanic tuff and anionic resin have been exhausted, in order to determine the exact concentration of iron and manganese ions, respectively chloride concentration and also, their evolution during the ionic exchange process.

Experiments were carried out without any modification of temperature (room temperature).

Exchange capacities, CEC, q in mg/g ZVT-H, mg/g A-21, and removal efficiencies, E (%), were calculated to establish the efficiency of the considered process [31].

$$q = \frac{c_0 - c_e}{m} \cdot \frac{V}{1000} \quad (\text{batch mode}) \quad (8) \quad q_t = \frac{c_0 - c_e}{m} \cdot \frac{V}{1000} + q_{t-1} \quad (9)$$

*(fixed bed)*

$$E(\%) = \frac{C_0 - C_e}{C_0} \cdot 100 \quad (10)$$

where:

$C_0$ ,  $C_e$  are the initial and equilibrium zinc ion concentrations (mg/L);

$q$  is the ionic exchange capacity (mg/g),

$V$  is the volume of water (mL),

$m$  is the mass of ionic exchanger (g).

The kinetics of iron and manganese sorption on ZVT-H respectively chloride sorption on Amberlite A-21, was studied taken into consideration the three models: Thomas, Wolbraska and Yoon-Nelson.

## REFERENCES

1. R.S. Ayers, DW. Westcot DW, "Water quality for agriculture. FAO irrigation and drainage", 29, Rome. ISBN 92-5-102263-1, **1976**.
2. S. Barlow, G. Kozianowski, G. Wurtzen, J. Schlatter, *Chemical Toxicology*, **2001**;39, 893.
3. T. Asano, F.L. Burton, H. Leverenz, R. Tsuchihashi, G. Tchobanoglous, "Water reuse: issues, technologies and applications", New York, McGraw-Hill, **2007**.
4. J. Bartram, T. A. Stenstrom, L. Fewtrell, "Harmonised assessment of risk for water-related microbiological hazards: an overview". In: L. Fewtrell, J. Bartram, (eds) *Water-quality: guidelines, standards and health – risk assessment and management for water-related infectious disease*. IWA, London, for the WHO; **2001**.
5. G. Bitton, R. Harvey, "Transport of pathogens through soils and aquifers". In: R. Mitchell New concepts in environmental microbiology, 19th edn. New York, Wiley, **1992**.
6. U.J. Blumenthal, D.D. Mara, A. Peasey, G. Ruiz-Palacios, R. Stott, *Bulletin of the World Health Organization*, **2000**, 78, 1104.
7. T. Asano, A.D. Levine, *Wastewater reclamation, recycling, and reuse: an introduction*. Asano, T. *Wastewater reclamation and reuse*. Technomic, Lancaster, PA, **1998**.
8. W. Kondro, "E. coli outbreak deaths spark judicial inquiry in Canada", *Lancet* 355, **2000**, 2058.
9. P. Mouchet, *Journal American Water Works Association*, **1992**;84, 158.
10. B. Gaskill, *Water Technology*, **1996**, 19, 10.
11. D. H. Paul, *Water Science Technology*, **1995**, 18(9), 30.
12. G.S. Pellington, *Water Review*, **1992**, 10(1), 1.

13. J. Mc, Peak, *Water Technology*, **1988**, 11(5), 40.
14. M.C. Keller, *Water Technology*, **1995**, 18(9), 35.
15. Y.H. Zhang, *Water Technology*, **1996**, 19(8), 81.
16. Water Quality Association. *Water Treatment Fundamentals*. Lisle, Ill; **1983**.
17. J. Van Newenhizen, *Water Technology*, **1983**, 21(1), 40.
18. T. Asano, D. Richard, R.W. Crites, G. Tchobanoglous, *Water Science Technology*, **1992**, 4, :37.
19. Codex Alimentarius Commission. Proposed draft guidelines for the hygienic reuse of processing water in food plants. Codex Committee on Food Hygiene, FAO, 36th session, Washington D.C., USA, CX/FH/01/9; **2004**.
20. Codex Alimentarius Commission. Procedural manual, 16th edn. ISSN 1020-8070. FAO, Rome; **2006**.
21. Commission of the European Communities. Quality of water intended for human consumption, Council Directive 98/83/EC; **1998**.
22. Law 458/2002, 311/2004, 107/1996, 311/2004, 112/2006, HG 188/2002, HG 352/2005 (in Roumanian).
23. E. Gilca, A. Maicaneanu, P. Ilea, *Central European Journal Chemistry*, **2014**, 12(8), 821.
24. A. Wolborska, *Water Research*, **1989**, 23, 85.
25. H.C. Thomas, *Jornal of the American Chemical Society*, **1944**, 66, 1466.
26. M. Dubinin, K. Nikolaev, N. Poljakov, L. Petrova, "Investigation of adsorption dynamics in a broad range of breakthrough concentrations". Institute of Physical Chemistry, Academy of Sciences of the USSR. Translated from *Izvestiya Akademii Nauk SSSR, Seriya Khimicheskaya*. **1972**, 2, 1265.
27. M. Dubinin, K. Nikolaev, N. Poljakov, G. Pirozhkov, "Pore structures of charcoal adsorbents and the adsorbability of gases and vapors". Institute of Physical Chemistry, Academy of Sciences of the USSR, Moscow. Translated from *Izvestiya Akademii Nauk SSSR, Seriya Khimicheskaya*, **1980**;8, 1728.
28. Y.H. Yoon, J.H. Nelson, *American Industrial Hygiene Association Journal*, **1984**, 45, 509.
29. H. Bedeleian, M. Stanca, A. Maicaneanu, S. Burca, *Studies and Research, Geology-Geography*, **2005**, 10(53), 881.
30. M Stanca, A. Maicaneanu, S. Burca, H. Bedeleian, *Studia Universitatis Babeş-Bolyai Chemia*, **2006**, LI(2), 187.
31. A. Maicaneanu, H. Bedeleian, M. Stanca, "Zeolitii naturali. Caracterizare si aplicatii in protectia mediului", Cluj University Press, Cluj-Napoca, Romania; **2008**.